Novel Evolutionary Engineering Approach for Accelerated Utilization of Glucose, Xylose, and Arabinose Mixtures by Engineered *Saccharomyces cerevisiae* Strains\(^7\)

H. Wouter Wisselink,\(^1,2\) Maurice J. Toirkens,\(^1,2\) Qixiang Wu,\(^1,2\) Jack T. Pronk,\(^1,2\) and Antonius J. A. van Maris\(^1,2,8\)

*Department of Biotechnology, Delft University of Technology, Julianalaan 67, 2628 BC Delft, The Netherlands,\(^1\) and Kluyver Centre for Genomics of Industrial Fermentation, P.O. Box 5057, 2600 GA Delft, The Netherlands\(^2\)

Received 2 October 2008/Accepted 5 December 2008

Lignocellulosic feedstocks are thought to have great economic and environmental significance for future biotechnological production processes. For cost-effective and efficient industrial processes, complete and fast conversion of all sugars derived from these feedstocks is required. Hence, simultaneous or fast sequential fermentation of sugars would greatly contribute to the efficiency of production processes. One of the main challenges emerging from the use of lignocellulosics for the production of ethanol by the yeast *Saccharomyces cerevisiae* is efficient fermentation of D-xylose and L-arabinose, as these sugars cannot be used by natural *S. cerevisiae* strains. In this study, we describe the first engineered *S. cerevisiae* strain (strain IMS0003) capable of fermenting mixtures of glucose, xylose, and arabinose with a high ethanol yield (0.43 g g\(^{-1}\) of total sugar) without formation of the side products xylitol and arabinitol. The kinetics of anaerobic fermentation of glucose-xylose-arabinose mixtures were greatly improved by using a novel evolutionary engineering strategy. This strategy included a regimen consisting of repeated batch cultivation with repeated cycles of consecutive growth in three media with different compositions (glucose, xylose, and arabinose; xylose and arabinose; and only arabinose) and allowed rapid selection of an evolved strain (IMS0010) exhibiting improved specific rates of consumption of xylose and arabinose. This evolution strategy resulted in a 40% reduction in the time required to completely ferment a mixture containing 30 g liter\(^{-1}\) glucose, 15 g liter\(^{-1}\) xylose, and 15 g liter\(^{-1}\) arabinose.

\(^{1}\) Corresponding author. Mailing address: Julianalaan 67, 2628 BC Delft, The Netherlands. Phone: 31 15 278 1616. Fax: 31 15 278 2355. E-mail: A.J.A.vanMaris@TUDelft.nl.

\(^{2}\) Published ahead of print on 12 December 2008.
The goal of the present study was to evaluate and optimize selection strategies for evolutionary optimization of the utilization of substrate mixtures. Fermentation of glucose, xylose, and arabinose mixtures by engineered strains was used as the model.

### MATERIALS AND METHODS

**Strains and maintenance**. The *S. cerevisiae* strains used in this study are listed in Table 1. Culture samples from either shake flasks, chemostats, or (sequential) batch cultivation were prepared by addition of 30% (vol/vol) glycerol and were stored as 2-ml aliquots at −80°C.

**Media and shake flask cultivation**. Shake flask cultivation was performed at 30°C in synthetic medium (MY) containing 5 g liter⁻¹ (NH₄)₂SO₄, 3 g liter⁻¹ KH₂PO₄, 0.5 g liter⁻¹ MgSO₄, 7H₂O, 0.05 ml liter⁻¹ silicon antifoam, and trace elements (23). For cultivation in shake flasks, the pH of the medium was adjusted to 6.0 with 2 M KOH prior to sterilization. After heat sterilization (121°C, 20 min), a filter-sterilized vitamin solution (23) and an appropriate carbon and energy source were added. Shake flask cultures were prepared by inoculating 100 ml medium containing the appropriate sugar with 500-ml shake flask with a frozen stock culture and were incubated at 30°C in an orbital shaker (200 rpm).

Solid MY plates containing 20 g liter⁻¹ xylose (MY-X) or 20 g liter⁻¹ arabinose (MY-A) were prepared by adding 1.5% agar to MY. Plates were incubated at 30°C until colonies appeared. Separate colonies were reseeded twice on solid MY-A. Single colonies were cultivated at 30°C in shake flasks containing 100 ml MY-A. Frozen stock cultures were prepared by addition of sterile 30% (vol/vol) glycerol in the stationary growth phase and storage of 2-ml aliquots at −80°C.

**Preparation of single-colony isolate cultures**. Culture samples, either from the chemostat or from sequential batch cultures, were diluted and spread on solid MY-A and incubated at 30°C until colonies appeared. Separate colonies were reseeded twice on solid MY-A. Single colonies were cultivated at 30°C in shake flasks containing 100 ml MY-A. Frozen stock cultures were prepared by addition of sterile 30% (vol/vol) glycerol in the stationary growth phase and storage of 2-ml aliquots at −80°C.

**Preparation of single-colony isolate cultures**. Culture samples, either from the chemostat or from sequential batch cultures, were diluted and spread on solid MY-A and incubated at 30°C until colonies appeared. Separate colonies were reseeded twice on solid MY-A. Single colonies were cultivated at 30°C in shake flasks containing 100 ml MY-A. Frozen stock cultures were prepared by addition of sterile 30% (vol/vol) glycerol in the stationary growth phase and storage of 2-ml aliquots at −80°C.

### Table 1. *S. cerevisiae* strains used in this study

<table>
<thead>
<tr>
<th>Strain</th>
<th>Characteristics</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>IMS0002</td>
<td>MATα ara1-52 lou2-112, loxP-PTPI: (−266,−1)TAL1 gre3::hphMX pUGPTPI-TKLI pUGPTPI-RPE1 loxP-PTPI: (−49,−1)RK1 (pRW231, pRW243); strain harboring Pirromyces sp. strain E2 Gly4 and Lactobacillus plantarum araA and araD on 2µg-based plasmid pRW231 and XKS1 and L. plantarum araB on integration plasmid pRW243; selected for anaerobic growth on l-arabinose</td>
<td>26</td>
</tr>
<tr>
<td>IMS0003</td>
<td>Single-colony isolate of strain IMS0002 cultivated anaerobically on solid MY-X; capable of cofermenting mixtures of glucose, xylose, and arabinose to ethanol</td>
<td>This study</td>
</tr>
<tr>
<td>IMS0007</td>
<td>Single-colony isolate obtained after long-term chemostat cultivation in MY-XA</td>
<td>This study</td>
</tr>
<tr>
<td>IMS0010</td>
<td>Single-colony isolate obtained after repeated consecutive batch cultivation in MY supplemented with a mixture of glucose, xylose, and arabinose as the carbon and energy sources, in MY supplemented with a mixture of xylose and arabinose as the carbon and energy sources, and in MY supplemented with arabinose as the sole carbon and energy source</td>
<td>This study</td>
</tr>
</tbody>
</table>

### Chemostat cultivation.** Anaerobic chemostat cultivation was carried out at 30°C in 2-liter laboratory fermentors (Applikon, Schiedam, The Netherlands) with a working volume of 1 liter. Cultures were grown in MY supplemented with 0.01 g liter⁻¹ ergosterol, 0.42 g liter⁻¹ Tween 80 dissolved in ethanol (1, 2), silicon antifoam, trace elements (23), and the appropriate carbon and energy source and were maintained at pH 5.0 by automatic addition of 2 M KOH. Cultures were stirred at 800 rpm and sparged with 0.5 liters min⁻¹ nitrogen gas (<10 ppm oxygen). To minimize diffusion of oxygen, the fermentors were equipped with Norpette tubing (Cole Palmer Instrument Company, Vernon Hills, IL). In addition, the presence of oxygen was monitored with an oxygen electrode (Applisens, Schiedam, The Netherlands). After inoculation and completion of the batch phase, chemostat cultivation was initiated by addition of MY containing 20 g liter⁻¹ xylose and 20 g liter⁻¹ arabinose (MY-XA) to the fermentor at a fixed dilution rate. The working volume of the culture was kept constant using an effluent pump controlled by an electric level sensor.

### Sequential batch cultivation.** For anaerobic sequential batch cultivation (in sequential batch reactors [SBR]) the fermentor setup and MY composition that were used for chemostat cultivation were used. Each fermentor was filled to obtain a working volume of 1 liter using a feed pump controlled by an electric level sensor. When the carbon and energy source was depleted, as indicated by a CO₂ level in the exhaust gas that was less than 0.05% after the CO₂ production peak, a new cycle of batch cultivation was initiated by either manual or automated replacement of approximately 90% of the culture with fresh synthetic medium containing the appropriate carbon and energy source. For each cycle, the maximum specific growth rate (μmax) was estimated from the CO₂ profile in the exponential growth phase.

**Preparation of single-colony isolate cultures.** Culture samples, either from the chemostat or from sequential batch cultures, were diluted and spread on solid MY-A and incubated at 30°C until colonies appeared. Separate colonies were reseeded twice on solid MY-A. Single colonies were cultivated at 30°C in shake flasks containing 100 ml MY-A. Frozen stock cultures were prepared by addition of sterile 30% (vol/vol) glycerol in the stationary growth phase and storage of 2-ml aliquots at −80°C.

**Rate calculation.** For calculation of the specific rates of glucose, xylose, and xylitol consumption, the time-dependent sugar concentrations [S(t)] were fitted to be equal to the measured cumulative production of CO₂ minus the CO₂ production that occurred due to biomass synthesis (5.85 mmol CO₂ per g biomass [22]) and acetate formation.
where $A_i$ is the initial value (left horizontal asymptote), $A_e$ is the final value (right horizontal asymptote), $t_0$ is the center (point of inflection), and $B \cdot t - C$ is the time-dependent width $r$, which was defined as the change in $t$ corresponding to the most significant change in $y$.

For each time point, the specific sugar consumption rates were calculated by dividing the derivative of the fitted curves by the dry weight.

RESULTS

Cofermentation of glucose, xylose, and arabinose by *S. cerevisiae* IMS0003. In a previous study, we described construction and evolutionary optimization of *S. cerevisiae* strain IMS0002, which is capable of fermenting arabinose to ethanol under anaerobic conditions (26). Although this strain was engineered for both xylose and arabinose utilization, the ability to ferment xylose to ethanol was lost during the long-term selection for anaerobic l-arabinose fermentation. The ability to ferment xylose was recovered by anaerobic cultivation of single colonies of strain IMS0002 on solid MY-X at 30°C. Small colonies appeared after 10 days, and after 30 days of incubation, colonies were transferred to solid MY-A, cultivated in a shake flask containing MY-A, and designated strain IMS0003. To verify that there was anaerobic utilization of both xylose and arabinose, strain IMS0003 was cultivated anaerobically in MY supplemented with a mixture containing 30 g liter$^{-1}$ glucose, 15 g liter$^{-1}$ D-xylose, and 15 g liter$^{-1}$ L-arabinose. All sugars present were consumed within approximately 60 h, as shown by the sugar consumption profiles (Fig. 1A). The xylose and arabinose profiles had a long “tail,” indicating that further improvement of the affinity for these sugars is possible.

Minor improvement of the fermentation kinetics by prolonged chemostat cultivation on xylose-arabinose mixtures. To select for spontaneous mutants with an improved affinity for xylose and arabinose, strain IMS0003 was cultivated in an anaerobic chemostat containing MY-XA at a dilution rate of 0.03 h$^{-1}$. For the first 6 days, the steady-state xylose and arabinose concentrations were stable at 10.4 and 4.0 g liter$^{-1}$, respectively (Fig. 2). After this, a decrease in the residual xylose concentration from 10 g liter$^{-1}$ to approximately 1.3 g liter$^{-1}$ was observed, while the dry weight of cells increased from 1.9 to 2.5 g liter$^{-1}$. This resulted in an increase in the specific xylose consumption rate from 0.15 to 0.21 g g$^{-1}$ (dry weight)$^{-1}$ h$^{-1}$. The specific arabinose consumption rate, however, decreased from 0.24 to 0.19 g g$^{-1}$ (dry weight)$^{-1}$ h$^{-1}$ as a result of only a minor decrease in the residual arabinose concentration and the increased dry weight of cells. As expected for chemostat cultivation without drastic shifts in the biomass yield, the sum of the specific consumption rates of the two pentoses remained constant at approximately 0.4 g g$^{-1}$ (dry weight)$^{-1}$ h$^{-1}$ throughout the first 21 days of cultivation.

After 21 days, the dilution rate was increased to 0.05 h$^{-1}$. After the initial increase in the residual arabinose concentration due to the increased dilution rate, the only significant change during this phase was a decrease in the residual arabinose concentration. After approximately 40 days of selective chemostat cultivation, a sample was removed, and multiple single colonies were isolated. The behavior of cultures resulting from evolutionary engineering can be due to either one dominant strain or a mixture of strains with different phenotypes. To exclude the latter possibility, where none or very few of the individual strains had the phenotype of the combined culture, two randomly selected single-colony isolates were compared to the heterogeneous sample from the chemostat by recording the CO$_2$ production profile during anaerobic batch cultivation in MY supplemented with 30 g liter$^{-1}$ glucose, 15 g liter$^{-1}$ D-xylose, and 15 g liter$^{-1}$ L-arabinose. Since these two single-colony isolates appeared to be representative of the phenotype of the heterogeneous sample, no additional isolates were characterized. The sugar consumption profile of single-colony isolate IMS0007 (Fig. 1B) showed that the total time necessary to consume the three sugars was only approximately 5 h less than the time necessary for strain IMS0003 to consume the three sugars (Fig. 1A).

Fermentation kinetics deteriorated after selective repeated batch cultivation with a xylose-arabinose mixture. As described above, prolonged cultivation in the chemostat resulted in only a minor improvement in the fermentation performance in medium containing a mixture of glucose, xylose, and arabinose (Fig. 1). To obtain an *S. cerevisiae* strain with either improved xylose and arabinose coconsumption or faster sequential consumption of both pentose sugars, further selection was performed using automated SBRs. To do this, anaerobic SBR cultivation in MY-XA was performed. The selective SBR cultivation was started by inoculation with a sample from the previous chemostat selection, after which the repeated anaerobic batch cultivation regimen was performed by automated emptying and filling of the fermentor with MY-XA at the end of each batch after both pentoses were consumed. When the CO$_2$ production profiles of the individual sequential batches were compared, a clear increase in the maximum CO$_2$ production rate and a progressively faster increase in CO$_2$ production were observed (Fig. 3A), which were confirmed by the increase in the $\mu_{max}$ from 0.08 to 0.13 h$^{-1}$ (Fig. 3B). Interestingly, the increase in the $\mu_{max}$ was accompanied by a gradual shift from a single CO$_2$ production peak in the first cycle of the SBR to a typical diauxic CO$_2$ production profile in cycle 16 (Fig. 3A). Analyses of sugars in supernatant samples from the SBR cultivation showed that the increase in the $\mu_{max}$ was a result of increasingly higher xylose consumption rates. The arabinose consumption rates, however, decreased during SBR selection. Clearly, the consumption phases for xylose and arabinose were separated more over time, which eventually resulted in an increase in the total time necessary to completely consume xylose and arabinose from 55 h to more than 70 h (Fig. 3A). The deteriorating arabinose consumption was confirmed by anaerobic batch cultivation of a 100-ml sample from the SBR culture (batch 13) in MY supplemented with 30 g liter$^{-1}$ glucose, 15 g liter$^{-1}$ D-xylose, and 15 g liter$^{-1}$ L-arabinose. Sugar analysis of supernatant samples revealed that the arabinose consumption phase was delayed and longer than that of chemostat isolate IMS0007, which was confirmed by the long “tail” of the CO$_2$ production profile (Fig. 3C).

Substantially improved fermentation kinetics due to repeated consecutive anaerobic batch cultivation in media with alternating glucose, xylose, and arabinose compositions. Repeated batch cultivation in medium with a fixed sugar compo-
position (MY-XA) resulted in a shift from coconsumption of xylose and arabinose to more sequential utilization of the two sugars. One explanation of this is that the starting culture started consuming xylose before it started consuming arabinose. As a consequence, the number of generations of growth on xylose was higher than the number of generations of growth on arabinose, resulting in an undesired increase in the selection pressure for faster growth on xylose alone and a reduction in the selection pressure for improved growth on arabinose. To select for cells that exhibited improved rates of consumption of

FIG. 1. Product formation and sugar consumption (A, B, and C) and specific consumption rates (D, E, and F) during anaerobic batch cultivation of strains IMS0003 (A and D), IMS0007 (B and E), and IMS0010 (C and F) in MY containing a mixture of 30 g liter⁻¹ glucose, 15 g liter⁻¹ D-xylose, and 15 g liter⁻¹ L-arabinose. The data are data from single batch cultivations and are representative of duplicate experiments. To correct for small differences in the initial biomass, the profiles were aligned using the beginning of the glucose consumption peak. Ethanol concentrations were derived from the cumulative CO₂ production. (A, B, and C) Symbols: ▼, concentration of glucose; ○, concentration of xylose; □, concentration of arabinose; ●, concentration of ethanol; △, dry weight of cells. (D, E, and F) Symbols: ●, specific consumption rate for xylose; ○, specific consumption rate for arabinose; ■, sum of the specific consumption rates for the two pentoses. DW, dry weight of cells.
both xylose and arabinose, the selection pressure for utilization of arabinose was enhanced by increasing the number of generations of cells growing on arabinose without compromising the selection pressure for xylose. To accomplish this, a new selective anaerobic SBR cultivation strategy was designed, consisting of three consecutive phases of cultivation in (i) MY containing 20 g liter\(^{-1}\) glucose, 20 g liter\(^{-1}\) xylose, and 20 g liter\(^{-1}\) arabinose (MY-GXA), (ii) MY-XA, and (iii) MY-A.

The new SBR selection regimen was started by inoculation with a (whole-broth) sample taken from the selective chemostat cultivation. Consecutive cultivation in MY-GXA, MY-XA, and MY-A resulted in typical CO\(_2\) production profiles, as shown by the profiles for cycle 1 in Fig. 4A. During cultivation in MY-GXA, glucose was consumed first, followed by xylose and arabinose, and this resulted in a typical diauxic CO\(_2\) production pattern. Xylose and arabinose were consumed simultaneously during batch cultivation in MY-XA, which was illustrated by the single CO\(_2\) production peak. When the complete three-phase cycle was repeated 20 times, progressively increasing CO\(_2\) production rates were observed for the three medium compositions, which was shown by the gradual increase in the \(\mu_{\text{max}}\). Within 20 cycles of batch cultivation, the \(\mu_{\text{max}}\) (as determined by CO\(_2\) production) increased from 0.19 h\(^{-1}\) to approximately 0.23 h\(^{-1}\) in MY-GXA and from 0.08 to 0.12 h\(^{-1}\) in MY-XA (Fig. 4B). Only a minor increase in the growth rate was observed for batch cultivation with arabinose as the sole carbon source. In addition to the increased \(\mu_{\text{max}}\) values, the total time necessary to consume all sugars present in the medium was substantially reduced from 50 h to approximately 35 h for MY-GXA and from 50 to 30 h for MY-XA (Fig. 4A). Furthermore, the single CO\(_2\) production peak for the batch cultivation in MY-XA indicates that the ability to utilize xylose and arabinose simultaneously was preserved during the repeated batch cultivation; this is in contrast to the findings obtained previously with SBR using medium with a fixed sugar composition. In addition to improved cococonsumption of xylose and arabinose, the batch cultivation times in medium with only arabinose were shortened from approximately 55 to 40 h.

After 20 cycles of repeated batch cultivation, a sample was removed, and single colonies were isolated by plating on solid MY-A. Two single-colony isolates were characterized to determine their CO\(_2\) production profiles with a mixture containing 30 g liter\(^{-1}\) glucose, 15 g liter\(^{-1}\) D-xylose, and 15 g liter\(^{-1}\) L-arabinose, and they were found to be representative of the sample obtained directly from the SBR. One of the single-cell isolates, designated strain IMS0010, was used for further characterization. Comparison of the sugar consumption and ethanol production profiles of strains IMS0007 and IMS0010 (Fig. 1B and 1C) revealed that the total time necessary to ferment the glucose-xylose-arabinose mixture was reduced from 55 h to approximately 35 h.
Fermentation characteristics and kinetics of strains selected for improved mixed-sugar fermentation. To compare physiological characteristics of the *S. cerevisiae* strains obtained by the different selection methods described in this paper, anaerobic batch cultivation of strains IMS0003, IMS0007, and IMS0010 in mixtures containing 30 g liter\(^{-1}\) glucose, 15 g liter\(^{-1}\) xylose, and 15 g liter\(^{-1}\) arabinose was performed (Fig. 1). Strain IMS0003 was able to consume the sugar mixture within approximately 65 h (Fig. 1A), and the maximum specific consumption rates (\(q_{\text{max}}\)) were 0.27 and 0.23 g g\(^{-1}\) (dry weight)\(^{-1}\) h\(^{-1}\) for xylose and arabinose, respectively (Fig. 1D and Table 2). As expected from the marginal reduction in total fermentation time after chemostat selection, no drastic changes in sugar consumption occurred when IMS0003 and IMS0007 were compared. While the \(q_{\text{max}}\) for xylose increased from 0.21 to 0.31 g g\(^{-1}\) (dry weight)\(^{-1}\) h\(^{-1}\), the \(q_{\text{max}}\) for arabinose did not change significantly (Table 2). The slightly improved coconsumption of both pentoses observed for IMS0007 is illustrated by the increased sum of the two pentose-specific consumption rates (Fig. 1E) compared to that of IMS0003.

The strain resulting from repeated consecutive batch cultivation with sugar mixtures having alternating compositions (IMS00010) exhibited clearly improved fermentation performance. While the \(q_{\text{max}}\) for xylose, 0.35 g g\(^{-1}\) (dry weight)\(^{-1}\) h\(^{-1}\), was comparable to that of strain IMS00007, arabinose was consumed with a \(q_{\text{max}}\) of 0.53 g g\(^{-1}\) (dry weight)\(^{-1}\) h\(^{-1}\), which corresponded to a twofold increase compared to strain IMS00007 (Fig. 1F and Table 2). As a result, the sum of the specific consumption rates of the two pentoses increased 1.5-fold (Fig. 1F), which contributed to a great extent to the reduction in the total fermentation time from 55 h to approximately 35 h (Fig. 1C).

All three xylose- and arabinose-consuming strains described in this study exhibited high ethanol yields (0.43 to 0.44 g g\(^{-1}\) of total sugar) (Table 2). In line with these yields, the levels of xylitol and arabinitol remained below the detection level. This indicated that the ethanol yields were not (negatively) affected either by the chemostat selection or the selective sequential batch cultivation. The biomass yield of strain IMS00010, however, was significantly higher than that of strains IMS00003 and IMS00007. The increased biomass yield of strain IMS00010 can be explained as an effect of a decreased cumulative energy requirement for maintenance as a result of the decreased fermentation times.

**DISCUSSION**

The use of evolutionary engineering has proven to be a very successful tool for selecting for recombinant *S. cerevisiae* strains capable of anaerobically utilizing sugars such as xylose and arabinose (5, 13, 19, 26). Natural selection even enabled a non-metabolically engineered *S. cerevisiae* strain to utilize xylose as a sole carbon and energy source, which demonstrates that trace activities of enzymes in the targeted metabolic route can result in growth, provided that strong selection pressure is applied (4). Hitherto, research on evolutionary engineering of pentose utilization by *S. cerevisiae* has focused primarily on the use of single sugars, and there have been only a few examples of improved utilization of multiple sugars (12). Improvement of the utilization of multiple substrates has specific challenges, as it is rather complicated to select for multiple mutations in different metabolic routes that require different kinds of (potentially conflicting) selective pressure (17). In line with this observation, previous evolutionary engineering for anaerobic utilization of arabinose as the sole carbon source resulted in the loss of the xylose-utilizing capacities of an engineered *S. cerevisiae* strain (26).

Given the complexity of biomass hydrolysates, evolutionary engineering strategies that enable efficient utilization of mixtures containing three (or more) different sugars are required. The sequencing batch evolution strategy with 20 g liter\(^{-1}\) xylose and 20 g liter\(^{-1}\) arabinose (Fig. 3) clearly demonstrated that a slight preference for xylose over arabinose resulted in increasingly more generations on xylose and thus a shift of the selection pressure to the preferred sugar. Although, as a result, growth on xylose improved dramatically, the deteriorated kinetics of arabinose utilization resulted in overall less favorable fermentation characteristics. If the expression of a native or heterologous pathway for a less-preferred substrate confers even a slight selective disadvantage (for example, via protein burden [18]), repeated cultivation on a fixed substrate mixture may ultimately select for strains that exhibit the well-known phenomenon of diauxic growth (15). To minimize such unequal selection pressure on the utilization of glucose, xylose, and arabinose, a novel selection strategy involving consecutive anaerobic batch cultivation in MY-GXA, MY-XA, and MY-A was used. This selection strategy allows more even distribution of the number of generations grown on each carbon source,
The amounts ethanol were derived from the cumulative CO2 production. The values are the averages.

<table>
<thead>
<tr>
<th>Strain</th>
<th>Biomass</th>
<th>CO2</th>
<th>Ethanol</th>
<th>Glycerol</th>
<th>Acetate</th>
<th>Lactate</th>
<th>Pyruvate</th>
<th>Succinate</th>
<th>Xylose</th>
<th>Arabinose</th>
</tr>
</thead>
<tbody>
<tr>
<td>IMS0001</td>
<td>0.000 ± 0.001</td>
<td>0.44 ± 0.01</td>
<td>1.11 ± 0.01</td>
<td>0.007 ± 0.002</td>
<td>0.41 ± 0.01</td>
<td>0.001 ± 0.001</td>
<td>0.000 ± 0.001</td>
<td>0.006 ± 0.000</td>
<td>0.006 ± 0.000</td>
<td>0.006 ± 0.000</td>
</tr>
<tr>
<td>IMS0003</td>
<td>0.005 ± 0.001</td>
<td>0.44 ± 0.01</td>
<td>1.07 ± 0.01</td>
<td>0.007 ± 0.002</td>
<td>0.42 ± 0.01</td>
<td>0.001 ± 0.001</td>
<td>0.000 ± 0.001</td>
<td>0.006 ± 0.000</td>
<td>0.006 ± 0.000</td>
<td>0.006 ± 0.000</td>
</tr>
<tr>
<td>IMS0007</td>
<td>0.008 ± 0.002</td>
<td>0.43 ± 0.01</td>
<td>1.05 ± 0.01</td>
<td>0.008 ± 0.002</td>
<td>0.43 ± 0.01</td>
<td>0.001 ± 0.001</td>
<td>0.000 ± 0.001</td>
<td>0.007 ± 0.000</td>
<td>0.007 ± 0.000</td>
<td>0.007 ± 0.000</td>
</tr>
<tr>
<td>IMS0010</td>
<td>0.009 ± 0.001</td>
<td>0.44 ± 0.01</td>
<td>1.05 ± 0.01</td>
<td>0.007 ± 0.002</td>
<td>0.43 ± 0.01</td>
<td>0.001 ± 0.001</td>
<td>0.000 ± 0.001</td>
<td>0.007 ± 0.000</td>
<td>0.007 ± 0.000</td>
<td>0.007 ± 0.000</td>
</tr>
</tbody>
</table>

Strain dry wt (g g \(^{-1}\) of total sugar)

<table>
<thead>
<tr>
<th>Strain</th>
<th>Observed qmax (g g (^{-1}) of total sugar)</th>
</tr>
</thead>
<tbody>
<tr>
<td>IMS0001</td>
<td>0.21 ± 0.03</td>
</tr>
<tr>
<td>IMS0003</td>
<td>0.21 ± 0.03</td>
</tr>
<tr>
<td>IMS0007</td>
<td>0.21 ± 0.03</td>
</tr>
<tr>
<td>IMS0010</td>
<td>0.21 ± 0.03</td>
</tr>
</tbody>
</table>

**TABLE 2.** Products yields, levels of carbon recovery, and specific consumption rates determined for anaerobic batch cultivation of strains IMS0003, IMS0007, and IMS0010 in MY supplemented with 30 g liter \(^{-1}\) glucose, 15 g liter \(^{-1}\) xylose, and 15 g liter \(^{-1}\) arabinose.

The regimen involving repetitive consecutive cultivation with the three different sugar mixtures yielded an *S. cerevisiae* strain (strain IMS0010) that exhibited rapid anaerobic fermentation of mixtures of glucose, xylose, and arabinose to ethanol. The underlying genetic changes involved in the improvement in xylose and arabinose utilization by the evolved strain IMS0010 remain to be investigated. The possible changes include mutations that resulted in improved codon usage of introduced genes, as codon optimization for *AraA*, *AraB*, and *AraD* in engineered *S. cerevisiae* strains has been shown to result in improved arabinose conversion rates (25). Alternatively, changes in plasmid copy numbers may have played a role in fine-tuning the levels of expression of introduced genes, as described previously for *S. cerevisiae* strains evolved for lactose utilization (8). Interestingly, the specific combined pentose consumption rate of IMS0010 nearly equals the maximum xylose consumption rate of RWB218 with glucose-xylose mixtures (12), which might indicate that there is a common flux-controlling step during pentose utilization by these strains.

In plant biomass hydrolysates, xylose, and especially arabinose comprise only a small fraction of the total carbon. The use of such hydrolysates for evolutionary engineering would limit the selection pressure for the substrates that are less preferred by the organism of choice and therefore might not result in improved kinetics for utilization of the minor components of the medium. A simple solution would be to add these substrates to the hydrolysates in various stages of the evolutionary engineering process to obtain enough generations on the desired substrates. In this way, rapid consumption of the less predominant substrates, which often are crucial for the overall process economics, in the production environment may be achieved. Equal selection pressures for multiple sugars can be achieved in multiple ways, and the fermentation setup described above is not unique. For example, repeated cultivation with a (fixed) mixture of 5 g liter \(^{-1}\) glucose, 15 g liter \(^{-1}\) xylose, and 45 g liter \(^{-1}\) arabinose would also result in a more even distribution of the selection pressures. However, although the number of generations is expected to be similar, the simultaneous presence of various sugars at high (repressing) concentrations in the strategy described in this paper add selective pressure for rapid subsequent or even partially simultaneous use of the substrates.

To our knowledge, the strategy described here for improving the utilization of mixtures of three or more substrates via consecutive batch cultivation in media with alternating sugar compositions has not been described previously. Moreover, although the strategy has to be tested in practice, its applications do not seem to be limited to the selection of improved *S. cerevisiae* phenotypes for ethanol production; this strategy might also be applicable to other microorganisms used in (industrial) biotechnological processes based on the conversion of lignocellulosic hydrolysates or other substrate mixtures.

**ACKNOWLEDGMENTS**

This project was financially supported by The Netherlands Ministry of Economic Affairs and the B-Basic partner organizations (www.b-basic.nl) through B-Basic, a public-private NWO-Advanced Chemical Technologies for Sustainability program. The Kluyver Centre for
REFERENCES


