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Microbially mediated kinetic sulfur isotope fractionation: reactive transport modeling benchmark

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Abstract

Microbially mediated sulfate reduction is a ubiquitous process in many subsurface systems. Isotopic fractionation is characteristic of this anaerobic process, since sulfate-reducing bacteria (SRB) favor the reduction of the lighter sulfate isotopologue ($\text{S}^{32}\text{O}_4^{2-}$) over the heavier isotopologue ($\text{S}^{34}\text{O}_4^{2-}$). Detection of isotopic shifts has been utilized as a proxy for the onset of sulfate reduction in subsurface systems such as oil reservoirs and aquifers undergoing heavy metal and radionuclide bioremediation. Reactive transport modeling (RTM) of kinetic sulfur isotope fractionation has been applied to field and laboratory studies. We developed a benchmark problem set for the simulation of kinetic sulfur isotope fractionation during microbially mediated sulfate reduction. The benchmark problem set is comprised of three problem levels and is based on a large-scale laboratory column experimental study of organic carbon amended sulfate reduction in soils from a uranium-contaminated aquifer. Pertinent processes impacting sulfur isotopic composition such as microbial sulfate reduction and iron-sulfide reactions are included in the problem set. This benchmark also explores the different mathematical formulations in the representation of kinetic sulfur isotope fractionation as employed in the different RTMs. Participating RTM codes are the following: CrunchTope, TOUGHREACT, PHREEQC, and PHT3D. Across all problem levels, simulation results from all RTMs demonstrate reasonable agreement.

Keywords Reactive transport modeling · Benchmark · Microbial sulfate reduction · S isotopes · Kinetic isotope fractionation

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1 Introduction

Dissimilatory sulfate reduction (DSR) occurs when sulfate-reducing microorganisms (SRM) oxidize organic matter anaerobically using sulfate as electron acceptor. Isotopic fractionation is typically characteristic of this anaerobic process, since sulfate-reducing bacteria (SRB) favor the reduction of the lighter sulfate isotopologue ($^{32}\text{SO}_4^{2-}$) over the heavier isotopologue ($^{34}\text{SO}_4^{2-}$) under the majority of environmental conditions. DSR occurs ubiquitously and plays an important role in sulfur cycling in natural terrestrial and marine sediments, causing wide range of $\delta^{34}\text{S}$ signatures [1–3]. Fractionations in the $^{34}\text{S}/^{32}\text{S}$ ratio have been extensively reported in marine, terrestrial, and subsurface environments and can vary widely. Sim et al. [3] reported values that range from < 10 to > 60‰. Recent pure culture experiments have demonstrated fractionation as large as 66‰ [3].

DSR is a key process in many subsurface applications such as remediation of uranium and acid mines. SRB populations are commonly limited by labile organic carbon substrate availability and are thus stimulated through the injection of organic carbon compounds in order to increase DSR rates, leading to

high rates of sulfate removal [4]. Organic carbon biostimulation experiments conducted in aquifers undergoing heavy metal and radionuclide remediation have pointed to a link between the onset of DSR and enhanced long-term stability of reduced uranium precipitates [5]. In offshore oil recovery operations, seawater is typically injected into the production reservoir to maintain pressure. Interaction of this sulfate-rich seawater with the organic carbon of the reservoir matrix creates favorable conditions for DSR, producing hydrogen sulfide, H_2S . The toxic nature of this H_2S poses health risks [6] to workers on site and compromises the integrity of metallic structure and pipeline equipment. Despite this range of environmental, infrastructure, and health effects, detection of DSR remains challenging, due to fluctuations in onsite background sulfate concentrations [7]. Further, effluent sulfide concentrations can react with iron minerals [8–10], thus masking their appearance in fluid samples. Breakthrough curves of sulfate $\delta^{34}S$ have therefore been utilized as a more reliable and accurate proxy for the onset of DSR in subsurface systems.

In subsurface systems characterized by both chemical and physical complexity, the usage of traditional Rayleigh-type equations to model biological kinetic isotope fractionation has been shown to be inadequate [10–12]. Such simple distillation assumes that reactions are unidirectional, irreversible, and the only source of change to reactant concentrations [13], while reactions in soil systems are by and large, mediated by a broad range of factors including microorganisms via enzymatic reactions that follow Michaelis-Menten kinetics. Recent studies showed that the Rayleigh equation systematically underestimated the rate of biodegradation in subsurface systems undergoing enhanced remediation of organic pollutants [11, 14, 15]. Thus, reactive transport models (RTMs) are often necessary to quantitatively describe isotopic effects such as kinetic sulfur isotope fractionation. These simulations have been applied to both field and laboratory studies using a variety of mathematical formulations [10, 16–20].

Despite these studies and the availability of a large number of codes that essentially solve the same governing equations, benchmarking exercises are useful to document conceptual and numerical capabilities of models and identify needs for further improvement [21]. It is vital to note that benchmarking studies on isotopes are severely limited. For example, an earlier benchmark study of RTMs focused on simulating chromium (Cr) isotope fractionation during aqueous kinetic Cr(VI) reduction [22]. However, the simulation approaches employed by the RTMs in the Wanner et al. [22] benchmark were solely tested for first-order reaction kinetics and would not be applicable in simulating isotope fractionation associated with Monod-type rate laws. Similarly, the carbon isotope benchmark study by Druhan et al. [23] focused exclusively on a system where multiple aqueous species exist in tandem and require distinct equilibrium partitioning of the stable isotopes of carbon. In contrast, this particular benchmark helps provide

users' differences (or similarities) across codes based on capabilities for kinetic isotope fractionation, biomass growth, and different rate law formulations. Moreover, the coupled abiotic-biotic Fe-S cycling provides a new reference for future isotopic modeling studies.

The focus of this paper is thus to present a benchmark problem set for the simulation of kinetic sulfur isotope fractionation during DSR. Problems in this benchmark are derived from a well-characterized large column experiment [10]. Processes impacting sulfur isotopic composition such as microbial sulfate reduction and iron-sulfide reactions are included in the problem set. In addition, this benchmark also explores the different mathematical formulations in the representation of kinetic sulfur isotope fractionation as employed in the different RTMs. The multicomponent reactive transport codes used in this benchmark are the following: CrunchTope, TOUGHREACT, PHREEQC, and PHT3D. Capabilities and key features of the codes are described in Steefel et al. [24] and a quick summary is provided in Table 1.

2 Mathematical model description

2.1 Mass balance

All RTM software described herein utilize some form of a general reactive transport equation for a chemical species i as follows:

$$\frac{\partial(\phi S_L C_i)}{\partial t} = \nabla \cdot (\phi S_L D_i \nabla C_i) - \nabla \cdot (q C_i) - \sum_{j=1}^{N_j} \nu_{ij} R_j - \sum_{l=1}^{N_l} \nu_{il} R_l - \sum_{m=1}^{N_m} \nu_{im} R_m \quad (1)$$

where the term on the left hand side is the mass accumulation rate, the terms on the right hand side are diffusion/dispersion, advection terms, and reaction terms: aqueous phase reactions, R_j , mineral reactions, R_l , and gas reactions, R_m . N_x (where $x = j, l, \text{ or } m$) represents the total number of reactions (aqueous, mineral, and gas phase respectively) that involve species i ; ν_{ix} represents the stoichiometric coefficient of i associated with reaction x . Here ϕ is porosity, S_L is liquid saturation, C_i is concentration (mol kgw^{-1}), D is the diffusion/dispersion coefficient ($\text{m}^2 \text{ s}^{-1}$), and q is the Darcy flux (m s^{-1}). The reaction network consists of aqueous speciation reactions, ion exchange, kinetic mineral dissolution and precipitation reactions, and microbially mediated redox reactions, which are described in the following subsections.

2.2 Aqueous equilibrium reactions and mineral dissolution and precipitation reactions

Aqueous speciation reactions between independent chemical components (i.e., primary species) and their derivatives (e.g.,

Table 1 Reactive transport models used in this benchmark study and their capabilities. Adapted from Steefel et al. [24] and Arora et al. [42]

Reactive transport model	Capabilities
PHREEQC	<ul style="list-style-type: none"> • 1D reactive multicomponent transport model • Transport reaction coupling based on sequential non-iterative approach (SNIA) • Discretization scheme–finite differences (mixing cells)
TOUGHREACT (TR)	<ul style="list-style-type: none"> • 3D reactive multicomponent transport model • SNIA • Integrated finite differences (finite volumes)
CrunchTope (CT)	<ul style="list-style-type: none"> • 3D reactive multicomponent transport model • SNIA (also global implicit approach) • Integrated finite differences (finite volumes)
PHT3D	<ul style="list-style-type: none"> • 3D reactive multicomponent transport model • Transport reaction coupling based on sequential non-iterative approach (SNIA) • Discretization scheme–finite differences (mixing cells)

secondary species) are assumed to equilibrate instantaneously. Concentrations of secondary species are calculated as a function of the primary species using the law of mass action:

$$C_i = K_i^{-1} \gamma_i^{-1} \prod_{j=1}^{N_c} (\gamma_j C_j)^{v_{ij}} \quad (2)$$

where C is the concentration, K is the thermodynamic equilibrium constant, γ is the activity coefficient, v_{ij} is the stoichiometric coefficients in the reaction, N_c is the number of primary species, and subscripts j and i refer to the primary and secondary species respectively. In this benchmark, the following primary species are included in the model: H^+ , Fe^{2+} , Fe^{3+} , Na^+ , Ca^{2+} , Mg^{2+} , Cl^- , $^{32}SO_4^{2-}$, $^{34}SO_4^{2-}$, $H_2^{32}S_{(aq)}$, $H_2^{34}S_{(aq)}$, CH_3COO^- , $CO_{2(aq)}$, $O_{2(aq)}$, and NH_4^+ . Secondary species and corresponding reaction stoichiometry are listed in Table S1.

Mineral dissolution and precipitation rates are calculated following a transition state theory (TST) rate expression [25]:

$$R_l = k_m A_m \left[1 - \left(\frac{Q}{K_m} \right)^{m_2} \right]^{m_1} \quad (3)$$

where R_l is the reaction rate ($\text{mol kgw}^{-1} \text{s}^{-1}$), k_m is the rate constant ($\text{mol m}^{-2} \text{s}^{-1}$), A_m is the mineral reactive surface area ($\text{m}^2_{\text{mineral kgw}^{-1}}$), Q refers to the ion activity product of a mineral dissolution/precipitation reaction, and K_m is the corresponding equilibrium constant. Exponents m_1 and m_2 are fitting parameters determined from experiments. A linear form to Eq. 3 is assumed, and the exponents m_1 and m_2 are set to 1. Table 2 shows the reaction stoichiometry of mineral reactions involved in these benchmark problems.

2.3 Representation of microbially-mediated reaction kinetics

The bioenergetics conceptual approach as developed by Rittman and McCarty [26] has been adopted to quantitatively

relate bacterial growth and energetics of redox reactions. In this framework, sulfate-reducing microorganisms (SRM, represented as $C_5H_7O_2N$) catalyze the reaction between an electron donor (acetate in this case) and an electron acceptor (sulfate in this case) to derive energy for respiration, growth, and maintenance. A dual Monod equation is utilized to mathematically represent the coupled microbial sulfate reduction and acetate oxidation:

$$R_k = \mu[SRM] \frac{[eDonor]}{[eDonor] + K_{eDonor}} \frac{[eAcceptor]}{[eAcceptor] + K_{eAcceptor}} \quad (4)$$

where R_k ($\text{mol kgw}^{-1} \text{day}^{-1}$) is the growth rate of the SRM, μ ($\text{mol mol-C}_5\text{H}_7\text{O}_2\text{N}^{-1} \text{day}^{-1}$) is the maximum specific growth rate, and K_e (mol kgw^{-1}) is the half saturation constant of the electron donor/acceptor. The decay of biomass is modeled with a first-order decay model with a decay constant of 0.00027 day^{-1} following Druhan et al. [10].

2.4 Representation of sulfur isotope fractionation kinetics

Druhan et al. [10] explicitly represented the kinetics of the individual isotopologues of sulfur, $^{32}SO_4^{2-}$, and $^{34}SO_4^{2-}$ through modification of the Monod rate law. Here we show this modification in terms of a single Monod expression concerning sulfate reduction, but it is equally applicable to a dual Monod expression as shown above (Eq. 4)

$$^{32}r = ^{32}\mu[SRM] \frac{[^{32}SO_4^{2-}]}{[^{32}SO_4^{2-}] + ^{32}K_s \left(1 + \frac{[^{34}SO_4^{2-}]}{^{34}K_s} \right)} \quad (5a)$$

$$^{34}r = ^{34}\mu[SRM] \frac{[^{34}SO_4^{2-}]}{[^{34}SO_4^{2-}] + ^{34}K_s \left(1 + \frac{[^{32}SO_4^{2-}]}{^{32}K_s} \right)} \quad (5b)$$

Table 2 Mineral reactions and associated parameters for problem level 3

Stoichiometry	k (mol/m ² /s)
$\text{CaCO}_{3(s)} + \text{H}^+ \leftrightarrow \text{Ca}^{2+} + \text{HCO}_3^-$	7.9433E-5
$\text{MgCO}_{3(s)} + \text{H}^+ \leftrightarrow \text{Mg}^{2+} + \text{HCO}_3^-$	4.1687E-10
$\text{Fe}^{2+} + \text{H}^{32}\text{S}^- \leftrightarrow \text{Fe}^{32}\text{S}_{(am)} + \text{H}^+$	1.0E-6
$\text{Fe}^{2+} + \text{H}^{34}\text{S}^- \leftrightarrow \text{Fe}^{34}\text{S}_{(am)} + \text{H}^+$	1.0E-6
$2\text{Fe}^{2+} + \text{H}^+ + {}^{32}\text{S}_{(s)} \leftrightarrow 2\text{Fe}^{3+} + \text{H}^{32}\text{S}^-$	1.0E-3
$2\text{Fe}^{2+} + \text{H}^+ + {}^{34}\text{S}_{(s)} \leftrightarrow 2\text{Fe}^{3+} + \text{H}^{34}\text{S}^-$	1.0E-3
$\text{Fe}(\text{OH})_{3(s)} + 3\text{H}^+ \leftrightarrow \text{Fe}^{3+} + 3\text{H}_2\text{O}$	3.715E-10

By assuming a common half saturation constant for both ${}^{32}r$ and ${}^{34}r$, we arrive at the following simplified set of isotope-specific dual Monod rate laws that also incorporate a dependency on electron donor (acetate) concentration:

$${}^{32}r = {}^{32}\mu[\text{SRM}] \frac{[\text{}^{32}\text{SO}_4^{2-}]}{[\text{}^{32}\text{SO}_4^{2-}] + K_S^{\text{SO}_4}} \frac{[\text{Ac}]}{[\text{Ac}] + K_S^{\text{Ac}}} \quad (6a)$$

$${}^{34}r = {}^{34}\mu[\text{SRM}] \frac{[\text{}^{34}\text{SO}_4^{2-}]}{[\text{}^{34}\text{SO}_4^{2-}] + K_S^{\text{SO}_4}} \frac{[\text{Ac}]}{[\text{Ac}] + K_S^{\text{Ac}}} \quad (6b)$$

As a result, the kinetic fractionation factor (α) is simply a ratio of the rate constants as shown below:

$$\alpha = \frac{{}^{34}\mu}{{}^{32}\mu} \quad (7)$$

In this work, all RTMs utilize kinetic expressions as described by Eqs. 6 and 7 for all problem sets.

One of the goals of this paper is the comparison of the different mathematical formulations in the representation of kinetic sulfur isotope fractionation as employed in the different RTMs. At problem level 2, another mathematical algorithm used in van Breukelen et al. [20, 27] is incorporated in PHREEQC for comparison with the algorithm as utilized in Druhan et al. [10]. In this treatment, for each compound, the light and the heavy isotope species were defined to reflect the compound's fraction of total concentration for the light and heavy isotopes, respectively. In other words, the concentration of an isotope species is here taken to be equal to its fraction multiplied by the compound's total concentration. Reaction rates are then given as follows:

$${}^{32}r = R_k \cdot \left(\frac{[\text{}^{32}\text{SO}_4^{2-}]}{[\text{SO}_4^{2-}]} \right) \quad (8a)$$

$${}^{34}r = R_k \cdot \left(\frac{[\text{}^{34}\text{SO}_4^{2-}]}{[\text{SO}_4^{2-}]} \right) \cdot [\varepsilon + 1] \quad (8b)$$

where ${}^{32}r$ and ${}^{34}r$ are the rates of the light and heavy isotopes, respectively, R_k is the overall rate of sulfate reduction (Eq. 4), and ε is the kinetic isotope enrichment factor of the reaction step ($\varepsilon = \alpha - 1$).

3 Benchmark problem setup

Problem sets in this work are adapted from a set of reactive transport modeling studies based on both column and field experiments of a well-established US Department of Energy research site: the Integrated Field Research Challenge (IFRC), located in Rifle, CO, USA [5, 28–33]. The Rifle site is a former uranium ore processing facility. Bioremediation of uranium in the Rifle aquifer has been demonstrated through the injection of excess labile organic carbon in the form of dissolved acetate. Specifically, acetate was injected into the subsurface over multiple experiments to stimulate both Fe(III) and SO_4^{2-} reducing microbes and mediate the reduction of soluble U(VI) to insoluble U(IV), effectively reducing U(VI) concentrations in the groundwater [5, 28, 29]. These field experiments coupled with reactive transport modeling studies [34–39] have yielded numerous insights pertaining to bioremediation of uranium and similar heavy metals and radionuclides in subsurface environments.

For the present benchmarking study, chemical and isotopic data used to constrain reaction rates, as simulated by the RTM, are taken from a large-scale laboratory column experimental study of organic carbon amended sulfate reduction in soils from the Rifle site [10, 39]. Druhan et al. [10] set up a large-scale (1 m in length) flow through column to replicate field conditions of acetate amended biostimulation experiments in Rifle. One of the goals of the study was to demonstrate incorporation of revised mathematical formulations for microbially mediated redox reactions which can accurately model kinetics of stable isotopes of sulfur (e.g., Eq. 7) over wide range of substrate availability. The mathematical formulation was originally incorporated and validated using the CrunchTope RTM software [24]. The column replicated patterns observed in the field [8]: including an initial period of time in which iron reduction was the dominant reduction process, after which, sulfate reduction became the principle source of acetate consumption (Fig. S1). The CrunchTope simulation was also able to accurately capture the observed enrichment in $\delta^{34}\text{S}$ of both sulfate and sulfide, including both aqueous phase reactant and product pools as well as reduced sulfur minerals. The problem levels in this benchmark are derived from this well-characterized large column experiment. The benchmark is divided into three levels of incremental complexity with a sequential addition of reactive processes.

3.1 Microbial sulfate reduction along a 1D flow path (problem level 1)

The large-scale flow column is modeled as a 1D system of 1 m (height of column) discretized into 100 grid blocks of 0.01 m, with a porosity of 0.32. A constant flow rate of 12.7 cm day^{-1} , similar to the experiments, was set in the model. The diffusion coefficient was assumed to be the same for aqueous chemical species and was set to $0.919 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$ and dispersivity is set at 4 cm [10]. Concentrations of the chemical species at the initial condition and influx boundary condition follow those from Druhan et al. [10] and are listed in Table 3. In this problem, only microbial sulfate reduction is simulated, and the concurrent reduction of Fe(III) by a separate population of microbial biomass is omitted for simplicity (see level 3). The terminal electron accepting process (TEAP), sulfate reduction, consists of two components: catabolic energy production and anabolic growth. This means that for each mole of electron donor (i.e., organic carbon substrate) utilized, a portion, f_s , is used by the SRM for cell synthesis (anabolic) while the remaining fraction, f_e , is used for energy production (catabolic) [26]. This catabolic component is the fraction of total acetate consumption that is coupled to the reduction of sulfate. For all simulations in this study, we assume a constant value of $f_s = 0.08$ and $f_e = 0.92$, such that 8% of the electrons provided by acetate oxidation are utilized for cell synthesis while the remaining 92% are used for energy production [10, 26]. The resulting stoichiometric equation is shown in Table 4.

Associated kinetic parameters for microbial sulfate reduction are also listed in Table 4.

3.2 Simulation of sulfate kinetic isotope fractionation along a 1D flow path (problem level 2a)

This problem builds on the processes and parameters described for problem level 1. The key difference is that at this level, kinetics of $^{32}\text{SO}_4^{2-}$, $^{34}\text{SO}_4^{2-}$, H^{32}S^- , and H^{34}S^- are explicitly represented (Table 3). The simulated kinetic fractionation factor is 0.987. Associated kinetic parameters for $^{32}\text{SO}_4^{2-}$ and $^{34}\text{SO}_4^{2-}$ reductions are listed in Table 4.

3.2.1 Simulation of sulfate kinetic isotope fractionation under reactant limitation (problem level 2b)

This problem is similar in setting to problem level 2a. The only modification is that the initial and influent concentrations of $^{32}\text{SO}_4^{2-}$ and $^{34}\text{SO}_4^{2-}$ are reduced by one order of magnitude (lower than the half saturation constant of sulfate) (Table 3). The aim of this problem level is to verify the appropriate behavior of the kinetic isotope fractionation algorithm (Eq. 6) when the substrate concentration (sulfate in this case) is lower than the half saturation constant. The Monod-type rate expressions are of the functional form: $x/(x + K_s)$. Under conditions where the concentration of sulfate (or acetate) remains much higher than K_s , the net rate is effectively subject to zero-order behavior. However, when the concentration of

Table 3 Initial and influent concentration of the primary aqueous species in all levels

Primary species	Initial concentration (mmol/kg H ₂ O)	Influent concentration (mmol/kg H ₂ O)
pH	7.2	7.2
Fe ²⁺	1.0E-08	1.0E-08
Fe ³⁺	In equilibrium with iron hydroxide	In equilibrium with iron hydroxide
Na ⁺	10.6	22.0
Ca ²⁺	4.47	5.28
Mg ²⁺	4.4	4.34
Cl ⁻	3.00	3.00
HCO ₃ ⁻	8.66	3.25
Acetate	0.0	9.70
O _{2(aq)}	15.6E-3	15.6E-3
NH ₄ ⁺	1.5	1.5
Br ⁻	0.0	1.32
$^{32}\text{SO}_4^{2-} \text{ } ^\wedge$	8.80	8.80
$^{34}\text{SO}_4^{2-} \text{ } ^\wedge\wedge$	3.9107925E-1	3.9107925E-1
H ³² S ⁻ **	1.0E-15	1.0E-15
H ³⁴ S ⁻ **	4.44083904E-17	4.44083904E-17

* Concentrations of $^{32}\text{SO}_4^{2-}$ and $^{34}\text{SO}_4^{2-}$ summed together in level 1

** Concentrations of H³²S⁻ and H³⁴S⁻ summed together in level 1

[^] Initial and influent concentrations of $^{32}\text{SO}_4^{2-}$ are 0.88 mmol/kg H₂O

^{^^} Initial and influent concentrations of $^{34}\text{SO}_4^{2-}$ are 3.9107925E-2 mmol/kg H₂O

Table 4 Sulfate reduction terminal electron accepting process (TEAP) stoichiometry and associated parameters in levels 2 and 3. Iron reduction TEAP stoichiometry and associated parameters in Level 3

Microbe-mediated redox reactions		μ	K_{TEAP} (mol/kg H ₂ O)	K_{AC}
(i)	$0.115 \text{ }^{32}\text{SO}_4^{2-} + 0.125 \text{ CH}_3\text{CHOO}^- + 0.004 \text{ NH}_4^+ + 0.121 \text{ H}^+ \rightarrow 0.004 \text{ C}_5\text{H}_7\text{O}_2\text{N}_{\text{SRB}} + 0.23 \text{ HCO}_3^- + 0.115 \text{ H}_2 \text{ }^{32}\text{S}_{(\text{aq})} + 0.012 \text{ H}_2\text{O}$	$3.21 \times 10^2 \text{ (mol/mol-C}_5\text{H}_7\text{O}_2\text{N/day)}^{++}$	5.0×10^{-3}	1.0×10^{-4}
(ii)	$0.115 \text{ }^{34}\text{SO}_4^{2-} + 0.125 \text{ CH}_3\text{CHOO}^- + 0.004 \text{ NH}_4^+ + 0.121 \text{ H}^+ \rightarrow 0.004 \text{ C}_5\text{H}_7\text{O}_2\text{N}_{\text{SRB}} + 0.23 \text{ CO}_{2(\text{aq})} + 0.115 \text{ H}_2 \text{ }^{34}\text{S}_{(\text{aq})} + 0.012 \text{ H}_2\text{O}$	$3.169212 \times 10^2 \text{ (mol/mol-C}_5\text{H}_7\text{O}_2\text{N/day)}^{++}$	5.0×10^{-3}	1.0×10^{-4}
(iii)	$\text{Fe(III)(S)}^s + 0.208 \text{ CH}_3\text{CHOO}^- + 1.925 \text{ H}^+ + 0.033 \text{ NH}_4^+ \rightarrow 0.033 \text{ C}_5\text{H}_7\text{O}_2\text{N}_{\text{FeRB}} + 1.6 \text{ H}_2\text{O} + 0.25 \text{ HCO}_3^- + \text{Fe}^{2+}$	$2.7 \times 10^{-7} \text{ (mol/m}^3 \text{ cell/day)}^{++}$		1.0×10^{-5}

⁺⁺ Units follow Druhan et al. (2014)

sulfate becomes much lower than the corresponding K_s value, the net rate approaches first-order behavior. The purpose of these isotope-specific rate law derivations is to allow smooth transition between the two end-member rate laws.

3.2.2 Comparison of kinetic isotope fractionation algorithms (problem level 2a and b)

In this level (2a and b), in addition to the formulation as shown in Druhan et al. [10] (Eq. 7a and b), the formulation as described in van Breukelen et al. [20, 27] (Eq. 8a and b) is incorporated in PHREEQC for comparison. Simulation results using this method are denoted as PHREEQC-alt in figures.

3.3 Simulation of sulfate kinetic isotope fractionation along a 1D flow path with microbial iron reduction and mineral dissolution/precipitation reactions (problem level 3)

This problem level adds mineral dissolution/precipitation reactions to level 2a. Seven minerals are included in this benchmark problem: calcite, magnesite, Fe(OH)_3 , Fe^{32}S , Fe^{34}S , $^{32}\text{S}^0$, and $^{34}\text{S}^0$. These minerals follow the mineral network described in Druhan et al. [10] and are considered to react under kinetic constraints and follow the TST rate law (eq. 3). $^{32}\text{S}^0$ and $^{34}\text{S}^0$ reaction rates are further dependent on the

activities of $\text{H}_2 \text{ }^{32}\text{S}_{(\text{aq})}$ and $\text{H}_2 \text{ }^{34}\text{S}_{(\text{aq})}$ respectively. The concurrent reduction of Fe(III) by a separate population of microbial biomass is also incorporated into this problem level. The mineral reactions used in this level are listed in Table 2. The resulting stoichiometric equation is shown in Table 4. Associated kinetic parameters for microbial iron and sulfate reduction are also listed in Table 4. Initial volume fraction and surface area of the minerals are listed in Table 5.

4 Results and discussion

4.1 Microbial sulfate reduction along a 1D flow path (problem level 1)

Simulation results show that the effluent pH of the system begins at 7.2 and increases asymptotically to 8 at the end of the experiment. Effluent sulfate concentrations remain near the initial and influent value of 9.2 mM for approximately the first 20 days of the experiment, a sign of insignificant microbial sulfate reduction. Beyond 20 days, the rate of microbial sulfate reduction began to increase, resulting in a corresponding decrease of both sulfate and acetate concentrations along the length of the column. Simulated effluent sulfate decreased to ~ 0 from 9.2 mM, while effluent acetate decreased to ~ 0 from initial values of 9.7 mM. At this problem

Table 5 Initial conditions of mineral volume fraction for problem level 3

Mineral	Surface area	Volume fraction ($\text{m}^3 \text{ m}^{-3}$) (mineral volume / total volume of solids)
Calcite	$1.04\text{E-}3 \text{ (cm}^2_{\text{mineral}} \text{ g}_{\text{mineral}}^{-1})$	0.069376
Magnesite	$1.00\text{E}6 \text{ (m}^2_{\text{mineral}} \text{ m}^3_{\text{mineral}}^{-1})$	1.47E-6
$\text{Fe}^{32}\text{S}_{(\text{am})}$	$315 \text{ (cm}^2_{\text{mineral}} \text{ g}_{\text{mineral}}^{-1})$	0
$\text{Fe}^{34}\text{S}_{(\text{am})}$	$315 \text{ (cm}^2_{\text{mineral}} \text{ g}_{\text{mineral}}^{-1})$	0
^{32}S	$315 \text{ (cm}^2_{\text{mineral}} \text{ g}_{\text{mineral}}^{-1})$	0
^{34}S	$315 \text{ (cm}^2_{\text{mineral}} \text{ g}_{\text{mineral}}^{-1})$	0
Fe(OH)_3	$1.06\text{E}3 \text{ (m}^2_{\text{mineral}} \text{ m}^3_{\text{mineral}}^{-1})$	0.001382

level, simulated trends of pH and all primary species from all RTMs are in good agreement (Fig. 1).

4.2 Simulation of sulfate kinetic isotope fractionation along 1D flow path (problem levels 2a and 2b)

In problem level 2a, explicitly representation of $^{32}\text{SO}_4^{2-}$, $^{34}\text{SO}_4^{2-}$, H^{32}S^- , and H^{34}S^- allows us to calculate the sulfur isotope ratios of SO_4^{2-} and HS^- . These values are plotted and compared across all RTMs for the effluent fluid composition through time. Sulfur isotope ratios are reported in standard delta notation, $\delta^{34}\text{S}$ (units of per mil, ‰), where $R_{\text{sample}} = (^{34}\text{S}/^{32}\text{S})_{\text{sample}}$ and $R_{\text{std}} = (^{34}\text{S}/^{32}\text{S})_{\text{std}}$. R_{std} is the Canyon Diablo troilite standard ($= 0.0441626$):

$$\delta^{34}\text{S} = \left[\left(\frac{R_{\text{sample}}}{R_{\text{std}}} \right) - 1 \times 1000 \right] \quad (9)$$

Effluent sulfate, sulfide, and acetate trends are similar to those in level 1. Specifically, the sum of the individual isotopologues (i.e., $^{32}\text{SO}_4^{2-} + ^{34}\text{SO}_4^{2-}$, $\text{H}^{32}\text{S}^- + \text{H}^{34}\text{S}^-$) at level 2a is equivalent to SO_4^{2-} and HS^- curves at level 1 respectively. $\delta^{34}\text{S}$ sulfate remained at the initial and up-gradient boundary value of +6.3‰ for roughly the first 20 days of the experiment. Beyond 20 days, $\delta^{34}\text{S}$ sulfate increased exponentially to a value of +46‰ at the end of the experiment (day 43). $\delta^{34}\text{S}$ of sulfide showed a similar trend, beginning at −6.5‰ and ending at a value of +4.9‰ on the last day of the

experiment. At this problem level, simulated trends of $\delta^{34}\text{S}$ sulfate and sulfide from all RTMs match (Fig. 2).

Figure 3 shows the temporal trends of sulfate concentration and $\delta^{34}\text{S}$ of sulfate at various locations along the column ($x = 20, 40, 60$, and 80 cm) for the complete duration of the 43-day acetate amendment. At any given point in time, sulfate concentration decreases with distance into the column. At day 43, sulfate concentrations are 1.9, 0.76, 0.45, and 0.34 mM at $x = 20, 40, 60$, and 80 cm respectively. $\delta^{34}\text{S}$ of sulfate similarly enriches with distance into the column. At day 43, $\delta^{34}\text{S}$ sulfate values are 22.92, 33.74, 40.61, and 44.47 at $x = 20, 40, 60$, and 80 cm respectively. Sulfate concentration at 20 cm is 1.9 mM, indicating that 80% of the total influent sulfate is removed in the first 20 cm of the column. Simulated spatial trends of sulfate concentration and $\delta^{34}\text{S}$ of sulfate from all RTMs match (Fig. 3).

At problem level 2b, with the initial and influent sulfate concentration much lower than the half saturation constant, the decrease in concentration in the effluent through time was relatively minor, from 0.919 to 0.915 mM over the 43-day period. Sulfide concentration increased correspondingly from 0 to 3.44×10^{-3} mM. Such high concentrations of sulfide only occur as a result of the lack of solid phase precipitation included in this portion of the benchmark. In the same period of time, $\delta^{34}\text{S}$ of sulfate increased from 6.3 to 6.36. At this problem level, simulated trends of $\delta^{34}\text{S}$ sulfate and from all RTMs match (Fig. 4).

Fig. 1 Simulated temporal trends of effluent pH, sulfate, sulfide, and acetate concentrations in level 1

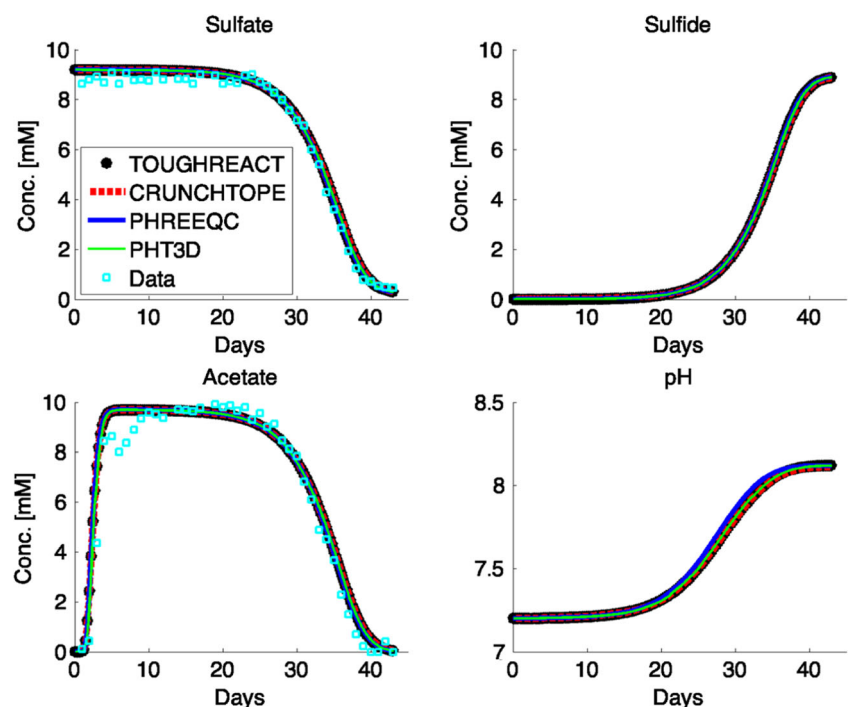
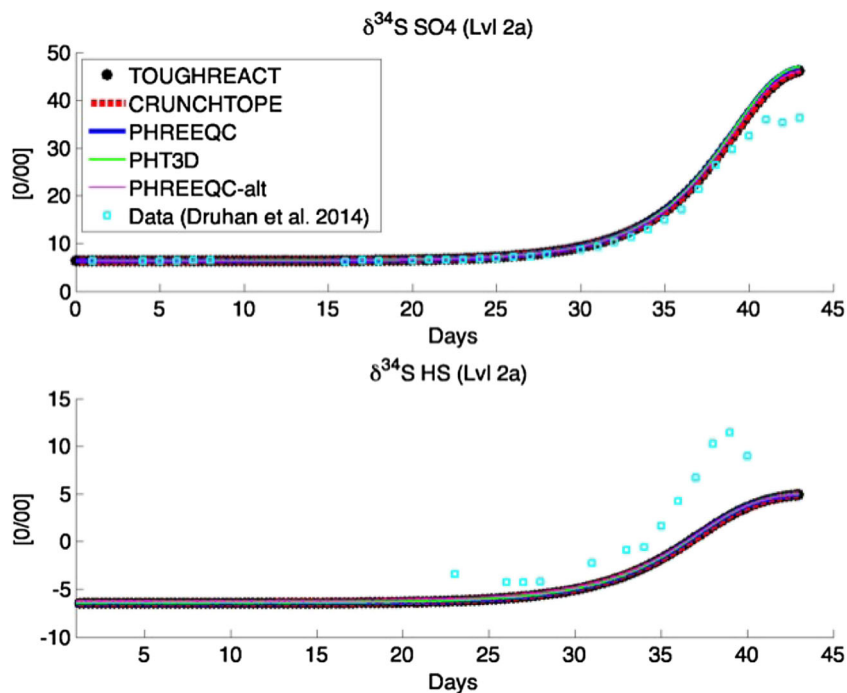


Fig. 2 Simulated temporal trends of effluent $\delta^{34}\text{S}$ sulfate (top) and sulfide (bottom) in level 2a



The results from level 2 show that the mathematical algorithms from van Breukelen et al. [20, 27] and Druhan et al. [10] yield similar behavior. Comparison of the mathematical expressions provides further support. Expanding Eq. 8b and substituting Eq. 7 and $\varepsilon = \alpha - 1$:

$$^{34}r = R_k \cdot \left(\frac{[^{34}\text{SO}_4^{2-}]}{[\text{SO}_4^{2-}]} \right) \cdot [\varepsilon + 1] = ^{32}\mu[\text{SRM}] \cdot \frac{[^{34}\text{SO}_4^{2-}]}{[\text{SO}_4^{2-}] + K_S^{\text{SO}_4^{2-}}} \cdot \frac{[\text{Ac}]}{[\text{Ac}] + K_S^{\text{Ac}}} \cdot \frac{^{34}\mu}{^{32}\mu} \quad (10)$$

Fig. 3 Simulated temporal trends of sulfate and $\delta^{34}\text{S}$ sulfate in level 2a and $x = 20$ (a), 40 (b), 60 (c), and 80 (d) cm at level 2a

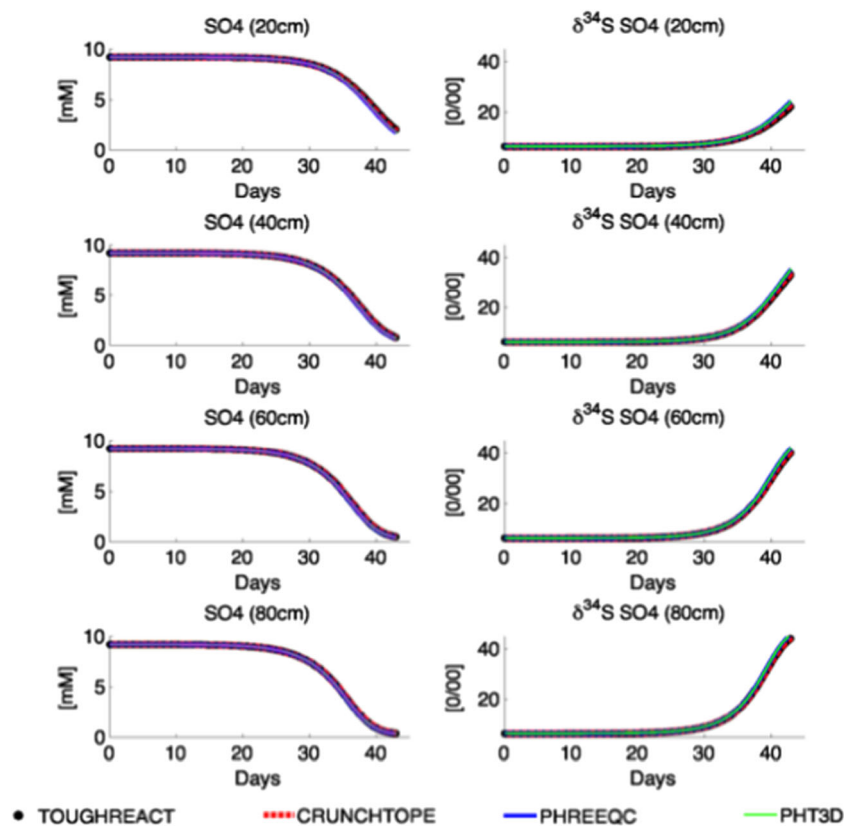
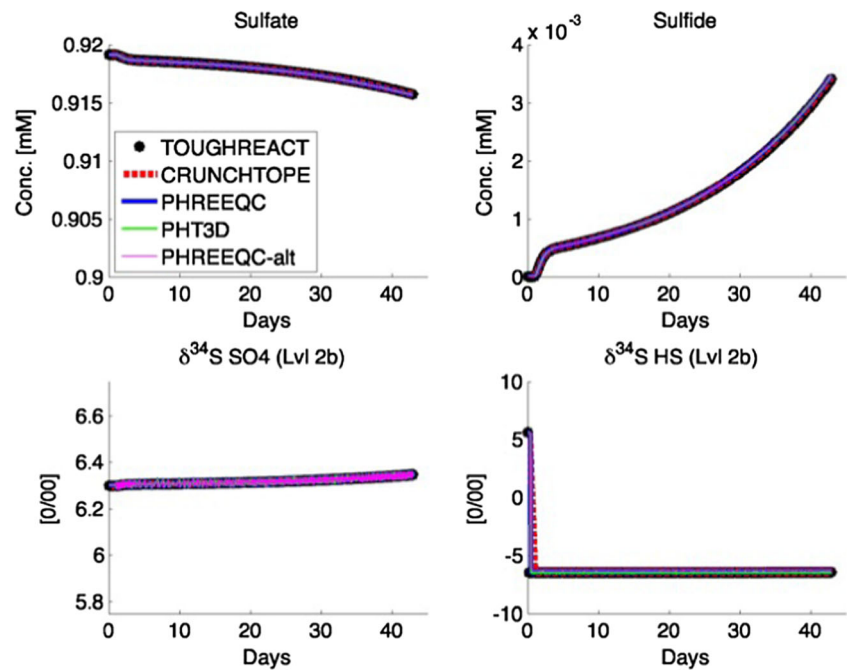


Fig. 4 Simulated temporal trends of effluent sulfate, sulfide concentrations, and $\delta^{34}\text{S}$ sulfate and sulfide in level 2b



We obtain the Eq. 6b under the simplifying assumption that the half saturation constants for the two isotopologues of sulfate are equivalent. These types of Monod formulations as utilized in van Breukelen et al. [20, 27] and Druhan et al. [10] coupled the two isotopologues to each other, thus enabling transition from zero-order to first-order reactions to occur (level 2a). The modified Monod formulation is distinct from the use of two fully independent Monod equations as utilized in earlier models, which may have limited these simulations to either zero-order or first-order conditions [16, 18].

It has been noted in van Breukelen et al. [20, 27] that a correction factor of the form:

$$\times \frac{1}{(A_H \times \alpha \times A_L)} \quad (11)$$

can be applied to each isotope/isotopologue reaction rate when the sum of the reaction rates of the individual isotopologues (e.g., Eq. 8) is lower than R_k , the overall rate. A_H and A_L are the abundances of the heavy and light isotope respectively. In this study, the correction factor was assumed to be ≈ 1 , permitting the derivation of equations as shown above. Further exploration of the impact of the correction factor is warranted. We reran simulation level 2a, this time with the inclusion of the correction factors. Results showed negligible differences between the simulation with and without the correction factor (Fig. S1). In addition, calculation of the correction factor at various time points of the simulation showed that the values ≈ 1 , justifying our assumption.

4.3 Simulation of sulfate kinetic isotope fractionation along a 1D flow path with microbial iron reduction and mineral dissolution/precipitation reactions (problem level 3)

Effluent sulfate, sulfide, acetate, $\delta^{34}\text{S}$ sulfate, and $\delta^{34}\text{S}$ sulfide trends are similar to those in level 2a (Fig. 5). Similar to the original study [9], $\delta^{34}\text{S}$ sulfide trend was simulated through inclusion of a Fe-S precipitate as well as the formation of elemental sulfur. Effluent pH remained relatively stable at ~ 7.2 throughout the experiment, indicating buffering as a result of water-rock interaction. Carbonate and Fe^{2+} concentrations showed increasing trends, while Ca^{2+} and Mg^{2+} concentrations showed decreasing trends. Fe^{3+} concentration increased rapidly within the first 2 days and thereafter slowly decreased to zero. Calcite volume fraction increased through time while magnesite content remained relatively constant throughout the duration of the experiment (Fig. 6). $^{32}\text{S}^0$ ($^{34}\text{S}^0$) minerals showed increasing trends in response to the increased production of sulfide. At this problem level, simulated trends of aqueous and mineral species of CrunchTope and TOUGHREACT matched reasonably well (Figs. 5 and 6).

In terms of process complexity, this problem level differs from the original Druhan et al. [10] simulation in the omission of a thermodynamic potential factor. This thermodynamic function, F_T limits the microbial-mediated reaction rates based on energetic constraints from the environment. In CrunchTope, F_T follows the Jin and Bethke [40] formulation, which represents

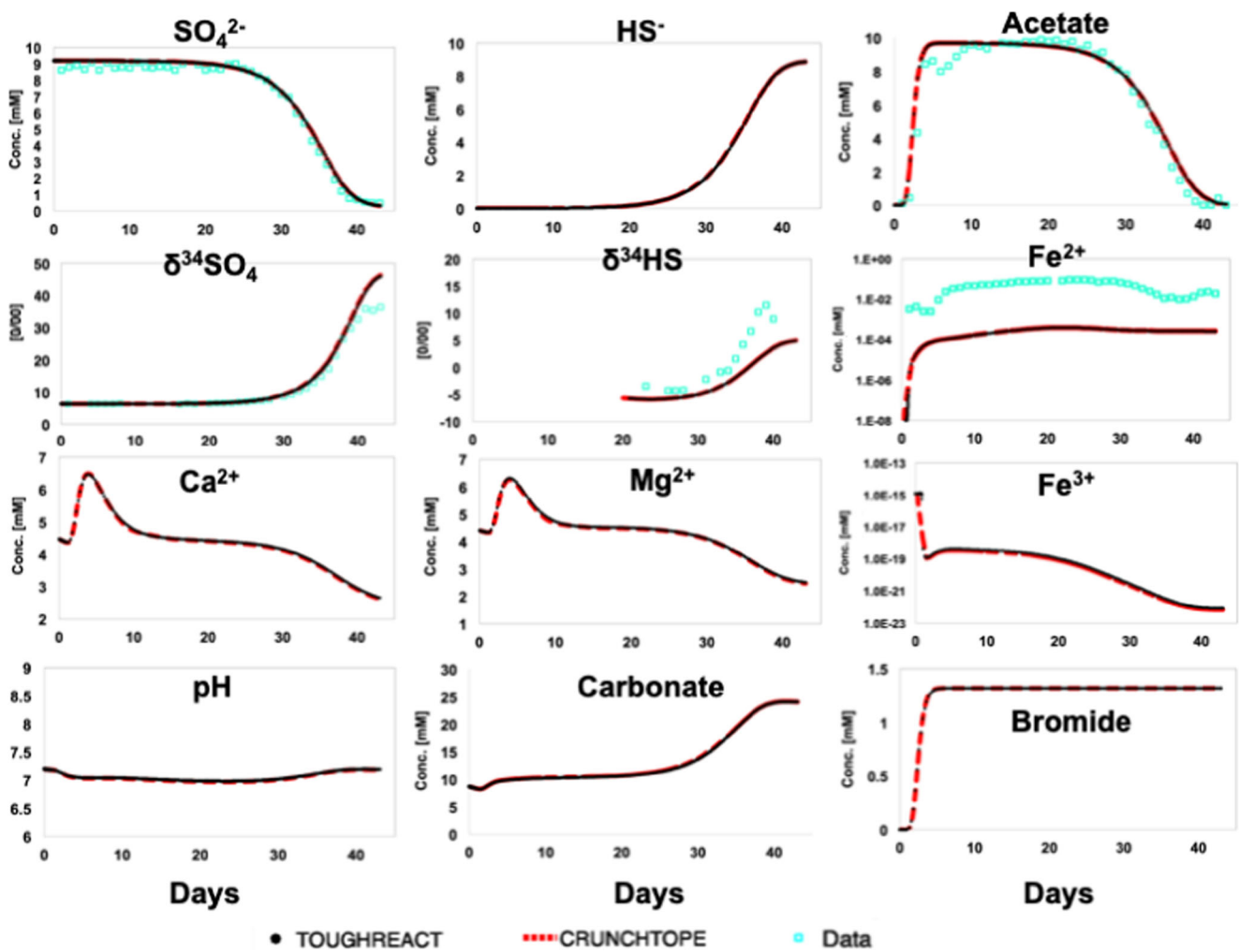
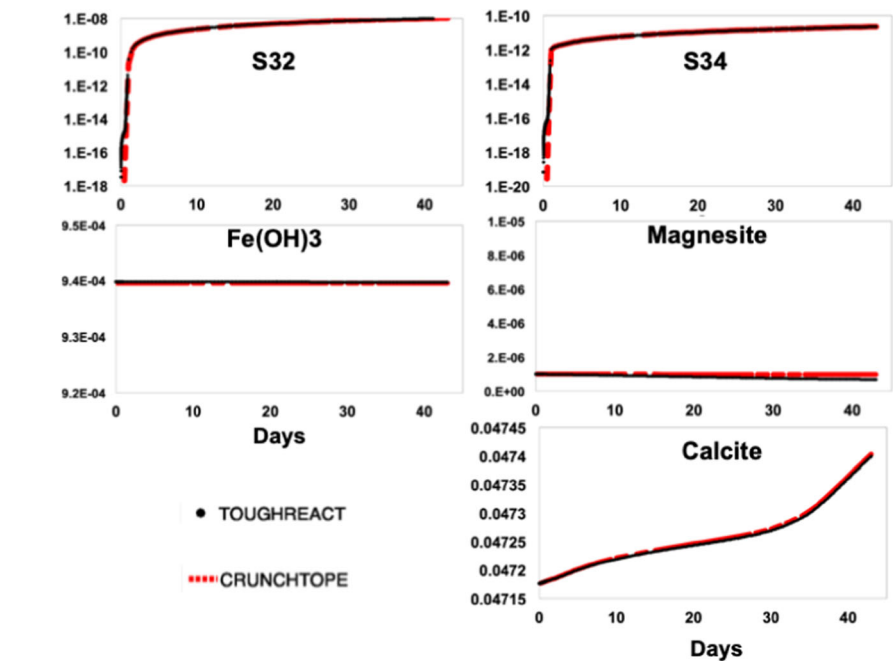


Fig. 5 Simulated temporal trends of effluent major aqueous chemical species and mineral species level 3

Fig. 6 Simulated temporal trends of mineral species level 3



ΔG_{\min} , minimum amount of energy microbial cells harvest for growth and maintenance, as the energetics of ATP synthesis by microorganisms, such that

$$F_T = 1 - e^{\left(\frac{\Delta G_r + m \cdot \Delta G_{ATP}}{\chi \cdot R \cdot T}\right)} \quad (12)$$

where ΔG_r is the Gibbs free energy of a reaction per electron transferred, m is the number of moles of ATP produced per reaction, R is the gas constant ($8.314 \text{ J mol}^{-1} \text{ K}^{-1}$), T is temperature (K), and χ represents the average stoichiometric number for the reaction or the number of times the rate-determining step occurs in the overall reaction. ΔG_{ATP} is the Gibbs free energy required to synthesize 1 mol of ATP, which is typically assumed to be $\sim 60 \text{ kJ mol}^{-1}$ [41]. $m \Delta G_{ATP}$ is therefore the energy threshold required to synthesize ATP. While this thermodynamic function is coded in CrunchTope at the time of this writing, it is not yet available in TOUGHREACT, PHREEQC, and PHT3D. As such, we chose to omit this function in this level so as to allow comparison between CrunchTope and another RTM, TOUGHREACT.

5 Summary and conclusion

A benchmark problem set consisting of three problem levels was developed for the simulation of S isotope fractionation in a 1D flow through column. It was adapted from a large-scale laboratory column experimental study of organic carbon amended sulfate reduction in soils from a uranium-contaminated aquifer. This benchmark adds to the limited benchmarking studies on isotopes.

The first level considered the general conditions of flow and transport and basic sulfate reduction. The second level incorporated the kinetic fractionation of S isotopes under different reactant concentrations. The results were also calculated and compared using two different kinetic fractionation algorithms. In the final problem level, mineral precipitation/dissolution and coupled abiotic-biotic Fe-S cycling processes are systematically introduced to the system. This particular benchmark helps provide users' differences (or similarities) across codes based on capabilities for kinetic isotope fractionation, biomass growth, and different rate law formulations.

The benchmark problem set was solved with the following four well-established reactive transport modeling codes: CrunchTope, TOUGHREACT, PHREEQC, and PHT3D. For the first two problem levels, model-to-model comparisons showed excellent agreement, suggesting that for the tested geochemical processes (i.e., fractionation during microbial sulfate reduction) all of the tested codes are capable to accurately simulate the fate of individual S isotopes. In the final level, when mineral precipitation/dissolution and coupled abiotic-biotic Fe-S cycling processes are incorporated,

CrunchTope and TOUGHREACT simulations match. Future benchmarking efforts should expand to 2D (or 3D) problem levels that provide base case for validation, and cases involving heterogeneous redox and/or permeability distribution to tease apart the impacts of hydrodynamic dispersion and sulfate reduction on sulfur isotope fractionation.

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