

**Tyrosine kinases regulate chondrocyte hypertrophy
promising drug targets for Osteoarthritis**

Ferrao Blanco, M. N.; Domenech Garcia, H.; Legeai-Mallet, L.; van Osch, G. J.V.M.

DOI

[10.1016/j.joca.2021.07.003](https://doi.org/10.1016/j.joca.2021.07.003)

Publication date

2021

Document Version

Final published version

Published in

Osteoarthritis and Cartilage

Citation (APA)

Ferrao Blanco, M. N., Domenech Garcia, H., Legeai-Mallet, L., & van Osch, G. J. V. M. (2021). Tyrosine kinases regulate chondrocyte hypertrophy: promising drug targets for Osteoarthritis. *Osteoarthritis and Cartilage*, 29(10), 1389-1398. <https://doi.org/10.1016/j.joca.2021.07.003>

Important note

To cite this publication, please use the final published version (if applicable).
Please check the document version above.

Copyright

Other than for strictly personal use, it is not permitted to download, forward or distribute the text or part of it, without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license such as Creative Commons.

Takedown policy

Please contact us and provide details if you believe this document breaches copyrights.
We will remove access to the work immediately and investigate your claim.

Osteoarthritis and Cartilage

Review

Tyrosine kinases regulate chondrocyte hypertrophy: promising drug targets for Osteoarthritis

M.N. Ferrao Blanco [†], H. Domenech Garcia [‡], L. Legeai-Mallet [‡], G.J.V.M. van Osch ^{† § || *}

[†] Department of Orthopaedics and Sports Medicine, Erasmus MC, University Medical Center Rotterdam, Rotterdam, the Netherlands

[‡] Université de Paris, INSERM U1163, Institut Imagine, Paris, France

[§] Department of Otorhinolaryngology, Erasmus MC, University Medical Center Rotterdam, Rotterdam, the Netherlands

^{||} Department of Biomechanical Engineering, Delft University of Technology, Delft, the Netherlands



ARTICLE INFO

Article history:

Received 11 February 2021

Accepted 8 July 2021

Keywords:

Tyrosine kinases

Chondrocyte hypertrophy

Disease modifying OA drugs

SUMMARY

Osteoarthritis (OA) is a major health problem worldwide that affects the joints and causes severe disability. It is characterized by pain and low-grade inflammation. However, the exact pathogenesis remains unknown and the therapeutic options are limited. In OA articular chondrocytes undergo a phenotypic transition becoming hypertrophic, which leads to cartilage damage, aggravating the disease. Therefore, a therapeutic agent inhibiting hypertrophy would be a promising disease-modifying drug. The therapeutic use of tyrosine kinase inhibitors has been mainly focused on oncology, but the Food and Drug Administration (FDA) approval of the Janus kinase inhibitor Tofacitinib in Rheumatoid Arthritis has broadened the applicability of these compounds to other diseases. Interestingly, tyrosine kinases have been associated with chondrocyte hypertrophy. In this review, we discuss the experimental evidence that implicates specific tyrosine kinases in signaling pathways promoting chondrocyte hypertrophy, highlighting their potential as therapeutic targets for OA.

© 2021 The Authors. Published by Elsevier Ltd on behalf of Osteoarthritis Research Society International.

This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

OA is a highly prevalent degenerative joint disorder worldwide¹. OA is characterized by a slow-progressive degeneration of the articular cartilage, dysregulation of subchondral bone remodeling and synovial inflammation which ultimately lead to loss of joint function and chronic pain². Despite the main risk factors for OA have been identified³, there is still no effective disease-modifying treatment available.

Accumulating data suggest that articular chondrocytes undergo a phenotypic shift and become hypertrophic in OA⁴. This phenotypic change resembles the process of endochondral ossification, which naturally occurs in the growth plate for the elongation of long bones and lasts from early ante natal period until puberty. Within this process, the temporary cartilage found at the early embryonic stages of endochondral bones differentiates into

hypertrophic cartilage and eventually enters apoptosis or differentiates to osteoblasts⁵, leading to replacement by bone. However, this process should not occur in articular cartilage. Recent studies have confirmed by single cell RNA-seq that hypertrophic differentiation takes place in osteoarthritic articular cartilage^{6,7}.

One of the main characteristics of hypertrophic chondrocytes is the upregulation of proteolytic enzymes, which drive the degradation of extracellular matrix (ECM) components such as type II Collagen and Aggrecan. These enzymes are mainly Matrix Metalloproteinase (MMP) 13 and a disintegrin and Metalloproteinase with Thrombospondin Motif (ADAMTS) 4 and 5. In addition, hypertrophic chondrocytes produce Alkaline Phosphatase (ALPL) that enhances the calcification of the matrix and increase the expression of hypertrophic-related genes, such as type 10 Collagen (COL10A1) and Runt-related transcription factor 2 (RUNX2)². Whereas, a healthy and proliferative chondrocyte is characterized by the expression of SRY-box transcription factor (SOX9) and type II collagen as well as Bagpipe homeobox homolog (BAPX1) (Fig. 1). Therefore, understanding the mechanisms that drive the terminal differentiation of hypertrophic chondrocyte and control the levels of ECM-degrading enzymes is important for developing effective therapies for OA.

* Address correspondence and reprint requests to: G.J.V.M. van Osch, Erasmus MC, University Medical Center Rotterdam, Wytemaweg 80, Room Ee16.55C, 3015 CN Rotterdam, the Netherlands. Tel.: 31-107043661.

E-mail addresses: m.ferraoblanco@erasmusmc.nl (M.N. Ferrao Blanco), h.domenechgarcia@erasmusmc.nl (H. Domenech Garcia), laurence.legeai-mallet@inserm.fr (L. Legeai-Mallet), g.vanosch@erasmusmc.nl (G.J.V.M. van Osch).

Interestingly, recent insights into the field of OA pinpoint that Tyrosine kinases (TKs) may play an important role in regulating chondrocyte hypertrophy. These enzymes are highly involved in cellular differentiation and are the main regulators in several pathways³. However, there is no study that provides an overview of the TKs associated with chondrocyte hypertrophy and the pathways that are involved in this phenotypic transition. To this end, this review aims to evaluate which TKs have been related to chondrocyte hypertrophy during OA pathogenesis and provide a clear overview of the signaling pathways that they activate.

Main pathways driving chondrocyte phenotype

Various pathways have been associated with chondrocyte hypertrophy that takes place during bone development (Fig. 2). Indian hedgehog (IHH) and parathyroid hormone–related protein (PTHrP) are known to generate a negative feedback loop, which allows to maintain chondrocyte homeostasis. Namely, IHH promotes hypertrophy while PTHrP blocks it⁸. Moreover, TGF- β has been shown to prevent hypertrophy via Smad2/3 signaling, through modulating the levels of main chondrocyte phenotype-determining transcriptional regulators such as SOX9 and RUNX2. Conversely, TGF- β might promote hypertrophy when it activates Smad1/5/9 phosphorylation⁹. Another pathway that contributes to chondrocyte hypertrophy is wingless-type (Wnt) signaling. Wnt induces the stabilization of β -catenin in the cytoplasm, allowing its translocation to the nucleus and this eventually enables the

transcription of *RUNX2*¹⁰. Similarly, pro-inflammatory cytokines such as interleukin (IL)-1 β and tumor necrosis factor (TNF)- α , have also shown to upregulate the expression of hypertrophic markers and downregulate SOX9 by stabilizing nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) and hypoxia inducible factor 2 α (Hif-2 α)^{3,8,99}. Other pathways including mitogen-activated protein kinase (MAPK) signaling, phosphoinositide 3-kinase (PI3K)- protein kinase B (AKT) and janus kinase (JAK) 2- (signal transducer and activator of transcription factor (STAT), however, may lead to opposite responses depending on the ligand-receptor by which the transduction is initiated².

The involvement of tyrosine kinase members in driving chondrocyte hypertrophy

TKs are a large subclass of protein kinases, which play a major role in cellular signaling. Through their ability to phosphorylate other proteins at their tyrosine residues, TKs are able to control most fundamental cellular processes such as cell proliferation and differentiation, cell metabolism as well as cellular survival¹¹. Recent studies have entailed TKs as the most pursued targets in current pharmacological research^{3,12} and importantly, there are already several TK inhibitors that are FDA approved¹³.

Current advances in the field of OA have suggested that both, receptor and non-receptor TKs, are associated to articular chondrocyte catabolism (Table I). Some TKs have been associated to the synthesis of inflammatory mediators, such as anexlektin (AXL) and

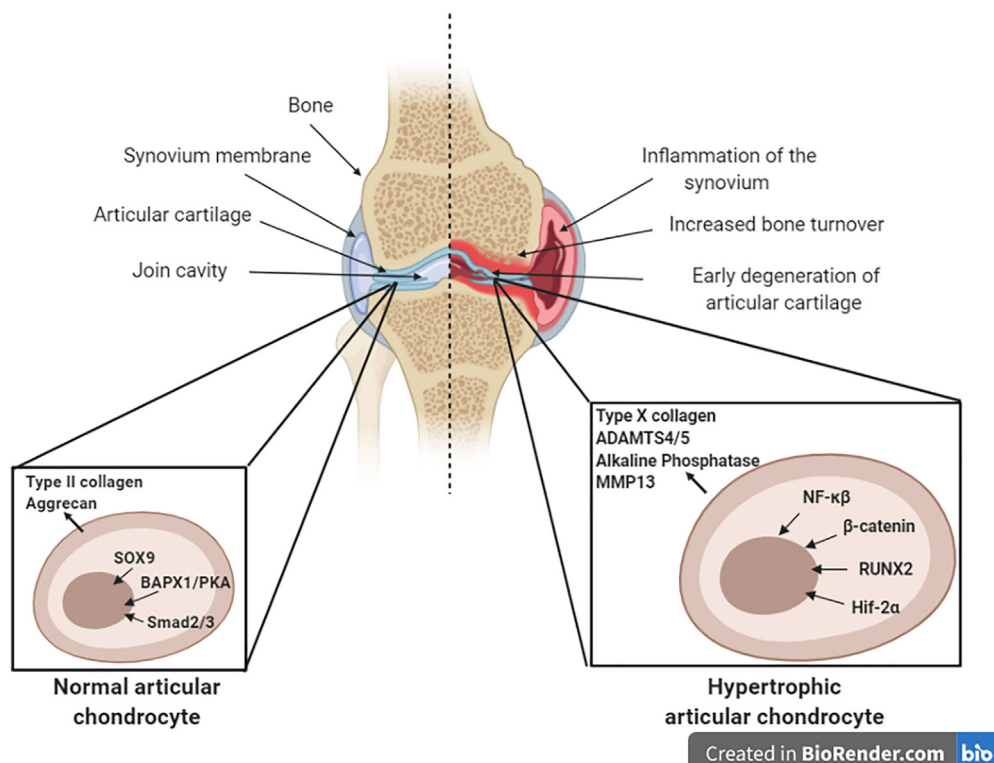


Fig. 1

Schematic representation of main characteristics of normal articular cartilage compared to an osteoarthritic cartilage. On the left, a normal articular chondrocyte is depicted together with the positive stimuli that induce chondrocyte homeostasis. Conversely, on the right a hypertrophic chondrocyte is shown, including the signaling and responses that occur during OA.

tyrosine-protein kinase receptor (TYRO)-3¹⁴, others have been shown to be increased in OA chondrocytes, such as human epidermal growth factor receptor 4 (ERBB4), insulin receptor (IR), c-mesenchymal epithelial transition factor (MET) and vascular endothelial growth factor (VEGFR)-3^{15–18}, to inhibit catabolic factors such as ephrin type B receptor (EPHB) 4¹⁹, or to modulate cartilage degradation such as Ick yes novel tyrosine kinase (LYN), protein tyrosine kinase 5 (FRK), hematopoietic cell kinase (HCK) and spleen tyrosine kinase (SYK)^{20,21}. In spite of this information, the specific role of these TKs in the mechanisms underlying chondrocyte hypertrophy remains to be elucidated.

Receptor tyrosine kinases

Receptor tyrosine kinases (RTK) are the major subgroup of the tyrosine kinase family and are essential components of signal transduction pathways that participate in intercellular communication. Out of 58 types of RTKs that are found in the human genome⁴⁶, ten have been associated to chondrocyte hypertrophy and are described below.

Fibroblast growth factor receptor 1 and 3

Fibroblast growth factor receptors (FGFRs) are ubiquitous regulators of development involved in wound healing, angiogenesis and cartilage homeostasis^{1,47}. Mutations of FGFR genes are the basis of a broad range of skeletal developmental disorders such as craniosynostoses and chondrodysplasia²⁹. FGFRs are activated through binding of fibroblast growth factors (FGFs) ligand and co receptor heparan sulfate to the extracellular domain of the receptor, which induces FGFR dimerization and transphosphorylation of tyrosine residues. Stimulation of the tyrosine kinase domain leads to activation of various downstream signaling proteins either from MAP kinases or PI3K/AKT pathway⁴⁷. So far, four signaling members of FGFRs have been reported FGFR1, FGFR2, FGFR3 and FGFR4⁴⁸. FGFR-1 and FGFR-3 thoroughly investigated in clinical trials as drug targets for various cancers and skeletal dysplasia^{49,50}, have shown to be the most predominant receptors expressed in human cartilage²⁸. Thus, they are suggested to play essential roles in cartilage upon interaction with the FGF-2 and fibroblast growth factor 18 (FGF-18) ligands^{1,2,30,51}. It has been consistently shown that the ratio of FGFR-3 to FGFR-1 is significantly reduced in OA, and that this is mediated by the upregulation of FGF-2^{1,2}. Hence, suggesting

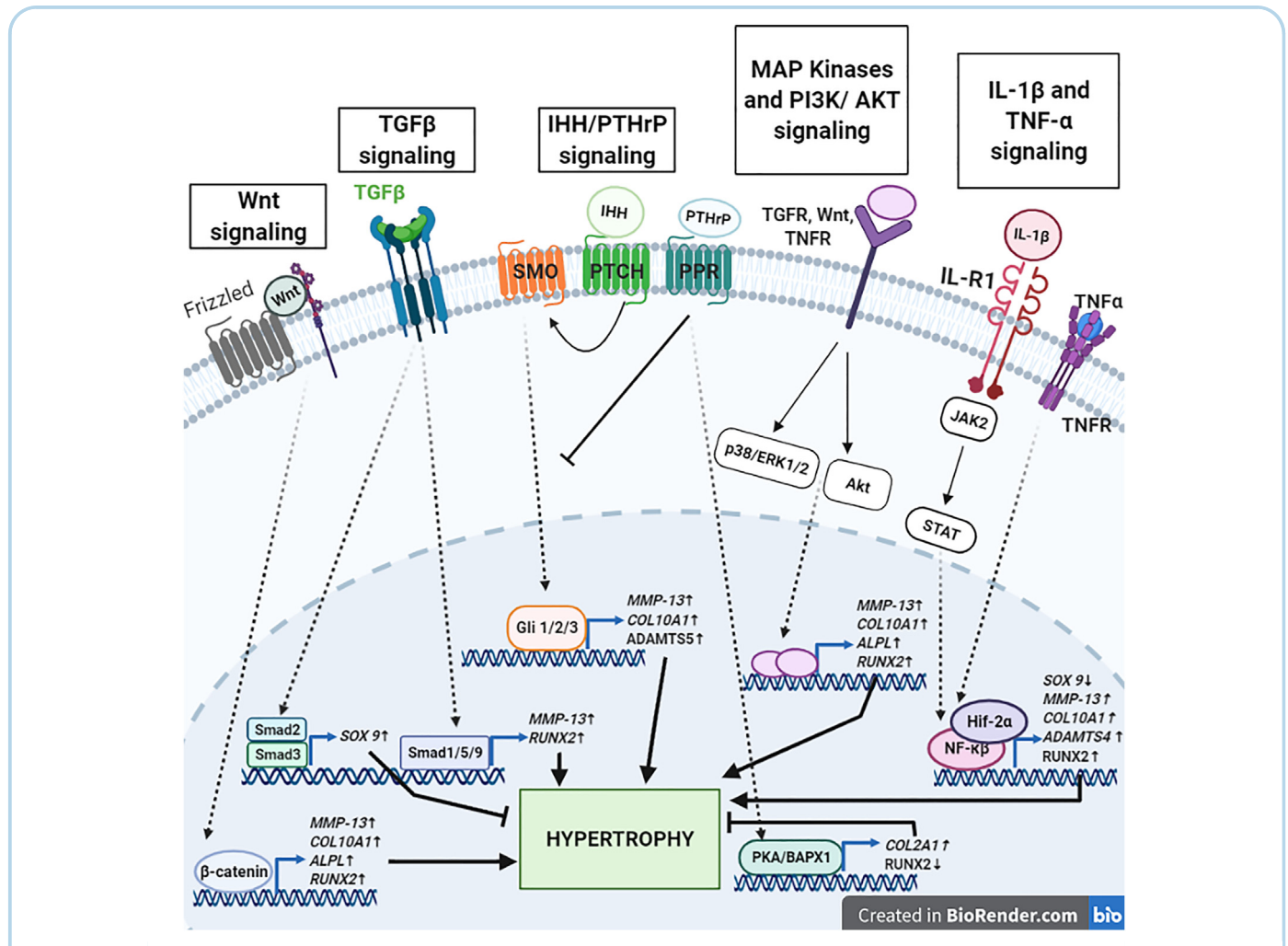


Fig. 2

Overview of the signaling pathways involved in the regulation of chondrocyte homeostasis and their relation to chondrocyte hypertrophy. The arrows and T-shaped lines signify positive and negative actions, respectively.

	Tyrosine Kinase	TK Family	Reference
Receptor	AXL	AXL	14
	TYRO-3		14
	DDR1	DDR	22,23
	DDR2		24–26
	EGFR	EGFR	27
	ERBB4		18
	EPHB 4	EPH	19
	FGFR-1	FGFR	1,2,14,28
	FGFR-3		29–31
	IGF1-R	INSR	32,33
	IR		34
	c-MET	MET	17
	ROR2	ROR	35
	TRK-A	TRK	8,36,37
	VEGFR-1	VEGFR	38,39
	VEGFR-2		16,38,39
	VEGFR-3		8,16,36,37,40
Non-receptor	FAK	FAK	41,42
	JAK2	JAK	43–45
	FRK	FRK	20
	FYN	SRC	3,14
	HCK		20
	LYN		20
	SYK	SYK	21

Table 1

Overview of the receptor and non-receptor tyrosine kinases that have been associated to chondrocyte catabolism

Osteoarthritis
and Cartilage

these FGFRs mediate opposing effects in human chondrocytes²⁸. Suppression of FGFR-1 by G141, a non-ATP-competitive FGFR-1 inhibitor, impaired the expression of *MMP13*, *ADAMTS5* and *COL10A1* as well as reduced the loss of aggrecan in FGF-2 treated human chondrocytes. Additionally, G141 could delay the progression of cartilage degradation in a mouse model of OA¹. Similarly, inhibition of components of the FGFR-1 signaling pathway, specifically FGFR-1 and ERK, abolished FGF-2 driven catabolic events in chondrocytes *in vitro*². Similar effects were observed using Fgfr-1 conditional knock out mice²⁸. Another morphogen that is overexpressed in OA-chondrocytes, FGF-23, can also activate FGFR-1. The addition of FGF-23 to healthy articular chondrocytes led to increased *RUNX2* expression and downregulation of *SOX9*, which is indicative of a hypertrophic phenotype⁵¹.

On the other hand, FGFR-3 is essential for cartilage protection and inhibition of hypertrophic phenotype. In fact, Fgfr-3 (–/–) mice develop spontaneous OA, evidenced by an accelerated degradation of cartilage in association with an upregulation of *Mmp13* and *Col10a1*^{1,30}. Further evidence demonstrated that Fgfr-3 levels were reduced in *mtorc1* knockout mice that displayed cartilage degradation and chondrocyte hypertrophy⁵². Interestingly, exogenous stimulation of FGFR-3 by FGF-18 was able to ameliorate OA³¹. Of note, in a randomized phase II clinical trial the intra-articular injection of FGF-18 increased cartilage thickness, and substantially reduced cartilage loss⁵³. Taken together, recent advances have shown that FGFR-1 induces chondrocyte hypertrophy while FGFR-3 prevents it.

Discoidin domain receptor 1 and 2

The discoidin domain receptors (DDRs) family comprises two members, DDR1 and DDR2, and are the only type I transmembrane receptor TKs that specifically bind to and are activated by

collagen⁵⁴. DDRs mediate cell–collagen interactions under both, normal and pathological conditions. Of note, DDRs regulate cell proliferation, migration, adhesion and tumor metastasis⁵⁵. DDRs can be activated by different types of collagen and each receptor has different affinities for each one. Namely, only DDR1 binds to collagen IV^{54,56}, whereas DDR2 preferentially binds collagen II⁵⁷ and X⁵⁸. Both DDRs activate signaling pathways including PI3K/AKT and rat Sarcoma virus (RAS)/extracellular signal regulated (ERK)/p38^{59–63}.

Provided the affinity of DDR2 for collagen type II and X, it was previously believed that this was the only DDR receptor involved in OA pathogenesis⁶⁴. Several *in vitro* studies investigating the role of DDR2 in OA suggested that cumulative injuries in joint tissues lead to increased collagen II cleavage that results in DDR2 activation by tyrosine autophosphorylation^{24–26}. DDR2 signaling induce overexpression of hypertrophic markers such as *Mmp13*, *Alpl* and *Col10a1* *in vivo*⁶⁵. OA severity was attenuated in *Ddr2*-deficient mutant mice⁶⁶. A recent publication has highlighted that inhibition of DDR1 in OA-mice models decreased expression of *Mmp13* and *Col10a1*²². In the light of these findings, it can be elucidated that both DDRs may be involved in promoting chondrocyte hypertrophy.

Insulin-like growth factor 1 receptor

Insulin-like growth factor receptor 1 (IGF1-R), along with insulin receptor (IR) and IGF2-R, is a member of the IGF signaling pathway that is essential for cell growth and tissue differentiation⁶⁷. Interestingly, a number of clinical trials involving IGF1-R inhibition in breast cancer patients have been published^{68–70}. This receptor mainly mediates the effect of insulin-like growth factor 1 (IGF1) and IGF2. Ligand-activated IGF1-R subsequently recruits and activates signaling intermediates, that include members of the insulin-receptor substrate (IRS) family and Shc. Activation of IRs induces activation of the PI3K/AKT pathway. Conversely, Shc leads to activation of the MAPK/ERK pathway⁷¹. Besides tumorigenesis, IGF1-R is thought to be involved in the regulation of articular cartilage metabolism via IGF-1 binding³³. Namely, stimulation of IGF-1R by IGF-1 impaired interleukin (IL)-1 β induced NF- κ B activation in human chondrocytes. Consequently, hypertrophic differentiation was prevented³². Together these findings suggest a protective role of IGF1-R in OA progression.

Epidermal growth factor receptor

The epidermal growth factor receptor (EGFR, ErbB1) is a member of the ErbB family of receptors, which also includes ErbB2, ErbB3 and ErbB4. EGFR is crucial for the development of several organs during embryonic and postnatal stages⁷². Through its interaction with its growth factor ligands, transforming growth factor (TGF)- α and epidermal growth factor (EGF), EGFR undergoes a transition from monomeric to an active homodimer. Subsequent dimerization leads to protein autophosphorylation and downstream activation of MAPK, AKT and c-jun N-terminal kinase (JNK) pathways. Importantly, EGFR has shown to promote inflammatory processes upon TGF- α binding⁷³. Overexpression of EGFR has been so far related to cancer, fibrosis and inflammatory diseases. Current literature on the field of OA, suggests that TGF- α and EGFR are involved in the molecular events that influence joint cartilage degeneration, since overexpression of both proteins was observed in degenerating cartilage in experimental knee OA⁷³. Moreover, levels of anabolic genes such as *Acan* and *Col2a1* were reduced, whereas *Mmp13* levels were increased in articular chondrocytes in response to TGF- α ⁷³. Particularly, EGFR signaling is thought to drive ECM degradation through the activation of MAPK pathways, which involves the phosphorylation of p38 and ERK kinases. This latter event is essential for the translocation of β -catenin into the nucleus

and the expression of catabolic genes. Moreover, downregulation of EGFR prevents thrombin-induced *Mmp-13* expression in human chondrocytes via PI3K/AKT²⁷. Clinical studies specifically targeting EGFR in OA have not hitherto been reported, albeit it could be of interest provided the participation of EGFR in the regulation of hypertrophic markers.

Vascular endothelial growth factor receptor 1 and 2

Vascular endothelial growth factor receptors (VEGFRs) are responsible for binding with their ligands VEGFs to promote angiogenesis and vasculogenesis during development, wound healing and endochondral ossification. Pathophysiological effects of VEGF/VEGFR signaling have been described in diseases such as cancer and rheumatoid arthritis^{74,75}. In fact, VEGFR inhibitors have been used in clinical trials and were approved by FDA as drug targets for the treatment of cancer, pulmonary fibrosis and macular degeneration^{76,77}. There are three subtypes of VEGF receptors, including VEGFR-1, VEGFR-2 and VEGFR-3, which can be activated by diverse structurally related VEGFs. Binding of VEGF to its receptor leads to protein homo- or heterodimerization, consequently inducing autophosphorylation of the tyrosine residues of the receptor and phosphorylation of downstream signal mediators that mainly participate in MAPK or PI3K/AKT pathways⁷⁸. Besides their involvement in cancer progression, all VEGFRs have also appeared to be highly expressed in OA chondrocytes^{16,38}. However, only the role of VEGF-A and its receptors VEGFR-1/-2 have been extensively studied in the pathophysiology of OA. Both receptor TKs are thought to drive vascular invasion of articular cartilage upon secretion of VEGF-A, a phenomena that is more pronounced in severe OA cartilage^{39,79}. Both, *in vitro* and *in vivo* studies, showed that shRNA-mediated knock-down of VEGF-A was able to ameliorate OA. Namely, VEGF-A inhibition protected cartilage from hypertrophy by decreasing *Mmp13*, *Runx2* and *Col10a1*³⁹. Inhibition of VEGF-A in an OA animal model reduced articular cartilage degradation. In contrast to healthy chondrocytes, VEGF-A downregulation increased expression of *Acan* and *Col2a1* and reduced *Mmp-13* and *Adamts5* levels. Since VEGF-A can bind to both VEGFR-1 and -2, further studies investigating the specific roles of these receptors and their targeted inhibition in OA progression would be of interest.

Tropomyosin receptor kinase A

Tropomyosin receptor kinase A (TrkA), also recognized as high affinity nerve growth factor (NGF) receptor, is a member of the neurotrophic tyrosine kinase receptor (NTRK) family. Two other members constitute this family and are designated TrkB and TrkC. As a kinase, TrkA is responsible for functional NGF signal transduction to promote neuronal differentiation and proliferation, nociceptor response and avoidance of apoptosis. Precisely, the NGF-activated TrkA undergoes dimerization and autophosphorylation at multiple tyrosine residues, leading to the activation of diverse intracellular pathways such as PI3K/AKT and MAPK signaling. In many occasions, the NGF-TrkA complex is internalized via endocytosis and subsequently activates several signaling pathways⁸⁰. NGF-TrkA signaling play a role in the pathophysiology of cancer and OA. Indeed, NGF and TrkA were found to be expressed in human articular chondrocytes and they were upregulated in osteoarthritic chondrocytes in accordance with the degree of tissue injury⁸¹. Besides the involvement of NGF-TrkA signaling in neuropathic pain in OA, this pathway was suggested to participate in chondrocyte homeostasis and hypertrophy⁸². Namely, it was found that the expression of an agonist of NGF and TrkA in human articular chondrocytes promoted chondrocyte hypertrophy. Conversely, inhibition of NGF receptor could also downregulate SOX-9 and increase the expression of hypertrophy-related genes, including *ALPL* and *RUNX2*. The fact that both, over-activation and inactivation, of

NGF-TrkA pathway give similar outcomes might be explained by the disruption of the feedback loop between IHH and PTHrP, two factors involved in regulating the homeostasis of articular chondrocytes⁸. Since 2015, a number of randomized clinical trials have been performed to evaluate the efficacy of TrkA inhibitors GZ389988, ASP7962 and ONO-4474 as treatment for OA knee pain and physical function. Whereas ASP7962 was unable to ameliorate pain and physical function, GZ389988 and ONO-4474 were able to safely reduce pain and gain physical function^{36,37,40}. However, no structural changes were measured that could be associated with hypertrophy. Further research is required to decipher whether TrkA inhibition is a disease-modifying OA drug.

Receptor tyrosine kinase-like orphan receptor 2

Receptor tyrosine kinase-like orphan receptor (ROR) family consists of two members, ROR1 and ROR2, which are essential for regulating skeletal and neuronal development, cell polarity and migration⁸³. The RORs are single-pass transmembrane receptors, with the unique feature that their cysteine-like rich domain resembles that of the Frizzled receptors and thus, non-canonical WNT ligands can bind to the receptor. Interaction of a non-canonical WNT ligand (WNT5A) with ROR triggers the association of ROR with other WNT co-receptors and subsequent RTK activation occurs. The signal is then transduced to downstream pathways such as WNT, PI3K/AKT, MAPK/ERK and Rho/yes-associated protein (YAP), which leads to the expression of genes related with cell proliferation and survival⁸⁴. Since RORs are involved in cell proliferation and they are rarely expressed in adult tissues, they have become an attractive target for cancer therapy^{85,86}. Nonetheless, a recent study showed that specifically ROR2 signaling was upregulated in OA cartilage and impaired chondrocyte differentiation via Rho/YAP pathway. ROR2 inhibition in primary human chondrocytes suppressed expression of *ADAMTS4/5* and increased *SOX9*. Furthermore, blocking ROR2 in a mouse OA model reduce pain and cartilage damage³⁵. This study suggests that ROR2 is associated with WNT5A and hypertrophy and thus, ROR2 could also be a potential target for OA.

Cytoplasmic non-receptor tyrosine kinases

Non-receptor tyrosine kinases (NRTKs) are enzymes that are involved in the transduction of signals initiating from extracellular clues and which often interact with transmembrane receptors⁸⁷. Three members (JAKs, FAK, Fyn) have been described to play a role in chondrocyte hypertrophy.

Janus kinase 2

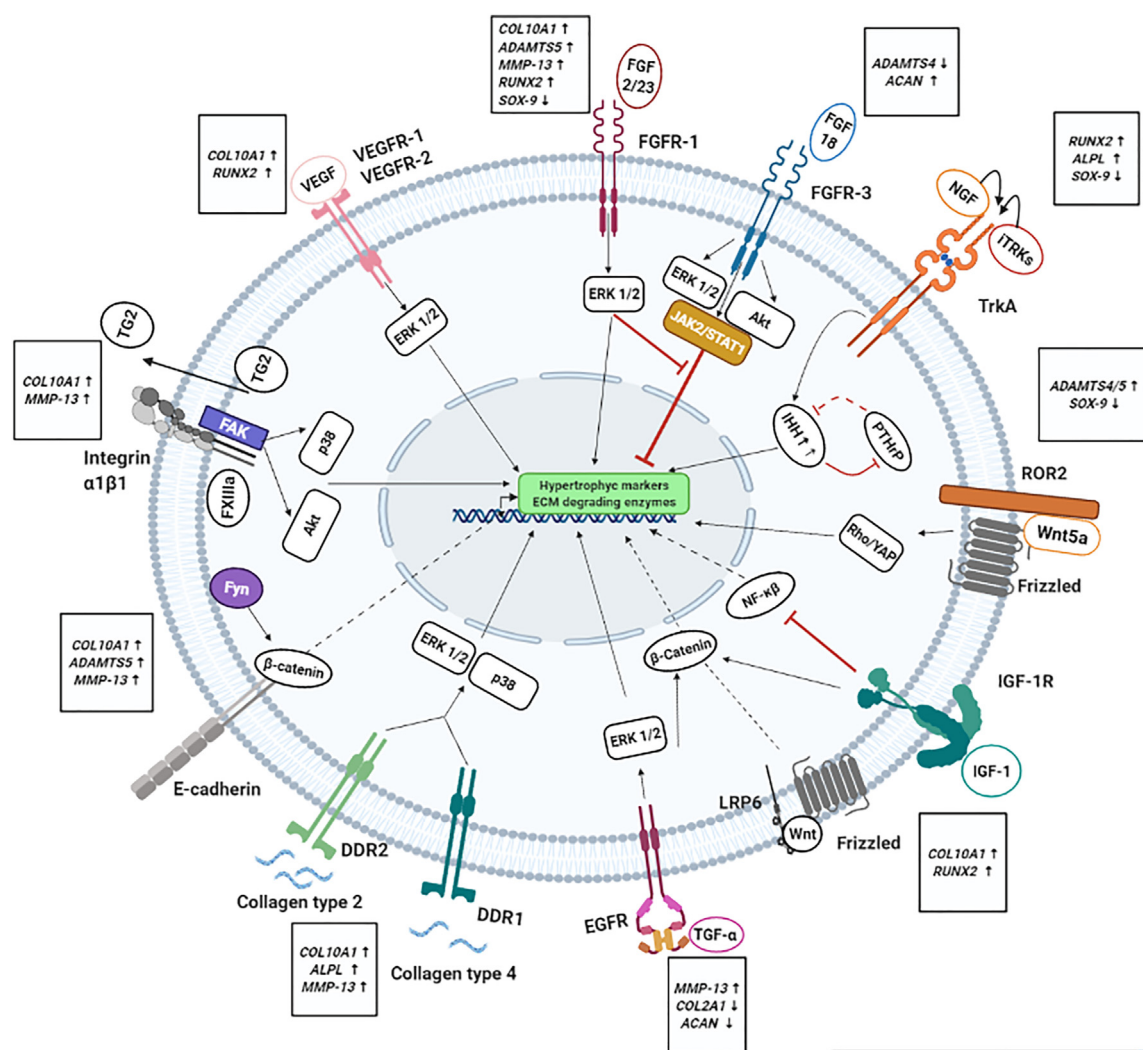
Janus kinases (JAKs) family is constituted by four members, including JAK1, JAK2, JAK3 and Tyrosine kinase 2 (Tyk2), which are crucial components of various signal transduction pathways that control cellular survival and differentiation, among others. JAKs can be activated by either cytokines/interferons or growth factors. Upon the activation of JAKs by ligand-induced receptor oligomerization, these enzymes then phosphorylate the receptor that initiated the signal transduction and create docking sites for signaling molecules, especially members of the signal transducer and activator of transcription (STAT) family. Subsequently, the phosphorylated STATs form homo- or heterodimers that translocate into the nucleus to induce transcription of specific gene targets⁸⁸. One of the members of the JAK family that is well-known to promote cell proliferation, JAK2, has recently been found to be activated in OA chondrocytes⁸⁹. IL-1 β highly increases JAK2/STAT3 phosphorylation in rat chondrocytes *in vitro*. Namely, the inhibition of JAK2/STAT3 pathway by JAK2 inhibitors such as AG490, prevented *Mmp-13* overexpression and the degradation of type II collagen⁴⁵. Conversely, other studies

have found JAK2 to present a protective role in OA. In fact, JAK2/STAT3 inhibition by miR-375 resulted in upregulation of *Adams5* and *Mmp13* and downregulation of *Col2a1* and *Acan* in mouse chondrocytes⁴⁴. Moreover, JAK2/STAT1 signaling could lead to the inhibition of hypertrophy in rat chondrocytes via parathyroid hormone (PTH)/PTHrP receptor down-regulation upon FGFR3 activation⁴³. In summary, JAK2 has a pivotal role in chondrocyte hypertrophy.

Focal adhesion kinase

Focal adhesion kinase (FAK) is a non-receptor tyrosine kinase that belongs to the FAK family, which also includes proline-rich tyrosine kinase 2 (PYK2). FAK has been suggested to play an essential role in integrin-mediated signal transductions and is important for early stages of cell migration and angiogenesis⁹⁰. Activation of FAK occurs by disruption of an auto-inhibitory

molecular interaction between the amino terminal four-point-one ezrin radixin moesin (FERM) domain and the central kinase domain. Upon activation, FAK associates with Src family kinases to initiate several signaling pathways including PI3K/AKT and MAPK. A number of studies *in vitro* suggested a critical role of FAK in angiogenesis during cancer progression⁹¹. On the other hand, FAK was also found to be involved in the pathophysiology of OA. Specifically, FAK could induce chondrocyte hypertrophic differentiation in response to factor XIIIa (FXIIIa) and transglutaminase 2 (TG2). Both transglutaminases are known to undergo physiologic upregulation in OA cartilage. In particular, TG2 mobilization to the cell surface is mediated by FXIIIa, upon its interaction with $\alpha 1\beta 1$ integrin and thus, results in the phosphorylation of FAK and p38 MAP kinase. The activation of these protein kinases then led to increased levels of *COL10A1* in bovine articular chondrocytes⁴². FAK may act as a positive regulator of chondrocyte hypertrophy since



Created in BioRender.com

Fig. 3

Overview of the receptor and non-receptor tyrosine kinases that have been associated with osteoarthritis and the pathways through which they regulate chondrocyte hypertrophy. In red, the inhibitory relations are depicted; the black dashed arrows indicate the translocation of the factors to the nucleus and the non-dashed arrows indicate the positive relations for each pathway. In white text-boxes there is written the genes that either are being upregulated or downregulated.

Osteoarthritis
and Cartilage

Drug	Target Tyrosine kinase	Target Disease	Side effects
Fedratinib	JAK2	Myelofibrosis	Anemia Diarrhea
Ruxolitinib	JAK2	Myelofibrosis	Fatigue Anemia Thrombocytopenia
Nintedanib	VEGFRs, FGFR1–3 and PDGFRs	Pulmonary fibrosis	Diarrhea

Table II Overview of FDA-approved tyrosine kinase inhibitors with potential interest for Osteoarthritis

Osteoarthritis
and Cartilage

FAK is able to activate PI3K, which in turns inhibits BAPX1 and that allows the transcription of *Col10a1* and *Mmp13*⁴¹. In spite of these findings further studies investigating the association between FAK and hypertrophy are required.

Fyn

Fyn is one of the nine members of the Src family, well-known proto-oncogenes that regulate cell growth and are frequently mutated in cancer⁹². Fyn is a tyrosine-specific phospho-transferase activated by autophosphorylation at Tyr416, upon stimulation of diverse transmembrane receptors including RTKs, G protein-coupled receptors, integrins and cytokine receptors⁹³. Thus, Fyn mediates multiple signal transduction pathways such as PKC δ /MAPK/NF- κ B, STAT3 or mTORC1, which regulate a large spectrum of biological processes involved in inflammation, fibrosis and cellular survival, respectively⁹⁴. Recently, Fyn has been found to accumulate in human OA cartilage, thereby suggesting a potential role of Fyn also in the pathogenesis of OA³. Concretely, Fyn deficiency in mice protected against age-related or trauma-induced cartilage degradation and the development of OA. Furthermore, they revealed that Fyn phosphorylation led to the increased expression of hypertrophic markers such as *COL10A1*, *MMP13* and *ADAMTS5* in mice and human chondrocytes, through the stabilization of β -catenin in a Wnt-independent manner. Given the close relation between Fyn and hypertrophy of articular chondrocytes³, more pre-clinical studies about Fyn role in OA pathophysiology would be of high interest.

Discussion

In recent years, there has been increasing interest in the roles of TKs in the pathophysiology of various diseases, since these are essential players in cellular signaling processes³. In this review it was highlighted that some members of the tyrosine kinase family, including FGFR-1, DDR1, DDR2, EGFR, VEGFR, TrkA, ROR2, FAK and Fyn, have been found to induce chondrocyte hypertrophy in articular cartilage (Fig. 3)^{2,8,39,41,66,95}. Thus, the use of inhibitors that impair the activity of these TKs could be considered as a potential treatment for OA. On this regard, repositioning drugs is an attractive alternative to lower development costs and shorten bench to bedside timeline. Three molecules that have been approved for clinical use for the treatment of Myelofibrosis and Pulmonary fibrosis (Table II) might be interesting for drug repurposing as disease-modifying osteoarthritic drugs. Nintedanib is an oral tyrosine kinase inhibitor with specificity against VEGFRs, FGFRs and PDGFRs. Fedratinib and Ruxolitinib, both orally bioavailable, selectively inhibit JAK2, however, considering the pivotal role of JAK2 in chondrocyte hypertrophy, caution should be taken. Of note, the adverse effects associated with these medications are often

related to systemic application and could be decreased by dose-scalation or by intra-articular injection. The TKs FGFR-3 and IGF-1 receptor may be playing a protective role in cartilage homeostasis, therefore, its inhibition could lead to hypertrophy in articular chondrocytes (Fig. 3)^{30,32}. Currently, some companies are working on agonist against FGFR-3 due to their potentiality to treat OA in an efficient manner.

Despite these findings, a limitation of this review is that only few studies exist for each of these proteins. Moreover, some of these studies used growth plate chondrocytes which may behave differently from articular chondrocytes⁹⁶. Hence, it would be interesting to further study the role of TKs in human articular chondrocytes. Furthermore, since tyrosine kinase inhibition could affect other joint structures, such as the synovium and the sub-chondral bone, the role of TK in these tissues should also be evaluated. To improve the specificity of the treatment, the use of non-competitive inhibitors is recommended since these molecules do not target the active site of the TKs and thereby have less promiscuity in binding⁹⁷. Since TKs do not only modulate chondrocyte hypertrophy but also play a role in inflammation and autophagy⁹⁸, TKs inhibitors could significantly help in elaborating treatments that can hinder the progression of OA.

In conclusion, articular cartilage homeostasis is highly susceptible by changes in signaling pathways regulated by TKs that, when dysregulated, lead to chondrocyte hypertrophy. Unrevealing the mechanisms by which TKs influence cartilage hypertrophy could further provide new pharmacological targets for the elaboration of more effective treatments.

Declaration of conflicting interests

The authors declared no conflicts of interest.

Acknowledgments

This work was performed within the framework of the Erasmus Postgraduate School Molecular Medicine and the European Union's Horizon 2020 Research and Innovation Programme, Marie Skłodowska-Curie grant agreement no. 721432 CarBon.

References

- Xu W, Xie Y, Wang Q, Wang X, Luo F, Zhou S, et al. A novel fibroblast growth factor receptor 1 inhibitor protects against cartilage degradation in a murine model of osteoarthritis. *Sci Rep* 2016;6:24042.
- Yan D, Chen D, Cool SM, van Wijnen AJ, Mikecz K, Murphy G, et al. Fibroblast growth factor receptor 1 is principally responsible for fibroblast growth factor 2-induced catabolic

- activities in human articular chondrocytes. *Arthritis Res Ther* 2011;13:R130.
3. Li K, Zhang Y, Zhang Y, Jiang W, Shen J, Xu S, *et al.* Tyrosine kinase Fyn promotes osteoarthritis by activating the beta-catenin pathway. *Ann Rheum Dis* 2018;77:935–43.
 4. Aigner T, Dietz U, Stoss H, von der Mark K. Differential expression of collagen types I, II, III, and X in human osteophytes. *Lab Invest* 1995;73:236–43.
 5. Yang L, Tsang KY, Tang HC, Chan D, Cheah KS. Hypertrophic chondrocytes can become osteoblasts and osteocytes in endochondral bone formation. *Proc Natl Acad Sci U S A* 2014;111:12097–102.
 6. Ji Q, Zheng Y, Zhang G, Hu Y, Fan X, Hou Y, *et al.* Single-cell RNA-seq analysis reveals the progression of human osteoarthritis. *Ann Rheum Dis* 2019;78:100–10.
 7. Chou CH, Jain V, Gibson J, Attarian DE, Haraden CA, Yohn CB, *et al.* Synovial cell cross-talk with cartilage plays a major role in the pathogenesis of osteoarthritis. *Sci Rep* 2020;10:10868.
 8. Jiang Y, Tuan RS. Role of NGF-TrkA signaling in calcification of articular chondrocytes. *Faseb J* 2019;33:10231–9.
 9. Blaney Davidson EN, Remst DF, Vitters EL, van Beuningen HM, Blom AB, Goumans MJ, *et al.* Increase in ALK1/ALK5 ratio as a cause for elevated MMP-13 expression in osteoarthritis in humans and mice. *J Immunol* 2009;182:7937–45.
 10. Day TF, Guo X, Garrett-Beal L, Yang Y. Wnt/beta-catenin signaling in mesenchymal progenitors controls osteoblast and chondrocyte differentiation during vertebrate skeletogenesis. *Dev Cell* 2005;8:739–50.
 11. Islam S, Kermode T, Sultana D, Moskowitz RW, Mukhtar H, Malemud CJ, *et al.* Expression profile of protein tyrosine kinase genes in human osteoarthritis chondrocytes. *Osteoarthritis Cartilage* 2001;9:684–93.
 12. Murthy RK, Loi S, Okines A, Paplomata E, Hamilton E, Hurvitz SA, *et al.* Tucatinib, trastuzumab, and capecitabine for HER2-positive metastatic breast cancer. *N Engl J Med* 2020;382:597–609.
 13. Solassol I, Pinguet F, Quantin X. FDA- and EMA-approved tyrosine kinase inhibitors in advanced EGFR-mutated non-small cell lung cancer: safety, tolerability, plasma concentration monitoring, and management. *Biomolecules* 2019;9.
 14. Daouti S, Latario B, Nagulapalli S, Buxton F, Uziel-Fusi S, Chirn GW, *et al.* Development of comprehensive functional genomic screens to identify novel mediators of osteoarthritis. *Osteoarthritis Cartilage* 2005;13:508–18.
 15. Claassen H, Schicht M, Brandt J, Reuse K, Schädlich R, Goldring MB, *et al.* C-28/12 and T/C-28a2 chondrocytes as well as human primary articular chondrocytes express sex hormone and insulin receptors—Useful cells in study of cartilage metabolism. *Ann Anat* 2011;193:23–9.
 16. Shakibaei M, Schulze-Tanzil G, Mobasheri A, Beichler T, Dressler J, Schwab W. Expression of the VEGF receptor-3 in osteoarthritic chondrocytes: stimulation by interleukin-1 beta and association with beta 1-integrins. *Histochem Cell Biol* 2003;120:235–41.
 17. Pfander D, Cramer T, Weseloh G, Pullig O, Schuppan D, Bauer M, *et al.* Hepatocyte growth factor in human osteoarthritic cartilage. *Osteoarthritis Cartilage* 1999;7:548–59.
 18. Nawachi K, Inoue M, Kubota S, Nishida T, Yosimichi G, Nakanishi T, *et al.* Tyrosine kinase-type receptor ErbB4 in chondrocytes: interaction with connective tissue growth factor and distribution in cartilage. *FEBS Lett* 2002;528:109–13.
 19. Kwan Tat S, Pelletier JP, Amiable N, Boileau C, Lavigne M, Martel-Pelletier J. Treatment with ephrin B2 positively impacts the abnormal metabolism of human osteoarthritic chondrocytes. *Arthritis Res Ther* 2009;11:R119.
 20. Bursell L, Woods A, James CG, Pala D, Leask A, Beier F. Src kinase inhibition promotes the chondrocyte phenotype. *Arthritis Res Ther* 2007;9:R105.
 21. Kjelgaard-Petersen CF, Sharma N, Kayed A, Karsdal MA, Mobasheri A, Hägglund P, *et al.* Tofacitinib and TPCA-1 exert chondroprotective effects on extracellular matrix turnover in bovine articular cartilage ex vivo. *Biochem Pharmacol* 2019;165:91–8.
 22. Chou HC, Chen CH, Chou LY, Cheng TL, Kang L, Chuang SC, *et al.* Discoidin domain receptors 1 inhibition alleviates osteoarthritis via enhancing autophagy. *Int J Mol Sci* 2020;21.
 23. Schminke B, Muhammad H, Bode C, Sadowski B, Gerter R, Gersdorff N, *et al.* A discoidin domain receptor 1 knock-out mouse as a novel model for osteoarthritis of the temporomandibular joint. *Cell Mol Life Sci* 2014;71:1081–96.
 24. Zhang S, Zhong Y, Li R, Wang W, Zeng L, Wang Z, *et al.* Experimental chondrocyte hypertrophy is promoted by the activation of discoidin domain receptor 2. *Mol Med Rep* 2014;10:1543–8.
 25. Sunk IG, Bobacz K, Hofstaetter JG, Amoyo L, Soleiman A, Smolen J, *et al.* Increased expression of discoidin domain receptor 2 is linked to the degree of cartilage damage in human knee joints: a potential role in osteoarthritis pathogenesis. *Arthritis Rheum* 2007;56:3685–92.
 26. Xu L, Peng H, Glasson S, Lee PL, Hu K, Ijiri K, *et al.* Increased expression of the collagen receptor discoidin domain receptor 2 in articular cartilage as a key event in the pathogenesis of osteoarthritis. *Arthritis Rheum* 2007;56:2663–73.
 27. Huang CY, Lin HJ, Chen HS, Cheng SY, Hsu HC, Tang CH. Thrombin promotes matrix metalloproteinase-13 expression through the PKC δ c-Src/EGFR/PI3K/Akt/AP-1 signaling pathway in human chondrocytes. *Mediat Inflamm* 2013;2013:326041.
 28. Weng T, Yi L, Huang J, Luo F, Wen X, Du X, *et al.* Genetic inhibition of fibroblast growth factor receptor 1 in knee cartilage attenuates the degeneration of articular cartilage in adult mice. *Arthritis Rheum* 2012;64:3982–92.
 29. Tang J, Su N, Zhou S, Xie Y, Huang J, Wen X, *et al.* Fibroblast growth factor receptor 3 inhibits osteoarthritis progression in the knee joints of adult mice. *Arthritis Rheum* 2016;68:2432–43.
 30. Valverde-Franco G, Binette JS, Li W, Wang H, Chai S, Laflamme F, *et al.* Defects in articular cartilage metabolism and early arthritis in fibroblast growth factor receptor 3 deficient mice. *Hum Mol Genet* 2006;15:1783–92.
 31. Davidson D, Blanc A, Filion D, Wang H, Plut P, Pfeffer G, *et al.* Fibroblast growth factor (FGF) 18 signals through FGF receptor 3 to promote chondrogenesis. *J Biol Chem* 2005;280:20509–15.
 32. Montaseri A, Busch F, Mobasheri A, Buhrmann C, Aldinger C, Rad JS, *et al.* IGF-1 and PDGF-bb suppress IL-1 β -induced cartilage degradation through down-regulation of NF- κ B signaling: involvement of Src/PI-3K/AKT pathway. *PloS One* 2011;6, e28663.
 33. Shakibaei M, Seifarth C, John T, Rahmzadeh M, Mobasheri A. Igf-I extends the chondrogenic potential of human articular chondrocytes *in vitro*: molecular association between Sox9 and Erk1/2. *Biochem Pharmacol* 2006;72:1382–95.
 34. Rosa SC, Rufino AT, Judas F, Tenreiro C, Lopes MC, Mendes AF. Expression and function of the insulin receptor in normal and osteoarthritic human chondrocytes: modulation of anabolic gene expression, glucose transport and GLUT-1 content by insulin. *Osteoarthritis Cartilage* 2011;19:719–27.
 35. Thorup AS, Strachan D, Caxaria S, Poulet B, Thomas BL, Eldridge SE, *et al.* ROR2 blockade as a therapy for osteoarthritis. *Sci Transl Med* 2020;12.

36. Krupka E, Jiang GL, Jan C. Efficacy and safety of intra-articular injection of tropomyosin receptor kinase A inhibitor in painful knee osteoarthritis: a randomized, double-blind and placebo-controlled study. *Osteoarthritis Cartilage* 2019;27:1599–607.
37. Watt FE, Blauwet MB, Fakhoury A, Jacobs H, Smulders R, Lane NE. Tropomyosin-related kinase A (TrkA) inhibition for the treatment of painful knee osteoarthritis: results from a randomized controlled phase 2a trial. *Osteoarthritis Cartilage* 2019;27:1590–8.
38. Shen P, Jiao Z, Zheng JS, Xu WF, Zhang SY, Qin A, et al. Injecting vascular endothelial growth factor into the temporomandibular joint induces osteoarthritis in mice. *Sci Rep* 2015;5:16244.
39. Zhang X, Crawford R, Xiao Y. Inhibition of vascular endothelial growth factor with shRNA in chondrocytes ameliorates osteoarthritis. *J Mol Med (Berl)* 2016;94:787–98.
40. Ishiguro N, Oyama S, Higashi R, Yanagida K. Efficacy, safety, and tolerability of ONO-4474, an orally available pan-tropomyosin receptor kinase inhibitor, in Japanese patients with moderate to severe osteoarthritis of the knee: a randomized, placebo-controlled, double-blind, parallel-group comparative study. *J Clin Pharmacol* 2020;60:28–36.
41. Cao Z, Dou C, Li J, Tang X, Xiang J, Zhao C, et al. Cordycepin inhibits chondrocyte hypertrophy of mesenchymal stem cells through PI3K/Baxp1 and Notch signaling pathway. *BMB Rep* 2016;49:548–53.
42. Johnson KA, Rose DM, Terkeltaub RA. Factor XIIIa mobilizes transglutaminase 2 to induce chondrocyte hypertrophic differentiation. *J Cell Sci* 2008;121:2256–64.
43. Li M, Seki Y, Freitas PH, Nagata M, Kojima T, Sultana S, et al. FGFR3 down-regulates PTH/PTHrP receptor gene expression by mediating JAK/STAT signaling in chondrocytic cell line. *J Electron Microscop* (Tokyo) 2010;59:227–36.
44. Zou LX, Yu L, Zhao XM, Liu J, Lu HG, Liu GW, et al. MiR-375 mediates chondrocyte metabolism and oxidative stress in osteoarthritis mouse models through the JAK2/STAT3 signaling pathway. *Cells Tissues Organs* 2019;208:13–24.
45. Yao ZZ, Hu AX, Liu XS. DUSP19 regulates IL-1 β -induced apoptosis and MMPs expression in rat chondrocytes through JAK2/STAT3 signaling pathway. *Biomed Pharmacother* 2017;96:1209–15.
46. Lemmon MA, Schlessinger J. Cell signaling by receptor tyrosine kinases. *Cell* 2010;141:1117–34.
47. Oladipupo SS, Smith C, Santeford A, Park C, Sene A, Wiley LA, et al. Endothelial cell FGF signaling is required for injury response but not for vascular homeostasis. *Proc Natl Acad Sci U S A* 2014;111:13379–84.
48. Xie Y, Su N, Yang J, Tan Q, Huang S, Jin M, et al. FGF/FGFR signaling in health and disease. *Signal Transduct Target Ther* 2020;5:181.
49. Wu YM, Su F, Kalyana-Sundaram S, Khazanov N, Ateeq B, Cao X, et al. Identification of targetable FGFR gene fusions in diverse cancers. *Canc Discov* 2013;3:636–47.
50. Brodie SG, Kitoh H, Lachman RS, Nolasco LM, Mekikian PB, Wilcox WR. Platyspondylic lethal skeletal dysplasia, San Diego type, is caused by FGFR3 mutations. *Am J Med Genet* 1999;84:476–80.
51. Bianchi A, Guibert M, Cailotto F, Gasser A, Presle N, Mainard D, et al. Fibroblast Growth Factor 23 drives MMP13 expression in human osteoarthritic chondrocytes in a Klotho-independent manner. *Osteoarthritis Cartilage* 2016;24:1961–9.
52. Zhang H, Wang H, Zeng C, Yan B, Ouyang J, Liu X, et al. mTORC1 activation downregulates FGFR3 and PTH/PTHrP receptor in articular chondrocytes to initiate osteoarthritis. *Osteoarthritis Cartilage* 2017;25:952–63.
53. Eckstein F, Kraines JL, Aydemir A, Wirth W, Maschek S, Hochberg MC. Intra-articular sprifermin reduces cartilage loss in addition to increasing cartilage gain independent of location in the femorotibial joint: post-hoc analysis of a randomised, placebo-controlled phase II clinical trial. *Ann Rheum Dis* 2020;79:525–8.
54. Vogel W, Gish GD, Alves F, Pawson T. The discoidin domain receptor tyrosine kinases are activated by collagen. *Mol Cell* 1997;1:13–23.
55. Labrador JP, Azcoitia V, Tuckermann J, Lin C, Olaso E, Mañes S, et al. The collagen receptor DDR2 regulates proliferation and its elimination leads to dwarfism. *EMBO Rep* 2001;2:446–52.
56. Shrivastava A, Radziejewski C, Campbell E, Kovac L, McGlynn M, Ryan TE, et al. An orphan receptor tyrosine kinase family whose members serve as nonintegrin collagen receptors. *Mol Cell* 1997;1:25–34.
57. Leitinger B, Steplewski A, Fertala A. The D2 period of collagen II contains a specific binding site for the human discoidin domain receptor, DDR2. *J Mol Biol* 2004;344:993–1003.
58. Leitinger B, Kwan AP. The discoidin domain receptor DDR2 is a receptor for type X collagen. *Matrix Biol* 2006;25:355–64.
59. Reel B, Korkmaz CG, Arun MZ, Yildirim G, Ogut D, Kaymak A, et al. The regulation of matrix metalloproteinase expression and the role of discoidin domain receptor 1/2 signalling in zoledronate-treated PC3 cells. *J Canc* 2015;6:1020–9.
60. Zhang Y, Su J, Yu J, Bu X, Ren T, Liu X, et al. An essential role of discoidin domain receptor 2 (DDR2) in osteoblast differentiation and chondrocyte maturation via modulation of Runx2 activation. *J Bone Miner Res* 2011;26:604–17.
61. Lin KL, Chou CH, Hsieh SC, Hwa SY, Lee MT, Wang FF. Transcriptional upregulation of DDR2 by ATF4 facilitates osteoblastic differentiation through p38 MAPK-mediated Runx2 activation. *J Bone Miner Res* 2010;25:2489–503.
62. Suh HN, Han HJ. Collagen I regulates the self-renewal of mouse embryonic stem cells through $\alpha 2\beta 1$ integrin- and DDR1-dependent Bmi-1. *J Cell Physiol* 2011;226:3422–32.
63. Lu KK, Trcka D, Bendeck MP. Collagen stimulates discoidin domain receptor 1-mediated migration of smooth muscle cells through Src. *Cardiovasc Pathol* 2011;20:71–6.
64. Xiao L, Liu C, Wang B, Fei W, Mu Y, Xu L, et al. Targeting discoidin domain receptor 2 for the development of disease-modifying osteoarthritis drugs. *Cartilage* 2019. 1947603519852401.
65. Xu L, Peng H, Wu D, Hu K, Goldring MB, Olsen BR, et al. Activation of the discoidin domain receptor 2 induces expression of matrix metalloproteinase 13 associated with osteoarthritis in mice. *J Biol Chem* 2005 Jan 7;280:548–55.
66. Xu L, Servais J, Polur I, Kim D, Lee PL, Chung K, et al. Attenuation of osteoarthritis progression by reduction of discoidin domain receptor 2 in mice. *Arthritis Rheum* 2010;62:2736–44.
67. Brown J, Delaine C, Zaccheo OJ, Siebold C, Gilbert RJ, van Boxel G, et al. Structure and functional analysis of the IGF-II/IGF2R interaction. *EMBO J* 2008;27:265–76.
68. Gao J, Chesebrough JW, Cartledge SA, Ricketts SA, Incognito L, Veldman-Jones M, et al. Dual IGF-I/II-neutralizing antibody MEDI-573 potently inhibits IGF signaling and tumor growth. *Canc Res* 2011;71:1029–40.
69. Hidalgo M, Gomez MT, Lewis N, Vukj JL, Taylor G, Hayburn JL, et al. A phase I study of MK-0646, a humanized monoclonal antibody against the insulin-like growth factor receptor type 1

- (IGF1R) in advanced solid tumor patients in a q2 wk schedule. *J Clin Oncol* 2008;26: 3520–3520.
70. Higano CS, Yu EY, Whiting SH, Gordon MS, LoRusso P, Fox F, et al. A phase I, first in man study of weekly IMC-A12, a fully human insulin like growth factor-I receptor IgG1 monoclonal antibody, in patients with advanced solid tumors. *J Clin Oncol* 2007;25: 3505–3505.
 71. Yin W, Park JJ, Loeser RF. Oxidative stress inhibits insulin-like growth factor-I induction of chondrocyte proteoglycan synthesis through differential regulation of phosphatidylinositol 3-Kinase-Akt and MEK-ERK MAPK signaling pathways. *J Biol Chem* 2009;284:31972–81.
 72. Wieduwilt MJ, Moasser MM. The epidermal growth factor receptor family: biology driving targeted therapeutics. *Cell Mol Life Sci* 2008;65:1566–84.
 73. Appleton CTG, Usmani SE, Bernier SM, Aigner T, Beier F. Transforming growth factor α suppression of articular chondrocyte phenotype and Sox9 expression in a rat model of osteoarthritis. *Arthritis Rheum* 2007;56:3693–705.
 74. Molina AM, Hutson TE, Nosov D, Tomczak P, Lipatov O, Sternberg CN, et al. Efficacy of tivozanib treatment after sorafenib in patients with advanced renal cell carcinoma: cross-over of a phase 3 study. *Eur J Canc* 2018;94:87–94.
 75. Kong X, Zhang Y, Liu C, Guo W, Li X, Su X, et al. Anti-angiogenic effect of triptolide in rheumatoid arthritis by targeting angiogenic cascade. *PLoS One* 2013;8, e77513.
 76. Richeldi L, du Bois RM, Raghu G, Azuma A, Brown KK, Costabel U, et al. Efficacy and safety of nintedanib in idiopathic pulmonary fibrosis. *N Engl J Med* 2014;370:2071–82.
 77. Jackson TL, Boyer D, Brown DM, Chaudhry N, Elman M, Liang C, et al. Oral tyrosine kinase inhibitor for neovascular age-related macular degeneration: a phase 1 dose-escalation study. *JAMA Ophthalmol* 2017;135:761–7.
 78. Koch S, Tugues S, Li X, Gualandi L, Claesson-Welsh L. Signal transduction by vascular endothelial growth factor receptors. *Biochem J* 2011;437:169–83.
 79. Walsh DA, McWilliams DF, Turley MJ, Dixon MR, Fransès RE, Mapp PI, et al. Angiogenesis and nerve growth factor at the osteochondral junction in rheumatoid arthritis and osteoarthritis. *Rheumatology (Oxford)* 2010;49:1852–61.
 80. Huang EJ, Reichardt LF. Trk receptors: roles in neuronal signal transduction. *Annu Rev Biochem* 2003;72:609–42.
 81. Iannone F, De Bari C, Dell'Accio F, Covelli M, Patella V, Lo Bianco G, et al. Increased expression of nerve growth factor (NGF) and high affinity NGF receptor (p140 TrkA) in human osteoarthritic chondrocytes. *Rheumatology* 2002;41:1413–8.
 82. Driscoll C, Chanalaris A, Knights C, Ismail H, Sacitharan PK, Gentry C, et al. Nociceptive sensitizers are regulated in damaged joint tissues, including articular cartilage, when osteoarthritic mice display pain behavior. *Arthritis Rheum* 2016;68:857–67.
 83. Masiakowski P, Carroll RD. A novel family of cell surface receptors with tyrosine kinase-like domain. *J Biol Chem* 1992;267:26181–90.
 84. Yu J, Chen L, Cui B, Widhopf 2nd GF, Shen Z, Wu R, et al. Wnt5a induces ROR1/ROR2 heterooligomerization to enhance leukemia chemotaxis and proliferation. *J Clin Invest* 2016;126:585–98.
 85. Chien HP, Ueng SH, Chen SC, Chang YS, Lin YC, Lo YF, et al. Expression of ROR1 has prognostic significance in triple negative breast cancer. *Virchows Arch* 2016;468:589–95.
 86. Guo M, Ma G, Zhang X, Tang W, Shi J, Wang Q, et al. ROR2 knockdown suppresses breast cancer growth through PI3K/ATK signaling. *Aging (Albany NY)* 2020;12:13115–27.
 87. Gocsek E, Moulas AN, Studzinski GP. Non-receptor protein tyrosine kinases signaling pathways in normal and cancer cells. *Crit Rev Clin Lab Sci* 2014;51:125–37.
 88. Berry DC, Jin H, Majumdar A, Noy N. Signaling by vitamin A and retinol-binding protein regulates gene expression to inhibit insulin responses. *Proc Natl Acad Sci U S A* 2011;108:4340–5.
 89. Brooks AJ, Dai W, O'Mara ML, Abankwa D, Chhabra Y, Pelekanos RA, et al. Mechanism of activation of protein kinase JAK2 by the growth hormone receptor. *Science* 2014;344:1249783.
 90. Polte TR, Naftilan AJ, Hanks SK. Focal adhesion kinase is abundant in developing blood vessels and elevation of its phosphotyrosine content in vascular smooth muscle cells is a rapid response to angiotensin II. *J Cell Biochem* 1994;55:106–19.
 91. Shi Q, Hjelmeland AB, Keir ST, Song L, Wickman S, Jackson D, et al. A novel low-molecular weight inhibitor of focal adhesion kinase, TAE226, inhibits glioma growth. *Mol Carcinog* 2007;46:488–96.
 92. Parsons SJ, Parsons JT. Src family kinases, key regulators of signal transduction. *Oncogene* 2004;23:7906–9.
 93. Jelić D, Mildner B, Kostrun S, Nujić K, Verbanac D, Culić O, et al. Homology modeling of human Fyn kinase structure: discovery of rosmarinic acid as a new Fyn kinase inhibitor and in silico study of its possible binding modes. *J Med Chem* 2007;50:1090–100.
 94. Seo HY, Jeon JH, Jung YA, Jung GS, Lee EJ, Choi YK, et al. Fyn deficiency attenuates renal fibrosis by inhibition of phospho-STAT3. *Kidney Int* 2016;90:1285–97.
 95. Li XS, Chen H, Zhen P, Li SS, Zhou SH, Tian Q, et al. JAK2/STAT3 signal pathway mediating curcumin in cartilage cell metabolism of osteoarthritis. *Zhong Guo Gu Shang* 2016;29:1104–9.
 96. Wang L, Shao YY, Ballock RT. Thyroid hormone-mediated growth and differentiation of growth plate chondrocytes involves IGF-1 modulation of beta-catenin signaling. *J Bone Miner Res* 2010;25:1138–46.
 97. Kirkland LO, McInnes C. Non-ATP competitive protein kinase inhibitors as anti-tumor therapeutics. *Biochem Pharmacol* 2009;77:1561–71.
 98. Domigan CK, Warren CM, Antanesian V, Happel K, Ziyad S, Lee S, et al. Autocrine VEGF maintains endothelial survival through regulation of metabolism and autophagy. *J Cell Sci* 2015;128:2236–48.
 99. Ferrao Blanco Mauricio, et al. Effect of inflammatory signaling on human articular chondrocyte hypertrophy: potential involvement of tissue repair macrophages. *Cartilage* 2021, <https://doi.org/10.1177/19476035211021907>.