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MINIREVIEW – Biotechnology &amp; Synthetic Biology

# Multiplex genome editing of microorganisms using CRISPR-Cas

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**One sentence summary:** Current status of multiplex genome editing in bacteria and eukaryotic microorganisms using CRISPR-Cas tools.

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## ABSTRACT

Microbial production of chemical compounds often requires highly engineered microbial cell factories. During the last years, CRISPR-Cas nucleases have been repurposed as powerful tools for genome editing. Here, we briefly review the most frequently used CRISPR-Cas tools and describe some of their applications. We describe the progress made with respect to CRISPR-based multiplex genome editing of industrial bacteria and eukaryotic microorganisms. We also review the state of the art in terms of gene expression regulation using CRISPRi and CRISPRa. Finally, we summarize the pillars for efficient multiplexed genome editing and present our view on future developments and applications of CRISPR-Cas tools for multiplex genome editing.

**Keywords:** CRISPR-Cas; genome editing; multiplex; cell factories; Cas9; Cas12a

## INTRODUCTION

Industrial microbiology plays a key role in the transition towards a more sustainable industry to produce food and feed ingredients, bio-based materials, biofuels and direct synthesis of cosmetic and pharmaceutical compounds (Lee *et al.* 2019). Oil-based production processes are gradually being substituted by bio-based processes, in which genetically engineered microorganisms are generally crucial to achieve cost-effective productivities and yields (Hong and Nielsen 2012; Dai and Nielsen 2015). The implementation of CRISPR-Cas tools has revolutionized genome

editing and mitigated the investment in the metabolic engineering programs required to generate highly engineered microbial cell factories (Donohoue, Barrangou and May 2018; Choi *et al.* 2019).

CRISPR-Cas systems (Clustered Regularly Interspaced Short Palindromic Repeats and CRISPR-associated proteins) are bacterial and archaeal adaptive immune defence systems, which can be repurposed as versatile genetic editing or regulation tools in a broad range of organisms. The effector endonucleases of these systems are guided by short RNA molecules encoded

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by CRISPR arrays. Native CRISPR arrays consist of a succession of spacers originating from invader organisms separated by direct repeats (Mojica et al. 2005; Barrangou et al. 2007). Transcription of the CRISPR array results in a long precursor-crRNA transcript (pre-crRNA), that is subsequently being processed to short functional CRISPR RNA (crRNA) guides (Brouns et al. 2008). To date, six different types of CRISPR-Cas systems (I–VI) have been described that are divided into two major classes (Class 1 and Class 2) (Makarova and Koonin 2015; Makarova et al. 2015; Shmakov et al. 2015, 2017; Koonin, Makarova and Zhang 2017). This review focuses on the application of DNA-targeting class 2 CRISPR systems (included in types II and V), that all consist of large multi-domain effector proteins able to use crRNA guides to target complementary DNA. Recent reviews have covered applications of CRISPR-Cas editing in bacteria (Choi and Lee 2016), in *Streptomyces* (Alberti and Corre 2019), in filamentous fungi (Shi et al. 2017), in yeast (Stovicek, Holkenbrink and Borodina 2017; Raschmanová et al. 2018), in microalgae and cyanobacteria (Naduthodi, Barbosa and Van der Oost 2018), and in general industrial microorganisms (Ferreira, David and Nielsen 2018).

After an introduction of single target genome editing tools, we focus on the spectacular development of multiplexed genome editing by Cas9 (type II) and Cas12a (type V) in industrial microorganisms. Both bacterial and eukaryotic examples are described, although more attention is given to yeast and filamentous fungi, since the diversity of strategies using Cas endonucleases for genome editing applications is more extensive in this group of organisms.

### Single target genome editing and regulation

Since its establishment as a genome editing tool (Gasiunas et al. 2012; Jinek et al. 2012), Cas9 from *Streptococcus pyogenes* (SpCas9) has become the most widely used RNA-guided endonuclease for genome editing and transcription regulation purposes (Table 1). Expression of this type II Cas nuclease together with a guide RNA (gRNA) is sufficient for generating targeted blunt double-stranded breaks (DSBs). The gRNA bound by SpCas9 consists of two small RNA molecules: a CRISPR RNA (crRNA) and a transactivating CRISPR RNA (tracrRNA). To simplify gRNA expression, a synthetic chimeric construct named single guide RNA (sgRNA) can be synthesized by fusing the tracrRNA and the crRNA (Jinek et al. 2012). Targeting of complementary DNA sequences (protospacers) by the Cas9:gRNA complex requires a protospacer adjacent motif (PAM), in case of Cas9 positioned downstream of the target sequence (Deveau et al. 2008). Correct PAM identification and base-pairing will trigger cleavage of the non-target and target DNA strands by the RuvC and HNH nuclease domains, respectively (Gasiunas et al. 2012; Jinek et al. 2012) (Fig. 1).

The more recently characterized endonuclease Cas12a (formerly called Cpf1) (type V) can cleave dsDNA directed by a crRNA, hence without the requirement of a tracrRNA (Zetsche et al. 2015) (Table 1). Cas12a does not possess an HNH domain, and its RuvC domain has been demonstrated to cleave both the non-target and the target DNA strands (Swarts, Van der Oost and Jinek 2017; Swarts and Jinek 2018). Moreover, Cas12a is able to process its crRNA guide autonomously (Fonfara et al. 2016; Swarts, van der Oost and Jinek 2017; Zetsche et al. 2017), while Cas9 relies on the activity of an additional non-Cas, dsRNA (crRNA/tracrRNA) targeting ribonuclease (RNaseIII) (Deltcheva et al. 2011). Both Cas9 and Cas12a can use multiple crRNA guides for creating simultaneous DSBs at different target loci in the genome (Fig. 1). Recently, two distinct Cas12 subtypes (Cas12b,

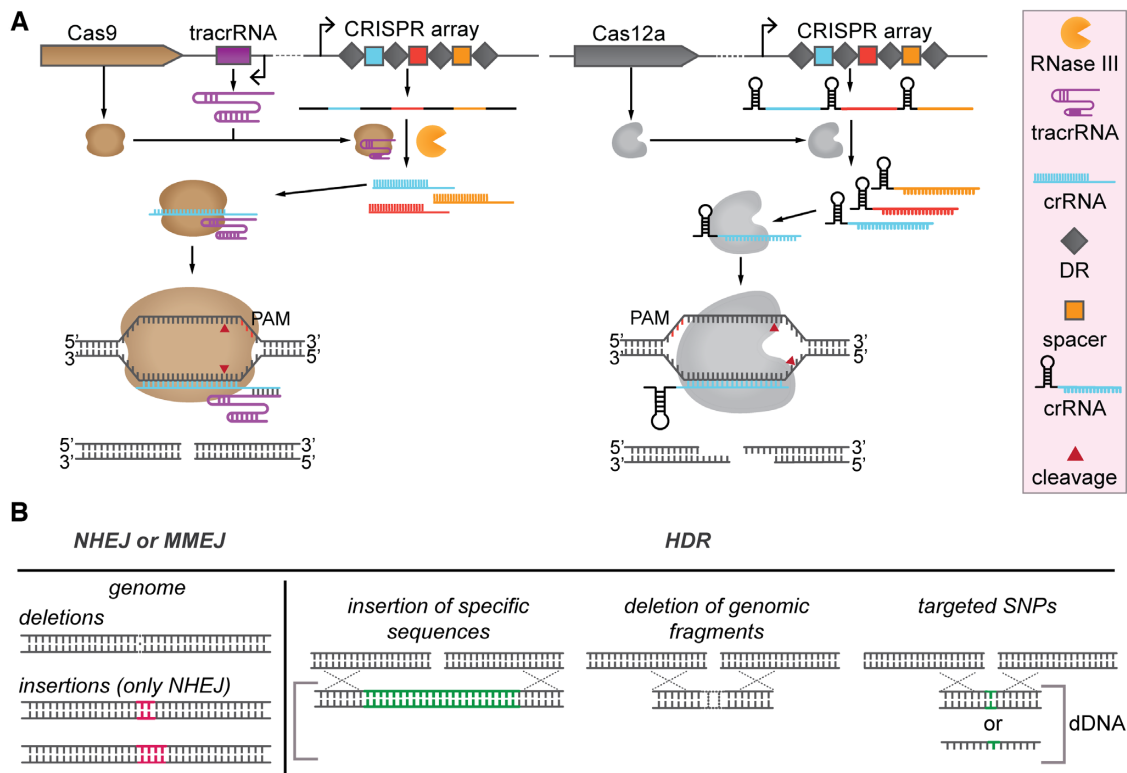
CasX/Cas12e) were shown to also edit genomes of bacteria and mammalian cells (Liu et al. 2019; Strecker et al. 2019).

Genomic DSBs can be repaired by homology-directed repair (HDR), non-homologous end joining repair (NHEJ) or alternative non-homologous end joining systems such as microhomology-mediated end joining (MMEJ) (Chayot et al. 2010; Sfeir and Symington 2015; Yao et al. 2017). The error-prone NHEJ repair system is often most prevalent in eukaryotes (Pawelczak et al. 2018) (Fig. 1), whereas it has been predicted to be encoded by only ~26% of publicly available prokaryotic genomes (Bowater and Doherty 2006; McGovern et al. 2016; Nayak and Metcalf 2017). The less studied alternative MMEJ repair system has been reported to also be present in bacteria and fungi (Sfeir and Symington 2015). This repair system has been proven to be active together with other repair mechanisms in the fungi *Aspergillus niger* and *Yarrowia lipolytica* (Shi et al. 2018) or in NHEJ-free bacteria (Chayot et al. 2010). HDR can be used in a targeted way: (i) to insert DNA fragments in targeted genomic locations; (ii) to delete small and large DNA fragments; or (iii) to introduce point mutations. HDR requires the introduction of a single or double-stranded DNA repair fragment into the cell, called donor DNA (dDNA), encoding the desired novel property or designed nucleotide change. To avoid targeting after the designed change, the recombinant sequence generally contains one or more silent mutations in the protospacer or PAM recognition sequence or partial deletion thereof. On the other hand, in organisms with highly active NHEJ or MMEJ repair systems, the introduction of non-specific insertions (only in case of NHEJ) and/or deletions (indels) in a certain target sequences can lead to gene disruption (Cong et al. 2013).

Inactive or deactivated versions of both Cas9 and Cas12a (named dCas9 and dCas12a) have been designed by substituting one or more of the catalytic amino acids in the nuclease domains (Gasiunas et al. 2012; Jinek et al. 2012; Zetsche et al. 2015; Swarts and Jinek 2018). These variants have been used to regulate gene expression in many organisms since they retain the target-binding ability (Berlec et al. 2018). By directing dCas9 or dCas12a to the promoter or coding sequence of a target gene, transcriptional repression (silencing) can be achieved by steric hindrance of the RNA polymerase and/or of transcription factors required for transcription of the target gene. This CRISPR interference (CRISPRi) technique has initially been established in *Escherichia coli*, resulting in significant transcriptional repression when targeting either the promoter or the non-template DNA strand of an open reading frame (Bikard et al. 2013; Qi et al. 2013). In eukaryotic microorganisms, gene repression is normally achieved by fusing repressor domains such as the mammalian transcriptional repressor domain Mxi1 or the Krüppel-associated box (KRAB domain) to the C-terminus of dCas9 or dCas12a (Jensen et al. 2017; Jensen 2018). Recently, native repression domains have been characterized in the yeast *Saccharomyces cerevisiae* with multiple Cas9 orthologs (Lian et al. 2017). This practice is more common in eukaryotic organisms since the use of only a dCas9:gRNA-complex seems not to be sufficient to significantly block transcription (Gilbert et al. 2013). Moreover, fusions of these deactivated variants to transcription activation domains are used to achieve gene activation (CRISPRa). In eukaryotes, VP16, VP64, Gal4<sup>AD</sup> or the synthetic VPR activator domains have been used successfully (Chavez et al. 2015; Jensen et al. 2017; Schwartz et al. 2018), while the omega ( $\omega$ ) subunit of the RNA polymerase has been used in bacteria (Bikard et al. 2013; La Russa and Qi 2015).

**Table 1.** Characteristics of the most commonly used Cas orthologues for genome editing.

Ortholog	Cas9		Cas12a	
	SpCas9	FnCas12a	AsCas12a	LbCas12a
Subtype	II-A		V-A	
Organism of origin	<i>Streptococcus pyogenes</i>	<i>Francisella novicida</i>	<i>Acidaminococcus</i> sp.	<i>Lachnospiraceae</i> bacterium
Nuclease domain	HNH, RuvC		RuvC	
tracrRNA	Yes		No	
PAM (5'-3')	NGG	TTTV	TTTV	TTTV
Size (amino acids)	1368	1302	1307	1228
RNA processing	No/RNaseIII	Yes/WED III	Yes/WED III	Yes/WED III
Minimum guide length (mature)	~100 nt		~44 nt	
Reference(s)	(Deltcheva et al. 2011)		(Zetsche, Heidenreich and Mohanraju et al. 2017)	



**Figure 1.** (A) Cas9 and Cas12a expression and cleavage schemes. Left panel: Cas9 requires tracrRNA transcription and RNase III expression for CRISPR array transcript processing. Cas9 forms a complex with crRNA and tracrRNA and cleaves target DNA generating blunt ends. Right panel: Cas12a processes its own CRISPR array transcript to obtain individual crRNAs without the requirement of any tracrRNA or RNase III co-expression. Cas12a stays in complex with crRNA and cleaves target DNA generating staggered ends. (B) Double strand break (DSB) repair mechanisms. DSBs can be repaired via non-homologous end joining (NHEJ), alternative non-homologous end joining repair pathways such as microhomology-mediated end joining (MMEJ), or via homologous direct recombination. NHEJ and MMEJ repair pathways can lead to the incorporation of deletions or insertions (only in case of NHEJ) in the targeted region. HDR is combined with the supplementation of donor DNA (dDNA), which can be double stranded or single stranded. dDNA can be used for insertion of long DNA sequences, deletion of genomic fragments, or introduction of single point mutations (SNPs).

### Multiplex genome editing

Editing of multiple loci is often required to introduce multiple heterologous genes and to fine-tune metabolic networks of microbial cell factories. In the pre-CRISPR era, iterative rounds of genome editing making use of selection markers were necessary to build strains expressing multiple-gene expression pathways.

Establishment of marker-free CRISPR-Cas tools brought powerful nuclease-mediated multiplex genome engineering capabilities, considerably saving time and resources in strain construction programs. The multiplexing capabilities of CRISPR-Cas systems as genome editing tools have been widely exploited with

Cas9 and more recently with Cas12a. One of the first microbial applications demonstrating the multiplexing capabilities of Cas9 was performed with the native Cas9 system of *Streptococcus pneumoniae* and two spacers expressed from a synthetic array integrated into the genome (Jiang et al. 2013). After this proof of principle, multiple studies explored the multiplexing capabilities of the endonuclease Cas9 in mammalian cells (Cong et al. 2013; Mali et al. 2013), as well as in industrially relevant prokaryotic and eukaryotic microorganisms (Table 2).

The crRNA processing activity of the recently characterized Cas12a increases the simplicity of multiplexing. By expressing a single CRISPR array containing multiple spacers under the transcriptional regulation of a single promoter and terminator, multiple loci can be targeted simultaneously (Zetsche et al. 2017). Therefore, there is no need of supplying multiple targeting expression constructs. Recent studies have demonstrated the multiplex editing potential of the Cas12a endonuclease in a wide range of microorganisms (Table 2).

Aneuploidy and polyploidy are common conditions among eukaryotic industrial microorganisms. The requirement for simultaneous targeting of multiple alleles in non-haploid strain results in a decrease of the CRISPR editing efficiencies (Mertens et al. 2019). The term 'cis-multiplexing' is used when targeting a single genomic locus found multiple times across the genome of a non-haploid organism. 'Trans-multiplexing' refers to the simultaneous introduction of modifications in multiple genes that occur in more than one copy in the genome (Ryan et al. 2014). Several examples of both cis- and trans- multiplexing have recently been described (Table 2).

### gRNA expression systems for efficient multiplex genome editing

In most reported cases of genome editing of industrial microorganisms, and in all bacterial examples, gRNAs are generally expressed from multicopy plasmids that, after successful editing, can be removed through counter selection (Bao et al. 2015; Si et al. 2017) or growth on non-selective medium (Mans et al. 2015; Wijsman et al. 2019). In case of multiplex genome editing, two strategies have been described for the delivery of the different gRNAs: (i) multiple gRNA expression cassettes transcribing a sgRNA molecule from one or more plasmids (Fig. 2); (ii) polycistronic expression of gRNAs, either inspired by native CRISPR systems or by synthetically designed ones (Fig. 3). Individual gRNAs can also be *in vitro* transcribed and supplied directly to filamentous fungi or microalgae (Table 2).

#### Multiplexing using multiple single gRNA expression cassettes

The initial attempts of multiplex genome editing using SpCas9 relied on combined expression of several individual gRNA expression cassettes. In bacteria, different strength natural or synthetic promoters (constitutive or inducible) are routinely used for gRNA expression (Table 2). In eukaryotes, efficient gRNA expression might be controlled by an RNA polymerase-III-dependent promoter. In a cell that expresses a nuclear-localized SpCas9, this gRNA can direct the nuclease to its target. In addition, fusions of gRNAs and tRNA auto-splicing sequences (used as promoters) have also been demonstrated to yield multiple functional gRNAs in eukaryotes (Schwartz et al. 2016; Song et al. 2018). Alternatively, gRNAs flanked with self-cleaving ribozymes on both ends have been expressed from RNA polymerase II-dependent promoters to provide transcripts with modified ends and increase transcript stability in the nucleus (Gao and Zhao 2014; Nødvig et al. 2015; Weninger et al. 2016; Wong et al. 2017).

Moreover, expression of a bacterial T7 RNA polymerase in the yeasts *Y. lipolytica* and *Kluyveromyces lactis* has been used for guide expression (Morse et al. 2018).

In bacteria, the maximal number of reported simultaneous editing events using SpCas9 varies from organism to organism (Table 2). Expression of multiple gRNAs from multiple expression cassettes is done from plasmid-borne or genome-integrated constructs. Most reported examples of multiplex genome editing using gRNAs expressed from multiple expression cassettes have been performed in *E. coli* (Jiang et al. 2015; Li et al. 2015; Ronda et al. 2016), with a maximum of four genes targeted simultaneously and an editing efficiency of ~30% (Feng et al. 2018). The authors of the study developed a CRISPR multiplex genome editing technique that uncouples transformation and editing. This separation is achieved by inducing Cas12a expression only after transformation and seems to be key, together with recombineering, for increased editing efficiencies (Reisch and Prather 2015; Feng et al. 2018).

In eukaryotes, the combination of several gRNA expression modules in a single amplicon or spread over several co-transformed plasmids resulted in successful editing in several organisms such as *S. cerevisiae* (Horwitz et al. 2015; Jakočiūnas et al. 2015; Lee et al. 2015; Mans et al. 2015; Ronda et al. 2015; Generoso et al. 2016; Deaner, Holzman and Alper 2018), *Komagataella phaffi* (Weninger et al. 2016), the xylose-utilizing yeast *Scheffersomyces stipitis* (Cao et al. 2018), the enzyme producer *Myceliophthora thermophila* (Liu et al. 2017), *K. lactis* (Horwitz et al. 2015), *Fusarium fujikuroi* (Shi et al. 2019), the methylotrophic yeast *Ogataea polymorpha* (Wang et al. 2018a) and the oleaginous yeast *Y. lipolytica* (Gao et al. 2016) (Table 2). In many organisms, high editing efficiencies allow for straightforward screening and selection of the desired mutant without introducing selectable markers in their genome. However, the reported efficiencies for targeting two or more sites simultaneously in eukaryotes vary significantly from one organism to another (i.e. from 2 to 100%). Using SpCas9, nine editing events in *S. cerevisiae* is the highest number of simultaneous modifications reported in microbes to date (Table 2) (Wijsman et al. 2019).

In some filamentous fungi, the gRNA molecules can be synthesized *in vitro* and co-transformed with Cas9-encoding plasmids. In microalgae, *in vitro* synthesized gRNAs can be delivered together with *in vitro* produced Cas proteins as ribonucleoprotein complexes (RNPs) as well (Liu et al. 2015; Pohl et al. 2016; Shi et al. 2017; Naduthodi, Barbosa and van der Oost 2018). Simultaneous double editing has been achieved by following this strategy in *Penicillium chrysogenum* (Pohl et al. 2016 2018), in *Phaeodactylum tricorutum* (Serif et al. 2018) and in the rice blast fungus *Magnaporthe oryzae* (Foster et al. 2018). Although transformation of *in vitro* synthesized gRNA simplifies gRNA cloning work and avoids the requirement of identifying effective RNA polymerase III promoters in eukaryotes, efficient selection of transformants may still require chromosomal integration of selectable markers located in the dDNA molecules (Liu et al. 2015; Pohl et al. 2016). These markers can be removed from the genome by using counter-selection markers or Cre-recombinase-based approaches.

Finally, although SpCas9 has been successfully implemented in microalgae, we found few examples for the use thereof for multiplexed genome editing (Behler et al. 2018; Naduthodi, Barbosa and van der Oost 2018). Recently, a triple knockout strain of the microalgae *P. tricorutum* was generated in a single step transformation using six different Cas9-based RNP complexes (Serif et al. 2018).

Table 2. Multiplexed genome editing events in industrial microorganisms using CRISPR-Cas systems.

Species [strain(poidy)]	Cas nuclease tool (expression), Plasmid (replication origin)/genome integrated	Strategy for multiplexed gRNA expression/delivery (expression), plasmid (replication origin)/genome integrated	Type of donor DNA: HFfs; amount/concentration	Type of modification: Number of target, editing efficiency	Reference
<b>PROKARYOTES</b>					
<i>Escherichia coli</i> [MG1655]	SpCas9 (inducible), Plasmid expression (repA101 ori)	Several sgRNA expression cassettes (constitutive), Plasmid expression	Circular dsDNA (pMB1 ori): ~300 bp	Knockouts: 2, 100%; 3, 88.3%; 4, > 30%	(Feng et al. 2018)
<i>Escherichia coli</i> [MG1655]	SpCas9 (constitutive), Plasmid expression (repA101 (Ts) ori)	Several sgRNA expression cassettes (constitutive), Plasmid expression (pMB1 ori)	Circular dsDNA (pMB1 ori): 250–550 bp	Knockouts: 2, 97% ± 4%; 3, 47% ± 8%	(Jiang et al. 2015)
<i>Escherichia coli</i> [MG1655]	SpCas9 (inducible), Plasmid expression (ColE1 ori)	Several sgRNA expression cassettes (inducible), Plasmid expression (pMB1 ori)	Linear ssDNA: 70 bp; 5 pmol	Short insertions: 2, ~70%	(Ronda et al. 2016)
<i>Escherichia coli</i> [MG1655]	SpCas9 (constitutive), Plasmid expression (p15A ori)	Several sgRNA expression cassettes (constitutive), Plasmid expression (ColE1 ori)	Linear ssDNA: ~89 bp; 50 pmol	Point mutations: 2, 83%; 3, 23%	(Li et al. 2015)
<i>Streptococcus pneumoniae</i> [crR6c]	SpCas9 (constitutive), Genome integrated	Native-like CRISPR array (constitutive), Genome integrated	Linear dsDNA: not mentioned; 0.7 ng/μl to 2.5 μg/μl	Deletions: 2, 75%	(Jiang et al. 2013)
<i>Streptomyces lividans</i>	SpCas9 (constitutive), Plasmid expression (pSG5rep)	Several sgRNA expression cassettes (constitutive), Plasmid expression (pSG5rep)	Circular dsDNA (oriT): 1 kB	Short deletions (20–34 bp): 2, 100% (4/4)	(Cobb, Wang and Zhao 2015)
<i>Streptomyces coelicolor</i> [M145]	SpCas9 (constitutive), Plasmid expression (pSG5rep)	Several sgRNA expression cassettes (constitutive), Plasmid expression (pSG5rep)	Circular dsDNA (pSG5): ~1 kB	Deletions (768–1053 bp): 2, 29–54%	(Huang et al. 2015)
<i>Escherichia coli</i> [MG1655]	FxCas12a (inducible), Plasmid expression (repA 101)	Native-like CRISPR array (constitutive), Plasmid expression (pSC101 ori)	Circular dsDNA (oriE): 500 bp	Gene insertions: 3, ~20%	(Ao et al. 2018)
<i>Streptomyces coelicolor</i> [M145]	FxCas12a (constitutive), Plasmid expression (pSG5rep)	Native-like CRISPR array (constitutive), Plasmid expression (pSG5)	Circular dsDNA (pSG5): ~1 kB	Knockouts: 2, 75%	(Li et al. 2018)
<b>EUKARYOTES</b>					
<i>Saccharomyces cerevisiae</i> [CEN.PK113-7D (n), CEN.PK2-1c (n), CEN.PK122 (2n)]	SpCas9 (constitutive), Genome integrated	Several sgRNA expression cassettes (RNA pol III promoter, constitutive), Plasmid expression (multicopy)	Linear dsDNA: 60 bp; 12 pmols	Knockouts: 2, 100%; 6, 65%	(Mans et al. 2015)
<i>Saccharomyces cerevisiae</i> [CEN.PK2-1c (n)]	SpCas9 (constitutive), Genome integrated	Several sgRNA expression cassettes with homology flanks to a linearized plasmid backbone, Plasmid expression (multicopy)	Linear dsDNA: 500 bp; 0.6–1.54 pmols	Knockouts: 3, 64%	(Horwitz et al. 2015)

Table 2. Continued

Species [strain(poidy)]	Cas nuclease tool (expression), Plasmid (replication origin)/genome integrated	Strategy for multiplexed gRNA expression/delivery (expression), plasmid (replication origin)/genome integrated	Type of donor DNA: HF's; amount/concentration	Type of modification: Number of target, editing efficiency	Reference
<i>Saccharomyces cerevisiae</i> [CEN.PK113-5D (n)]	FnCpf1 (constitutive), Genome integrated	Native-like CRISPR array (RNA pol III promoter, constitutive) Plasmid expression (multicopy)	Linear dsDNA: 60 bp; 12 pmols	Knockouts: 2, 100%; 4, 85%	(Swiat et al. 2017)
<i>Saccharomyces cerevisiae</i> [CEN.PK113-7D (n), Ethanol Red (2n)]	SpCas9 (constitutive), Plasmid expression (multicopy)	Several sgRNA expression cassettes (RNA pol III promoter, constitutive), Plasmid expression (multicopy)	Linear ssDNA: 40 bp; 300 pmols	Knockouts: 2, 91–98%	(Generoso et al. 2016)
<i>Saccharomyces cerevisiae</i> [BY4741 (n)]	FnCpf1 (constitutive), Plasmid expression (centromeric)	Native-like CRISPR array (RNA pol III promoter, constitutive), Plasmid expression (multicopy)	Linear dsDNA: 50 bp	Multi-gene integrations: 2, 52%; 3, 43%	(Li, Wang and Wei 2018)
<i>Saccharomyces cerevisiae</i> [204 508; ATCC (mated) (2n)]	SpCas9 (constitutive), Plasmid expression (multicopy)	Synthetic array of ribozyme-flanked sgRNA (RNA pol III promoter, constitutive), Plasmid expression (multicopy)	Linear dsDNA: 50 bp; 44.94 pmol	Deletions: 2, 43%; 3, 19%	(Ryan et al. 2014)
<i>Saccharomyces cerevisiae</i> [CEN.PK113-7D (n)]	LbCpf1, AsCpf1 or FnCpf1 (constitutive), Plasmid expression (centromeric)	Native-like crRNA-array with homology flanks to a linearized plasmid backbone (RNA pol III promoter, constitutive), Plasmid expression (multicopy)	Linear dsDNA: 50 bp; 90–120 fmols	Multi-gene integrations: 3, 91%	(Verwaal et al. 2018)
<i>Saccharomyces cerevisiae</i> [BY4741 (n), CEN.PK2-1c (n)]	ispCas9 (constitutive), Plasmid expression (multicopy)	Native-like crRNA-array (RNA pol III promoter, constitutive), separate expression of tracrRNA, Plasmid expression (multicopy)	Circular dsDNA, at 5' of each spacer sequence (multicopy): 50 bp; 142 fmols	Knockouts: 3, 27–87%	(Bao et al. 2015)
<i>Saccharomyces cerevisiae</i> [CEN.PK113-7D (n)]	SpCas9 (constitutive), Genome integrated	Synthetic crRNA-array (RNA pol III promoter), separate expression of PaCsy4, Plasmid expression (multicopy)	Linear dsDNA: 60 bp; 12 pmols	Knockouts: 2, 100%; 4, 96%	(Ferreira, et al. 2018)
<i>Saccharomyces cerevisiae</i> [BY4741 (n)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Several HDV ribozyme-sgRNA expression cassettes (RNA pol III promoter, constitutive), Plasmid expression (centromeric)	Linear dsDNA, barcoded: 60 bp; 55 pmols	Knockouts: 2, 65–87.5%; 3, 57.5–75%; 4, 27.5–15%	(Lee et al. 2015)
<i>Saccharomyces cerevisiae</i> [CEN.PK2-1C (n)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Several sgRNA expression cassettes (RNA pol III promoter, constitutive), Plasmid expression (multicopy)	Linear dsDNA: 500 bp; 700 fmols	Multi-gene integrations: 3, 84%	(Ronda et al. 2015)

Table 2. Continued

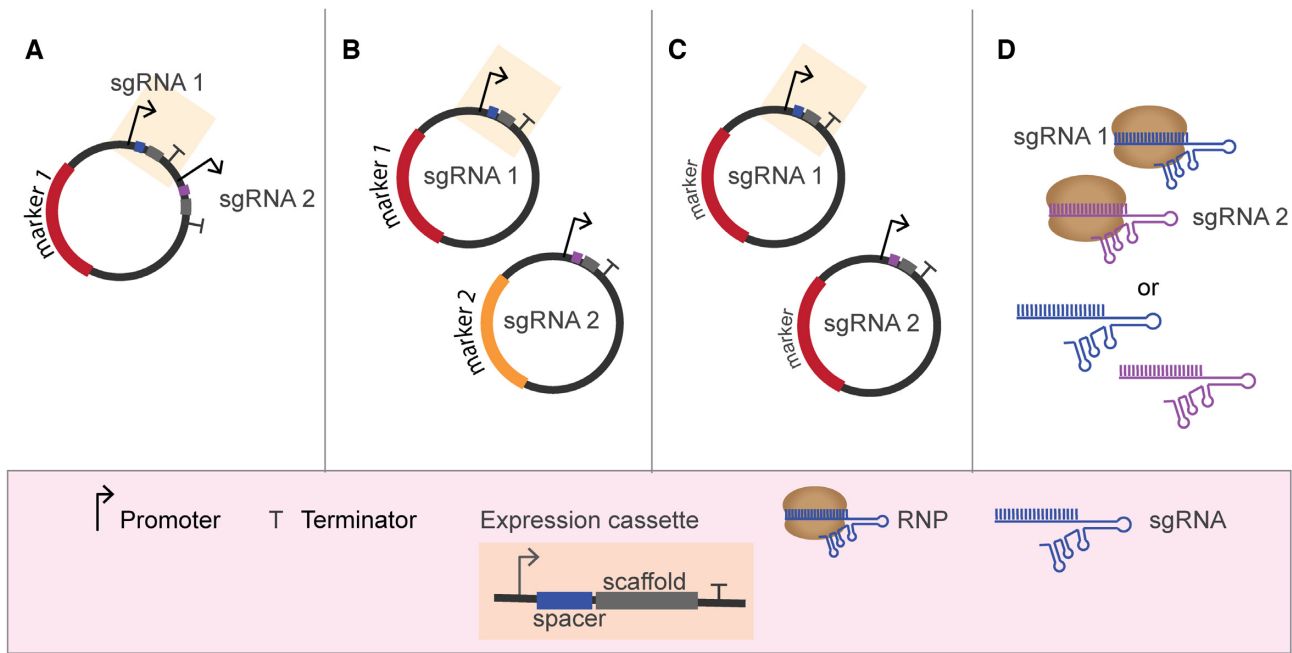
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<i>Saccharomyces cerevisiae</i> [CEN.PK111-27B (n)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Several sgRNA expression cassettes (RNA pol III promoter, constitutive), Plasmid expression (multicopy)	Linear dsDNA: 50 bp; 4 pmols	Multi-gene integrations: 2, 58%; 3, 30.6%	(Jakočiūnas et al. 2015)
<i>Saccharomyces cerevisiae</i> [CEN.PK2-1C]	SpCas9 (constitutive), Genome integrated	Several sgRNA expression cassettes (RNA pol III, constitutive), some target more than one site, Plasmid expression (multicopy)	Linear dsDNA: 60 bp; 26.96 pmol	Deletions: 9, 50% (only 2 transformants on plate)	(Wijisman et al. 2019)
<i>Saccharomyces cerevisiae</i> [CEN.PK 113-5D]	SpCas9 (constitutive), Multicopy plasmid	Synthetic crRNA-array (with one RNA pol III promoter for the expression of four gRNAs), gRNAs between trRNA <sup>gly</sup> sequences, Plasmid expression (multicopy)	Linear dsDNA: 50 bp; 266.9 pmol	Deletions (8 bp): 8, 86.7%	(Zhang et al. 2019)
<i>Ogataea parapolymorpha</i> [CBS 11 895 (n)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Synthetic array of ribozyme-flanked sgRNA (RNA pol II promoter, constitutive), Plasmid expression (centromeric)	Linear dsDNA: 480 bp, 1.6 pmols	Knockouts: 2, 2-5%	(Jurgens et al. 2018)
<i>Ogataea polymorpha</i> [CGMCC7.89 (n)]	iSpCas9 (constitutive), Genome integrated	Several sgRNA expression cassettes expressed (RNA pol III promoter, constitutive), Genome integrated	Linear dsDNA: 1 kb, 1.7 pmols	Multi-gene integrations: 3, 30.56 ± 2.40%	(Wang et al. 2018a)
<i>Yarrowia lipolytica</i> [ATCC 201 249 (n), ATCC MYA-2613 (n)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Synthetic array of ribozyme-flanked sgRNAs (RNA pol II promoter, constitutive), Plasmid expression (centromeric)	On multicopy plasmid: ~450bp	Knockouts: 2, 36.7 ± 8.5%; 3, 19.3 ± 9.2%	(Gao et al. 2016)
<i>Penicillium chrysogenum</i> [DS68530]	SpCas9 (transient), Delivered as a RNP	<i>In vitro</i> synthesized sgRNA in RNP, protoplast-mediated transformation, Transient expression	Linear dsDNA: ≥ 1 kb, 1-11 µg	Cassette integration: 2, 50%	(Pohl et al. 2016)
<i>Aspergillus nidulans</i> [IBT27263 (n)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Synthetic array of tRNA-flanked sgRNAs (RNA pol III promoter, constitutive), Plasmid expression (centromeric)	Linear ssDNA: 45 bp, 1 µmol	Multi-purpose: 3, 90%	(Nodvig et al. 2018)
<i>Trichoderma reesei</i> [ATCC 13 631, (n); ATCC 56 765 (n)]	SpCas9 (inductible), Genome integrated	<i>In vitro</i> synthesized sgRNA delivery by protoplasts transformation, Transient expression	Linear dsDNA: 200 bp, 296 pmols	Knockouts: 2, 16-45%; 3, 4.2%	(Liu et al. 2015)
<i>Scheffersomyces stipitidis</i> [UC7, (n)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Several sgRNA expression cassettes (RNA pol III promoter, constitutive), Plasmid expression (centromeric)	Linear dsDNA: 500 bp	Knockouts: 2, 40%	(Cao et al. 2018)



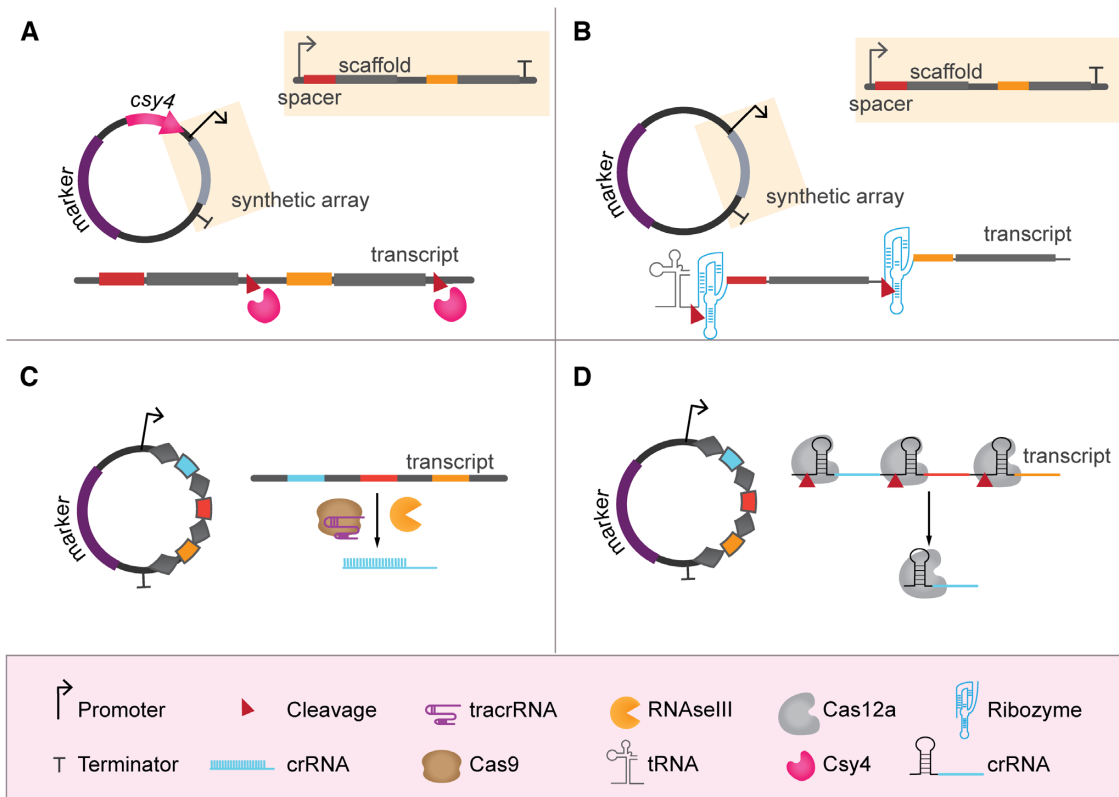
Table 2. Continued

Specie [strain(poidy)]	Cas nuclease tool (expression), Plasmid (replication origin)/genome integrated	Strategy for multiplexed gRNA expression/delivery (expression), plasmid (replication origin)/genome integrated	Type of donor DNA: HF's; amount/concentration	Type of modification: Number of target, editing efficiency	Reference
<i>Kluyveromyces lactis</i> [ATCC8585 (n)]	SpCas9 (constitutive), Genome integrated	Several sgRNA expression cassettes with homology flanks to a linearized plasmid backbone. Plasmid expression (multicopy)	Linear dsDNA: 1 kb, 0.6–1.54 pmols	Multi-gene integration: 3, 2.1%	(Horwitz et al. 2015)
<i>Myceliophthora thermophila</i> [ATCC 42 464, (n)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Several sgRNA expression cassettes (RNA pol III promoter, constitutive), Transient expression	Linear dsDNA: 600 bp; ~12pmols	Knockouts: 2, 61–70%; 3, 30%; 4, 22%	(Liu et al. 2017)
<i>Saccharomyces pastorianus</i> [CBS1483, (n#)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Synthetic array of ribozyme-flanked sgRNA (RNA pol II promoter, constitutive), Plasmid expression (centromeric)	Linear dsDNA: 60 bp; 12pmols	Knockouts: 2, 100%	(Gorter de Vries et al. 2017)
<i>Komagataella phaffii</i> [CBS7435 (n)]	SpCas9 (constitutive), Plasmid expression (centromeric)	Several sgRNA (ribozyme-flanked) expression cassettes (RNA pol II promoter, constitutive), Plasmid expression (centromeric)	Linear dsDNA: 1 kb; ~400–770fmols	Knockouts: 2, 69 ± 13%	(Weninger et al. 2016)
<i>Phaeodactylum tricornutum</i> (2n)	SpCas9 (transient), Delivered as a RNP	<i>In vitro</i> synthesized sgRNA in RNP, biolytic delivery, Transient expression	–	Knockouts: 2, 65–100%; 3, 15.4%	(Serif et al. 2018)
<i>Magnaporthe oryzae</i> (2n)	SpCas9 (transient), Delivered as a RNP	<i>In vitro</i> synthesized sgRNA in RNP, protoplast-mediated transformation, Transient expression	Linear dsDNA: ~40 bp	Knockouts by SNP: 2, 3.4–12.3%	(Foster et al. 2018)
<i>Fusarium fujikuroi</i> [NJtech 02, CCTCC M2015614]	SpCas9 (constitutive), Plasmid expression (centromeric)	Several sgRNA expression cassettes (RNA pol III promoter, constitutive), Plasmid expression (centromeric)	–	Knockouts by disruption: 2, 20.8%; 3, 4.2%	(Shi et al. 2019)

#Anaeuploid.



**Figure 2.** Multiplexing using single gRNA expression cassettes. (A) Expression of several sgRNA cassettes from a single expression vector. (B) Expression of several sgRNA cassettes from multiple expression vectors (each harboring a different marker). (C) Expression of several sgRNA cassettes from multiple expression vectors (all harboring the same marker). (D) Transient supplementation with *in vitro* assembled RNPs or *in vitro* transcribed gRNAs.



**Figure 3.** Multiplexing using gRNA polycistronic cassettes. (A) Expression of gRNAs from synthetic array dependent on Csy4 processing. In this case, Csy4 has to be co-expressed. (B) Expression of gRNAs from synthetic array dependent on endoribonuclease splicing. In most of the reviewed examples, these synthetic arrays are expressed using tRNAs as RNA pol III promoters. (C) Expression of gRNAs from native-like CRISPR array dependent on Cas9, tracrRNA and RNase III processing. (D) Expression of gRNAs from native-like CRISPR array dependent on Cas12a processing.

### Multiplexing using gRNA polycistronic cassette

Cas9-dependent multiplex genome editing efforts reported to date are generally based on the combination of individual expression cassettes required for single targeting editing experiments. Hence, they commonly suffer from (i) repeated usage of equal sets of promoters and terminators for gRNA expression, (ii) requirement for multiple selection markers, and more importantly, (iii) labor-intensive expression cassettes and plasmid construction. By expressing multiple gRNAs under the control of a single promoter, polycistronic expression cassettes overcome these hurdles. To date, two polycistronic cassette-based approaches have been shown that enable multiplex genome editing (Fig. 3).

The first approach uses tandem arrays of chimeric sgRNAs that require a dedicated maturation mechanism to release the individual sgRNAs after transcription (Fig. 3A and B). The Csy4 ribonuclease cleaves synthetic precursor sgRNA arrays when interspaced by 28-nt sequences recognized by Csy4 (Qi et al. 2012). As such, efficient quadruplex editing by SpCas9 has been accomplished in *S. cerevisiae* (Ferreira et al. 2018). Alternatively, cleavage of the crRNA array can be performed by self-cleaving RNA sequences. As such, efficient duplex editing was achieved in the non-conventional yeasts *O. parapolymorpha* and *Saccharomyces pastorianus* using sgRNAs flanked with Hammerhead and Hepatitis Delta Virus ribozymes (Gorter de Vries et al. 2017; Juergens et al. 2018). Triplex editing was achieved in a diploid *S. cerevisiae* strain with a synthetic array of sgRNAs flanked by ribozymes (Ryan et al. 2014). In addition, tandem fusion of multiple sgRNAs-tRNAs enabling sgRNAs processing by the native tRNA-maturation system was shown to promote SpCas9-assisted multiplexed genome editing in filamentous fungi (Nodvig et al. 2018).

The second approach for polycistronic gRNA expression resembles native CRISPR arrays (Fig. 3C and D). The expression of Cas9 is required together with the transcription of a CRISPR array with multiple crRNAs and a tracrRNA. With this approach, an array of two guides was expressed and processed in a self-targeting system in *E. coli* for plasmid removal. Processing of the CRISPR array transcript occurred via unknown native endoribonucleases (Ronda et al. 2016). Again, resembling native CRISPR systems, a three-spacers array and a tracrRNA were co-expressed in *S. cerevisiae* and up to three genes were deleted with efficiencies ranging from 27–87% (depending on the sequence of the targeting guides) (Bao et al. 2015). More recently, several orthologues from the class V endonuclease Cas12a (*AsCas12a*, *LbCas12a* and *FnCas12a*) (Zetsche et al. 2015) were shown to deliver efficient multiplex genome editing of *E. coli* (Ao et al. 2018), *Streptomyces coelicolor* (Li et al. 2018) and *S. cerevisiae* (Swiat et al. 2017; Verwaal et al. 2018; Li, Wang and Wei 2018).

### Efficient delivery of donor DNA for repair of CRISPR-mediated DNA breaks

Current multiplexing strategies are based on two different procedures for the simultaneous delivery of dDNA fragments. On one hand, transient delivery of these homology repair templates is accomplished by co-transforming multiple linear dDNAs with a corresponding set of gRNAs. This approach was implemented for efficient introduction of multiple barcoded repairs fragments (short DNA fragments that contain a unique sequence tag) (Ryan and Cate 2014), and editing of heterologous metabolic pathways in both eukaryotic and prokaryotic genomes (Jiang et al. 2013; Jakočiūnas et al. 2015; Li et al. 2015; Mans et al. 2015; Ronda

et al. 2015; Verwaal et al. 2018; Wang et al. 2018a). While dDNA is often delivered in double-stranded configuration, it should be noted that successful repair using single-stranded DNA (ssDNA) has also been achieved in *S. cerevisiae* (DiCarlo et al. 2013; Generoso et al. 2016), *Aspergilli* (Nodvig et al. 2018), in *E. coli* (Li et al. 2015; Ronda et al. 2016), and in mammalian cells (Richardson et al. 2016; Song and Stieger 2017). Alternatively, a stable source of dDNA sequences is provided when these are cloned in one multi-copy plasmid (Cobb, Wang and Zhao 2015; Huang et al. 2015; Jiang et al. 2015; Gao et al. 2016; Ao et al. 2018; Feng et al. 2018; Li et al. 2018). In a more elegant way, these sequences can be cloned in tandem to the corresponding gRNAs under control of a single promoter. These hybrids are long transcripts that include multiple repair template-crRNA sequences between CRISPR repeats to be further processed into shorter gRNAs. These gRNAs will still include the transcribed sequence of each dDNA template (Bao et al. 2015; Garst et al. 2017; Roy et al. 2018).

Repair efficiencies of the double-stranded DNA break at a targeted locus depend on the size of the homology flanks (HFs) of the dDNA. Commonly used dDNA sequences may vary from short-sized HFs (~50 bp for *S. cerevisiae* or ~50–100 bp for *E. coli*), to longer, PCR-based HFs for many bacteria, non-conventional yeasts and fungi (200–1000 bp) (Table 2). Long HFs have been shown to increase the efficiency of HDR for both single target and multiplex genome editing in bacteria and eukaryotic microorganisms (Ronda et al. 2015; Ao et al. 2018; Wang et al. 2018a). In *S. cerevisiae*, multiplex gene deletions can be obtained simultaneously by using oligo-sized dDNAs with short HFs (~50 bp), either as single-stranded (Generoso et al. 2016) or as annealed double-stranded dDNAs (Mans et al. 2015; Swiat et al. 2017; Ferreira, David and Nielsen 2018) (Table 2). The amount of dDNA has previously been considered a key factor for enhancing CRISPR-Cas-mediated editing (Horwitz et al. 2015). Cells are generally transformed with a relatively high concentration of dDNAs, in the order of picomols (Horwitz et al. 2015; Mans et al. 2015). This concentration is higher than the one usually used for *in vitro* plasmid assembly procedures in *S. cerevisiae* (Kuijpers et al. 2013).

### Methods for increasing HDR frequencies versus NHEJ and alternative NHEJ repair

The potential of CRISPR-Cas9/Cas12a for implementing precise HDR-based genome editing is often hindered by (i) the presence of NHEJ or alternative-NHEJ repair systems, (ii) the presence of inefficient HDR systems, or (iii) an unfavorable balance between NHEJ and HDR repair mechanisms.

In bacteria, Cas9 and Cas12a are mainly used as counter-selection tools: the endonucleases create DSBs causing cell death due to the absence or poor efficiency of NHEJ repair systems. Only those cells that successfully obtained and integrated appropriate repair templates into their genome avoid the occurrence of persistent chromosomal breaks (Mougiakos et al. 2016). The expression of recombining systems based on the  $\lambda$ -Red recombinase is extensively used in organisms such as *E. coli*, *Lactobacillus* (Oh and van Pijkeren 2014) or *Pseudomonas putida* (Aparicio, de Lorenzo and Martínez-García 2018) to boost the efficiency of HDR and increase the number of recombinants (Mougiakos et al. 2018).

In the case of a multiplex engineering approach in yeasts, the desired genome editing (and the cell viability) relies on the stochastic allocation of dDNAs into the nucleus. In *S. cerevisiae*,

Table 3. Multiplexed genome regulation events in industrial microorganisms using CRISPRi and CRISPRa.

Species(strain(poidy))	Cas nuclease tool (CRISPRi/CRISPRa), expression, promoterPlasmid (replication origin)/genome integrated	Strategy for multiplexed gRNA expression/delivery (expression)Plasmid (replication origin)/genome integrated	Goal	Number of targets	Reference
<b>PROKARYOTES</b>					
<i>E. coli</i> [B0013]	dSpCas9 (CRISPRi)—constitutive expression, tc promoterPlasmid expression	Several sgRNA expression cassettes (constitutive)Plasmid expression (CloDF13ori)	Increase malate titer	3 targets	(Gao et al. 2018)
<i>E. coli</i> [MG1655]	ddA-Cas12a (CRISPRi)—constitutive expression, j23100 promoterPlasmid expression (p15A ori)	Several sgRNA expression cassettes (j23119-SpeI constitutive promoter)Plasmid expression (ColE1 ori)	Proof of principle	3 targets	(Zhang et al. 2017)
<i>E. coli</i> [MG1655]	dSpCas9 (CRISPRi/CRISPRa)—constitutive expression, endogenous <i>S. pyogenes</i> promoterPlasmid expression	Several sgRNA expression cassettes (j23119 constitutive promoter) fused to protein binding sequences (scaffold RNAs)Plasmid expression	Proof of principle. Activation of ethanol biosynthesis	2 targets	(Dong et al. 2018)
<i>Bacillus subtilis</i> [BNY]	dSpCas9 (CRISPRi)—inducible expression, xyIA promoterGenome integrated	Several sgRNA expression cassettes (P <sub>reg</sub> constitutive promoter)Genome integrated	Increase N-acetylglucosamine titer	3 targets	(Wu et al. 2018b)
<i>Streptomyces coelicolor</i>	dSpCas9 (CRISPRi)—constitutive expression, ermE* <sub>p</sub> promoterGenome integrated	Several sgRNA expression cassettes (j23119p constitutive promoter)Genome integrated	Proof of principle (knock-out 4 pigmented antibiotic synthesis)	3 and 4 targets	(Zhao et al. 2018)
<i>Pseudomonas putida</i>	dSpaCas9 (CRISPRi)—inducible expression, LacI-P <sub>tac</sub> promoterGenome integrated	Several sgRNA expression cassettes (P <sub>tet</sub> promoter)Plasmid expression (oriV, Rep)	Proof of principle	2 targets	(Tan et al. 2018)
<i>Corynebacterium glutamicum</i>	dSpCas9 (CRISPRi)—inducible expression, P <sub>tac</sub> promoterPlasmid expression	Several sgRNA expression cassettes (P <sub>tac</sub> inducible promoter)Plasmid expression	Increase aminoacid production	3 targets	(Cleto et al. 2016)
<i>Bacillus subtilis</i> [BNY]	dSpCas9 (CRISPRi)—inducible expression, PxyIA promoterGenome integrated	Several sgRNA expression cassettes (P <sub>veg</sub> promoter)Genome integrated	Increase GlcNAc titer	3 targets	(Wu et al. 2018)

Table 3. Continued

Species[strain(poidy)]	Cas nuclease tool (CRISPRi/CRISPRa), expression, promoter/Plasmid (replication origin)/genome integrated	Strategy for multiplexed gRNA expression/delivery (expression)/Plasmid (replication origin)/genome integrated	Goal	Number of targets	Reference
<i>Synechocystis</i>	dSpCas9 (CRISPRi)—constitutive expression, PpsbA2 promoter/Genome integrated	Several sgRNA expression cassettes (P <sub>L31</sub> constitutive promoter)/Genome integrated	Reduction of PHB and glycogen accumulation during nitrogen starvation	4 targets	(Yao et al. 2016)
<i>Synechocystis</i>	dSpCas9 (CRISPRi)—inducible expression, P <sub>L22</sub> promoter/Genome integrated	Several sgRNA expression cassettes (P <sub>L22</sub> constitutive promoter)/Genome integrated	Carbon flux re-direction for production of fatty alcohols	6 targets	(Kaczmarzyk et al. 2018)
<i>Streptomyces coelicolor</i> [M145]	ddFrCas12a (CRISPRi)—constitutive expression, ermEp* promoter/Genome integrated	Native-like CRISPR array ( <i>kasOp*</i> constitutive promoter)/Genome integrated	Proof of principle (knock-out 3 pigmented antibiotic synthesis)	3 targets	(Li et al. 2018)
<b>EUKARYOTES</b>					
<i>Saccharomyces cerevisiae</i> [CEN.PK2-a and Sigma 10 560-4A]	dCas9-VPR (CRISPRi and CRISPRa)/Plasmid expression (centromeric)	TEF1p-tRNA-sgRNA-tRNA (constitutive)/Plasmid expression (centromeric)	Increase 2,3-butanediol titer	5 targets (4 interference, 1 activation)	(Deaner, Holzman and Alper 2018)
<i>Saccharomyces cerevisiae</i>	dSpCas9 (CRISPRi/CRISPRa)—inducible expression, pGal10 promoter/Genome integrated	Several sgRNA (with RNA scaffolds) expression cassettes (SNR52p constitutive RNA pol III promoter)/Plasmid expression (centromeric)	Proof of principle, obtention of different violacein biosynthetic pathway products	3 targets	(Zalatan et al. 2015)
<i>Saccharomyces cerevisiae</i>	dLbCas12a-VP (CRISPRa), dSpCas9-RD1152 (CRISPRi), SaCas9 (CRISPRd)—constitutive expression, pTDH3 promoter/Genome integrated	Several sgRNA and gRNA in synthetic array between Csy4 recognition sites (TEF1p RNA pol II constitutive promoter)/Plasmid expression	Increase $\beta$ -carotene production	3 targets (1 activation, 1 interference and 1 deletion)	(Lian et al. 2017)
<i>Kluyveromyces marxianus</i>	dSpCas9 (CRISPRi)—constitutive expression, TEF1p promoter/Plasmid expression (centromeric)	Several sgRNA expression cassettes (RPR1-tRNA <sup>gly</sup> , RNA pol III constitutive promoter)/Plasmid expression (centromeric)	Increase ethyl acetate production	6 targets (4 genes)	(Löbs et al. 2018)
<i>Yarrowia lipolytica</i>	dSpCas9-Mxi1 (CRISPRi)—constitutive expression, UAS1B8-TEF/Plasmid expression	Several sgRNA expression cassettes (SCR1-tRNA <sup>gly</sup> RNA pol III constitutive promoter)/Plasmid expression	Increase HR by repression of the NHEJ machinery	3 targets (2 genes)	(Schwartz et al. 2017)

Table 3. Continued

Species[strain(poidy)]	Cas nuclease tool (CRISPRi/CRISPRa), expression, promoterPlasmid (replication origin)/genome integrated	Strategy for multiplexed gRNA expression/delivery (expression)Plasmid (replication origin)/genome integrated	Goal	Number of targets	Reference
<i>Saccharomyces cerevisiae</i> [BY4741 (n)]	dSpCas9-VPR (CRISPRi and CRISPRa)Plasmid expression (centromeric)	Synthetic array of ribozyme-flanked sgRNAs (Gal1p, RNA pol II inducible promoter)Plasmid expression (centromeric)	Proof of principle	2 targets	(Deaner, Mejia and Alper 2017)
<i>Saccharomyces cerevisiae</i> [BY4741 (n)]	dSpCas9-VPR (CRISPRi and CRISPRa)Plasmid expression (centromeric)	Synthetic array of ribozyme-flanked sgRNAs (TEF1p, RNA pol II constitutive promoter)Plasmid expression (centromeric)	Proof of principle	4 targets	(Deaner, Mejia and Alper 2017)
<i>Yarrowia lipolytica</i> [ATCC 201 249]	dSpCas9 or dFrCas12a (CRISPRi)—constitutive expressionPlasmid expression (centromeric)	Several sgRNA expression cassettes (SCR1-tRNA <sup>gly</sup> RNA pol III constitutive promoter)Plasmid expression (centromeric)	Proof of principle, decrease protodeoxy-violaecinic acid	3 targets	(Zhang et al. 2018)

the relatively high activity of the HDR machinery facilitates precise editing at multiple loci simultaneously (Mans et al. 2015; Swiat et al. 2017; Ferreira, David and Nielsen 2018). In contrast, the prevalent NHEJ system in non-conventional yeasts (or other difficult-to-engineer organisms, such as microalgae) might need a more extensive screening due to the high heterogeneity of transformants (Serif et al. 2018; Wang et al. 2018a). To improve the efficiency of HDR in some yeasts such as *Y. lipolytica*, the *KU70* gene responsible for DSB repair in the NHEJ pathway was disrupted (Jang et al. 2018). A similar approach is often applied in non-conventional yeasts, e.g. in *Naumovozyma castellii*, where simultaneous deletion of the orthologues of *KU70* and *KU80* completely abolished NHEJ repair during CRISPR-based editing (Vyas et al. 2018). Alternatively, researchers have inhibited certain endogenous DNA repair components to favor HDR in CRISPR experiments (Vyas et al. 2018).

The balance between DSB repair pathways is further influenced by the cell cycle phase in which a cell is. By making use of hydroxyurea-mediated cell cycle arrest (S-phase), the frequency of targeted integration is significantly increased in multiple fungi (Tsakraklides et al. 2015) and demonstrated specifically for CRISPR-based genome editing of *Y. lipolytica* (Jang et al. 2018). Finally, an improvement of HDR efficiency in single target genome editing has recently been achieved by active recruitment of the dDNA to the DSB making use of the ability of some proteins to bind DSBs in *S. cerevisiae* (Roy et al. 2018). Similar methods could be considered for obtaining an increased HDR frequency in a multiplexing set-up for a broader variety of microbes.

### Multiplex gene repression and activation

Multiplexing can also be exploited for CRISPRi/CRISPRa approaches (Table 3). Again, both dCas9 and dCas12a have been used for controlling gene expression. In many microorganisms, CRISPRi and CRISPRa have mainly been used to re-direct the carbon flux towards the production of the desired product. This usually requires fine-tuning the expression of multiple genes, which can be achieved by multiplexed gRNA expression for polygenic targeting. Multiplexing using CRISPRi has been explored in several prokaryotic industrial organisms such as *E. coli* (Zhang et al. 2017; Gao et al. 2018; Tian et al. 2019), *S. coelicolor* (Li et al. 2018; Zhao et al. 2018), *P. putida* (Tan, Reisch and Prather 2018), *Bacillus subtilis* (Wu et al. 2018b), *Corynebacterium glutamicum* (Cleto et al. 2016) and the cyanobacterium *Synechocystis* (Yao et al. 2016; Kaczmarzyk et al. 2018). Furthermore, different approaches have been explored in the yeasts *S. cerevisiae* (Jensen et al. 2017; Lian et al. 2017), *Y. lipolytica* (Schwartz et al. 2017; Zhang et al. 2018) and *Kluyveromyces marxianus* (Löbs et al. 2018). Independent studies on the implementation of Cas12a for CRISPRi purposes reported changes in repression strength after altering the order of the spacers in the CRISPR array (Liao et al. 2018). Other studies did not observe the same effect (Wang et al. 2017; Zetsche et al. 2017; Zhang et al. 2017). This suggests that the phenomenon could be gRNA-sequence-dependent and therefore most likely related to transcript secondary structure formation.

Fewer examples can be found of the use of CRISPRa because of the requirement of functional transcription activators. Frequently, CRISPRa applications are used in combination with CRISPRi approaches (Lian et al. 2017). Combination of CRISPRi and CRISPRa using RNA scaffolds was accomplished using similar designs in *E. coli* (Dong et al. 2018) and *S. cerevisiae* (Zalatan et al. 2015; Lian et al. 2017). In both cases, protein-binding RNA

sequences were fused to the 3' end of sgRNAs of Cas9. These protein-recruitment RNA sequences have a high affinity for certain proteins, such as the MCP (used both in *E. coli* and *S. cerevisiae*) or the PCP and Com proteins (tested only in *S. cerevisiae*). By fusing the transcriptional activation domain SoxS (in *E. coli*) or VP64 (in *S. cerevisiae*) to these RNA binding proteins, expression of a gene downstream a target promoter can be enhanced, and pathway fluxes sequentially directed (Zalatan et al. 2015; Dong et al. 2018).

### Challenges and outlook of multiplexing

In recent years, remarkable progress has been achieved in the field of multiplexed genome editing. Besides broadening the spectrum of microorganisms that can be engineered using CRISPR-Cas endonucleases, novel approaches have recently been implemented in terms of gRNA and dDNA delivery for increasing multiplexing efficiency. Below, a summary is provided of the major challenges related to the development of efficient multiplexing CRISPR-Cas systems in microorganisms.

1. *Guide design*: availability of highly efficient guides (sgRNA, crRNA) obtained either through software prediction based on generalized well defined guide-design principles, preferential PAM domains and secondary structure prediction (Chari et al. 2015; Graham and Root 2015; Moreno-Mateos et al. 2015; Labuhn et al. 2018; Liao et al. 2018), or through pre-characterization of the functionality of individual guide performance in single target genome editing experiments.
2. *dDNA design and delivery*: dDNA can be part of a vector, provided as a linear fragment, single or double-stranded, a single fragment of multiple fused repair templates or provided as multiple DNA fragments. Proper characterization of such elements (e. g. by determination of optimal size of homology sequences) for the specific host organism to be edited is required. Additionally, dDNA might be stabilized by chemical modifications (Lee et al. 2017).
3. *Controllable expression of CRISPR elements*: tight expression control of guides and nucleases, via tuning of expression either by systematic variation of constitutive promoters or by using inducible systems. Optimized promoter and terminator sequences are required to fine-tune the expression of the different CRISPR elements (Feng et al. 2018).
4. *Innovative plasmid assembly methods*: smart DNA construction schemes to develop single or multi-plasmid systems containing elements of the CRISPR-Cas system (gRNA, Cas, dDNA, selective marker), including techniques to incorporate repetitive DNA elements such as the repeat sequences of CRISPR arrays (Cress et al. 2015; Deaner, Holzman and Alper 2018; Liao et al. 2018). Recently, a cloning-free approach was developed in *S. cerevisiae* for the obtainment of up to 6 simultaneous deletions using SpCas9 with a efficiency of 23.3%. The strategy combined multiple successful strategies presented in this manuscript. Three transcripts (each of them containing two gRNAs flanked by tRNA<sup>gly</sup> sequences were expressed from a single plasmid encoding for SpCas9. The assembly of the plasmid via Golden Gate reaction was performed in the yeast (Zhang et al. 2019).
5. *Organism-specific CRISPR tools*: smart choice of the CRISPR-Cas expression approach depending on final application of the production strain or the target organism of choice. For instance, CRISPR-Cas tools can be combined with the introduction of dDNA containing selective markers, which

makes screening and selection of positive clones more efficient. This approach can be used in proof of principle studies, whereas in other cases, marker-free strains are important. In a similar way, guides and Cas nucleases can be co-expressed from plasmid-borne expression cassettes or expressed sequentially in strains pre-expressing the Cas nuclease from a second plasmid or from a genome integrated copy.

6. *Editing conditions*: optimization of organism-specific CRISPR-Cas delivery systems and recovery protocols. Cell synchronization protocols in combination with CRISPR-Cas systems have been used in human cells to enhance HDR versus NHEJ repair (Lin et al. 2014). In some yeasts, the highest rate of HDR over NHEJ is shown during S-phase (Tsakraklides et al. 2015). Therefore, this strategy has been already proposed for its use in industrial microorganisms (Juergens et al. 2018).
7. *Novel or improved endonucleases*: these endonucleases should have alternative or less-stringent PAM recognition selection. In addition, nucleases are preferred that are smaller, more specific and more active, as reviewed by Kleinstiver et al. (2019). Cas9 variants with distinct PAM-recognizing features have been obtained by laboratory evolution (Hu et al. 2018), as well as by structure-guided protein engineering of Cas12a (Kleinstiver et al. 2019). The discovery and characterization of novel Cas endonucleases (Liu et al. 2019) can also extend the PAM compatibility (Burstein et al. 2017; Harrington et al. 2018; Strecker et al. 2019). At the same time, multiple endonucleases can be used in orthogonal designs in order to edit the genome and regulate gene expression simultaneously (Kweon et al. 2017; Lian et al. 2017). Alternatively, orthogonal designs can incorporate modified gRNAs, as described for Cas12a by Breinig et al. (2019).

In multiplexed genome editing experiments, a negative correlation is experienced between the number of targets and the amount of obtained colonies after transformation in most microorganisms. The introduction of DSBs dramatically reduces the cell survival rate, and this causes limited numbers of simultaneous modifications as a delicate balance between DNA cleavage and repair needs to be established. Alternatively, multiplexed single-base editing does not depend on DSB generation nor dDNA supply and can be used to introduce nucleic acid base changes at a targeted window of DNA (Eid, Alshareef and Mahfouz 2018; Wu et al. 2018a). In this approach, deactivated or nickase Cas nuclease variants coupled to base editors (cytidine or adenine deaminases) are directed to a target site by the gRNA (Marx 2018). Nucleotide changes are introduced in a targeted DNA window rather than in a precise DNA position, which makes this technique less accurate for the introduction of single point mutations. Multiplexed single-base editing has been recently implemented in prokaryotic microorganisms (Banno et al. 2018; Wang et al. 2018b). Useful applications of this technique remain limited to phenotype modifications linked to single nucleotide polymorphisms (SNPs) and to the possibility of introducing stop-codons for gene inactivation in coordination with the presence of a PAM sequence in a determined window (Arazoe, Kondo and Nishida 2018). Although very promising, further development and control of this base editing approach are required in order to broaden the type of modifications and to avoid unwanted mutations (Nishida et al. 2016).

Advances in multiplexed genome editing of microorganisms can significantly accelerate future strain construction programs of cell factories with unprecedented efficiencies. Therefore, the CRISPR revolution continues: new tools and workflows are being

developed to broaden the range of functionalities of currently used CRISPR-Cas systems as well as knowledge about the mechanism of these systems. As identified in this review, dedicated optimization of each of the elements involved in CRISPR-Cas genome editing is crucial for efficient multiplex genome editing and for stretching the number of simultaneous editing events.

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## REFERENCES

- Alberti F, Corre C. Editing streptomyces genomes in the CRISPR/Cas9 age. *Nat Prod Rep* 2019. DOI: 10.1039/C8NP00081F.
- Ao X, Yao Y, Li T et al. A multiplex genome editing method for *Escherichia coli* based on CRISPR-Cas12a. *Front Microbiol* 2018;9:2307. DOI:10.3389/fmicb.2018.02307.
- Aparicio T, de Lorenzo V, Martínez-García E. CRISPR/Cas9-Based Counterselection Boosts Recombineering Efficiency in *Pseudomonas putida*. *Biotechnol J* 2018;13:e1700161. DOI: 10.1002/biot.201700161.
- Arazoe T, Kondo A, Nishida K. Targeted nucleotide editing technologies for microbial metabolic engineering. *Biotechnol J* 2018;13:1700596.
- Banno S, Nishida K, Arazoe T et al. Deaminase-mediated multiplex genome editing in *Escherichia coli*. *Nat Microbiol* 2018;3:423–9.
- Bao Z, Xiao H, Liang J et al. Homology-integrated CRISPR-Cas (HI-CRISPR) system for one-step multigene disruption in *Saccharomyces cerevisiae*. *ACS Synth Biol* 2015;4:585–94.
- Barrangou R, Fremaux C, Deveau H et al. CRISPR provides acquired resistance against viruses in prokaryotes. *Science* 2007;315:1709–12.
- Behler J, Vijay D, Hess WR et al. CRISPR-based technologies for metabolic engineering in cyanobacteria. *Trends Biotechnol* 2018;36:996–1010.
- Berlec A, Škrlec K, Kocjan J et al. Single plasmid systems for inducible dual protein expression and for CRISPR-Cas9/CRISPRi gene regulation in lactic acid bacterium *Lactococcus lactis*. *Sci Rep* 2018;8:1009.
- Bikard D, Jiang W, Samai P et al. Programmable repression and activation of bacterial gene expression using an engineered CRISPR-Cas system. *Nucleic Acids Res* 2013;41:7429–37.
- Bowater R, Doherty AJ. Making ends meet: repairing breaks in bacterial DNA by non-homologous end-joining. *PLoS Genet* 2006;2:e8.
- Breinig M, Schweitzer AY, Herianto AM et al. Multiplexed orthogonal genome editing and transcriptional activation by Cas12a. *Nat Methods* 2019;16:51–4.
- Brouns SJJ, Jore MM, Lundgren M et al. Small CRISPR RNAs guide antiviral defense in prokaryotes. *Science* 2008;321:960–4.
- Burstein D, Harrington LB, Strutt SC et al. New CRISPR-Cas systems from uncultivated microbes. *Nature* 2017;542:237–41.



- Cao M, Gao M, Ploessl D et al. CRISPR-mediated genome editing and gene repression in *Scheffersomyces stipitidis*. *Biotechnol J* 2018;**13**:e1700598.
- Chari R, Mali P, Moosburner M et al. Unraveling CRISPR-Cas9 genome engineering parameters via a library-on-library approach. *Nat Methods* 2015;**12**:823–6.
- Chavez A, Scheiman J, Vora S et al. Highly efficient Cas9-mediated transcriptional programming. *Nat Methods* 2015;**12**:326–8.
- Chayot R, Montagne B, Mazel D et al. An end-joining repair mechanism in *Escherichia coli*. *Proc Natl Acad Sci U S A* 2010;**107**:2141–6.
- Choi KR, Jang WD, Yang D et al. Systems metabolic engineering strategies: integrating systems and synthetic biology with metabolic engineering. *Trends Biotechnol* 2019;**17**:44:1–21. DOI: 10.1016/j.tibtech.2019.01.003.
- Choi KR, Lee SY. CRISPR technologies for bacterial systems: current achievements and future directions. *Biotechnol Adv* 2016;**34**:1180–209.
- Cleto S, Jensen JV, Wendisch VF et al. *Corynebacterium glutamicum* metabolic engineering with CRISPR interference (CRISPRi). *ACS Synth Biol* 2016;**5**:375–85.
- Cobb RE, Wang Y, Zhao H. High-efficiency multiplex genome editing of *Streptomyces* species using an engineered CRISPR/Cas system. *ACS Synth Biol* 2015;**4**:723–8.
- Cong L, Ran FA, Cox D et al. Multiplex genome engineering using CRISPR/Cas systems. *Science* 2013;**339**:819–23.
- Cress BF, Toparlak ÖD, Guleria S et al. CRISPRi: modular combinatorial assembly of Type II-A CRISPR arrays for dCas9-mediated multiplex transcriptional repression in *E. coli*. *ACS Synth Biol* 2015;**4**:987–1000.
- Dai Z, Nielsen J. Advancing metabolic engineering through systems biology of industrial microorganisms. *Curr Opin Biotechnol* 2015;**36**:8–15.
- Deaner M, Holzman A, Alper HS. Modular ligation extension of guide RNA operons (LEGO) for multiplexed dCas9 regulation of metabolic pathways in *Saccharomyces cerevisiae*. *Biotechnol J* 2018;**13**:1700582.
- Deaner M, Mejia J, Alper HS. Enabling graded and large-scale multiplex of desired genes using a dual-mode dCas9 activator in *Saccharomyces cerevisiae*. *ACS Synth Biol* 2017;**6**:1931–43.
- Deltcheva E, Chylinski K, Sharma CM et al. CRISPR RNA maturation by trans-encoded small RNA and host factor RNase III. *Nature* 2011;**471**:602–7.
- Deveau H, Barrangou R, Garneau JE et al. Phage response to CRISPR-encoded resistance in *Streptococcus thermophilus*. *J Bacteriol* 2008;**190**:1390–400.
- DiCarlo JE, Norville JE, Mali P et al. Genome engineering in *Saccharomyces cerevisiae* using CRISPR-Cas systems. *Nucleic Acids Res* 2013;**41**:4336–43.
- Dong C, Fontana J, Patel A et al. Synthetic CRISPR-Cas gene activators for transcriptional reprogramming in bacteria. *Nat Commun* 2018;**9**:2489.
- Donohoue PD, Barrangou R, May AP. Advances in industrial biotechnology using CRISPR-cas systems. *Trends Biotechnol* 2018;**36**:134–46.
- Eid A, Alshareef S, Mahfouz MM. CRISPR base editors: genome editing without double-stranded breaks. *Biochem J* 2018;**475**:1955–64.
- Feng X, Zhao D, Zhang X et al. CRISPR/Cas9 assisted multiplex genome editing technique in *Escherichia coli*. *Biotechnol J* 2018;**13**:1700604.
- Ferreira R, David F, Nielsen J. Advancing biotechnology with CRISPR/Cas9: recent applications and patent landscape. *J Ind Microbiol Biotechnol* 2018;**45**:467–80.
- Ferreira R, Skrekas C, Nielsen J et al. Multiplexed CRISPR/Cas9 genome editing and gene regulation using Csy4 in *Saccharomyces cerevisiae*. *ACS Synth Biol* 2018;**7**:10–5.
- Fonfara I, Richter H, Bratovic M et al. The CRISPR-associated DNA-cleaving enzyme Cpf1 also processes precursor CRISPR RNA. *Nature* 2016;**532**:517–21.
- Foster AJ, Martin-Urdiroz M, Yan X et al. CRISPR-Cas9 ribonucleoprotein-mediated co-editing and counterselection in the rice blast fungus. *Sci Rep* 2018;**8**:14355.
- Gao C, Wang S, Hu G et al. Engineering *Escherichia coli* for malate production by integrating modular pathway characterization with CRISPRi-guided multiplexed metabolic tuning. *Biotechnol Bioeng* 2018;**115**:661–72.
- Gao S, Tong Y, Wen Z et al. Multiplex gene editing of the *Yarrowia lipolytica* genome using the CRISPR-Cas9 system. *J Ind Microbiol Biotechnol* 2016;**43**:1085–93.
- Gao Y, Zhao Y. Self-processing of ribozyme-flanked RNAs into guide RNAs in vitro and in vivo for CRISPR-mediated genome editing. *J Integr Plant Biol* 2014;**56**:343–9.
- Garst AD, Bassalo MC, Pines G et al. Genome-wide mapping of mutations at single-nucleotide resolution for protein, metabolic and genome engineering. *Nat Biotechnol* 2017;**35**:48–55.
- Gasiunas G, Barrangou R, Horvath P et al. Cas9-crRNA ribonucleoprotein complex mediates specific DNA cleavage for adaptive immunity in bacteria. *Proc Natl Acad Sci U S A* 2012;**109**:E2579–86.
- Generoso WC, Gottardi M, Oreb M et al. Simplified CRISPR-Cas genome editing for *Saccharomyces cerevisiae*. *J Microbiol Methods* 2016;**127**:203–5.
- Gilbert LA, Larson MH, Morsut L et al. CRISPR-mediated modular RNA-guided regulation of transcription in eukaryotes. *Cell* 2013;**154**:442–51.
- Gorter de Vries ARG, de Groot PA, van den Broek M et al. CRISPR-Cas9 mediated gene deletions in lager yeast *Saccharomyces pastorianus*. *Microb Cell Fact* 2017;**16**:222.
- Graham DB, Root DE. Resources for the design of CRISPR gene editing experiments. *Genome Biol* 2015;**16**:260.
- Harrington LB, Burstein D, Chen JS et al. Programmed DNA destruction by miniature CRISPR-Cas14 enzymes. *Science* 2018;**362**:839–42.
- Hong K-K, Nielsen J. Metabolic engineering of *Saccharomyces cerevisiae*: a key cell factory platform for future biorefineries. *Cell Mol Life Sci* 2012;**69**:2671–90.
- Horwitz AA, Walter JM, Schubert MG et al. Efficient multiplexed integration of synergistic alleles and metabolic pathways in yeasts via CRISPR-Cas. *Cell Syst* 2015;**1**:88–96.
- Huang H, Zheng G, Jiang W et al. One-step high-efficiency CRISPR/Cas9-mediated genome editing in *Streptomyces*. *Acta Biochim Biophys Sin* 2015;**47**:231–43.
- Hu JH, Miller SM, Geurts MH et al. Evolved Cas9 variants with broad PAM compatibility and high DNA specificity. *Nature* 2018;**556**:57–63.
- Jakočiūnas T, Bonde I, Herrgård M et al. Multiplex metabolic pathway engineering using CRISPR/Cas9 in *Saccharomyces cerevisiae*. *Metab Eng* 2015;**28**:213–22.
- Jang I-S, Yu BJ, Jang JY et al. Improving the efficiency of homologous recombination by chemical and biological approaches in *Yarrowia lipolytica*. *PLoS One* 2018;**13**:e0194954.

- Jensen ED, Ferreira R, Jakociūnas T et al. Transcriptional reprogramming in yeast using dCas9 and combinatorial gRNA strategies. *Microb Cell Fact* 2017;**16**:46.
- Jensen MK. Design principles for nuclease-deficient CRISPR-based transcriptional regulators. *FEMS Yeast Res* 2018;**18**. DOI: 10.1093/femsyr/foy039.
- Jiang W, Bikard D, Cox D et al. RNA-guided editing of bacterial genomes using CRISPR-Cas systems. *Nat Biotechnol* 2013;**31**:233–9.
- Jiang Y, Chen B, Duan C et al. Multigene editing in the *Escherichia coli* genome via the CRISPR-Cas9 system. *Appl Environ Microbiol* 2015;**81**:2506–14.
- Jinek M, Chylinski K, Fonfara I et al. A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science* 2012;**337**:816–21.
- Juergens H, Varela JA, de Vries ARG et al. Genome editing in *Kluyveromyces* and *Ogataea* yeasts using a broad-host-range Cas9/gRNA co-expression plasmid. *FEMS Yeast Res* 2018;**18**. DOI: 10.1093/femsyr/foy012.
- Kaczmarzyk D, Cengic I, Yao L et al. Diversion of the long-chain acyl-ACP pool in *Synechocystis* to fatty alcohols through CRISPRi repression of the essential phosphate acyltransferase PlsX. *Metab Eng* 2018;**45**:59–66.
- Kleinstiver BP, Sousa AA, Walton RT et al. Engineered CRISPR-Cas12a variants with increased activities and improved targeting ranges for gene, epigenetic and base editing. *Nat Biotechnol* 2019;**37**:276–82.
- Koonin EV, Makarova KS, Zhang F. Diversity, classification and evolution of CRISPR-Cas systems. *Curr Opin Microbiol* 2017;**37**:67–78.
- Kuijpers NGA, Solis-Escalante D, Bosman L et al. A versatile, efficient strategy for assembly of multi-fragment expression vectors in *Saccharomyces cerevisiae* using 60 bp synthetic recombination sequences. *Microb Cell Fact* 2013;**12**:47.
- Kweon J, Jang A-H, Kim D-E et al. Fusion guide RNAs for orthogonal gene manipulation with Cas9 and Cpf1. *Nat Commun* 2017;**8**:1723.
- Labuhn M, Adams FF, Ng M et al. Refined sgRNA efficacy prediction improves large- and small-scale CRISPR-Cas9 applications. *Nucleic Acids Res* 2018;**46**:1375–85.
- La Russa MF, Qi LS. The new state of the art: Cas9 for gene activation and repression. *Mol Cell Biol* 2015;**35**:3800–9.
- Lee K, Mackley VA, Rao A et al. Synthetically modified guide RNA and donor DNA are a versatile platform for CRISPR-Cas9 engineering. *Elife* 2017;**6**. DOI: 10.7554/eLife.25312.
- Lee ME, DeLoache WC, Cervantes B et al. A highly characterized yeast toolkit for modular, multipart assembly. *ACS Synth Biol* 2015;**4**:975–86.
- Lee SY, Kim HU, Chae TU, et al. A comprehensive metabolic map for production of bio-based chemicals. *Nature Catalysis* 2019;**2**:18–33.
- Lian J, Hamedirad M, Hu S et al. Combinatorial metabolic engineering using an orthogonal tri-functional CRISPR system. *Nat Commun* 2017;**8**:1688.
- Liao C, Ttofali F, Slotkowski RA et al. One-step assembly of large CRISPR arrays enables multi-functional targeting and reveals constraints on array design. *bioRxiv* 2018 [Preprint]. DOI: 10.1101/312421.
- Li L, Wei K, Zheng G et al. CRISPR-Cpf1-assisted multiplex genome editing and transcriptional repression in *Streptomyces*. *Appl Environ Microbiol* 2018;**84**:e00827–18.
- Lin S, Staahl BT, Alla RK et al. Enhanced homology-directed human genome engineering by controlled timing of CRISPR/Cas9 delivery. *Elife* 2014;**3**:e04766.
- Liu J-J, Orlova N, Oakes BL et al. CasX enzymes comprise a distinct family of RNA-guided genome editors. *Nature* 2019;**566**:218–23.
- Liu Q, Gao R, Li J et al. Development of a genome-editing CRISPR/Cas9 system in thermophilic fungal *Myceliophthora* species and its application to hyper-cellulase production strain engineering. *Biotechnol Biofuels* 2017;**10**:1.
- Liu R, Chen L, Jiang Y et al. Efficient genome editing in filamentous fungus *Trichoderma reesei* using the CRISPR/Cas9 system. *Cell Discov* 2015;**1**:15007.
- Li Y, Lin Z, Huang C et al. Metabolic engineering of *Escherichia coli* using CRISPR-Cas9 mediated genome editing. *Metab Eng* 2015;**31**:13–21.
- Li Z-H, Wang F-Q, Wei D-Z. Self-cloning CRISPR/Cpf1 facilitated genome editing in *Saccharomyces cerevisiae*. *Bioresour Bioprocess* 2018;**5**:36.
- Löbs A-K, Schwartz C, Thorwall S et al. Highly multiplexed CRISPRi repression of respiratory functions enhances mitochondrial localized ethyl acetate biosynthesis in *Kluyveromyces marxianus*. *ACS Synth Biol* 2018;**7**:2647–55.
- Makarova KS, Koonin EV. *Annotation and Classification of CRISPR-Cas Systems*. New York, NY: Humana Press, 2015, 47–75.
- Makarova KS, Wolf YI, Alkhnbashi OS et al. An updated evolutionary classification of CRISPR-Cas systems. *Nat Rev Microbiol* 2015;**13**:722–36.
- Mali P, Yang L, Esvelt KM et al. RNA-guided human genome engineering via Cas9. *Science* 2013;**339**:823–6.
- Mans R, van Rossum HM, Wijsman M et al. CRISPR/Cas9: a molecular Swiss army knife for simultaneous introduction of multiple genetic modifications in *Saccharomyces cerevisiae*. *FEMS Yeast Res* 2015;**15**. DOI:10.1093/femsyr/fov004.
- Marx V. Base editing a CRISPR way. *Nat Methods* 2018;**15**:767–70.
- McGovern S, Baconnais S, Roblin P et al. C-terminal region of bacterial Ku controls DNA bridging, DNA threading and recruitment of DNA ligase D for double strand breaks repair. *Nucleic Acids Res* 2016;**44**:4785–806.
- Mertens S, Gallone B, Steensels J et al. Reducing phenolic off-flavors through CRISPR-based gene editing of the FDC1 gene in *Saccharomyces cerevisiae* x *Saccharomyces eubayanus* hybrid lager beer yeasts. *PLoS One* 2019;**14**:e0209124.
- Mojica FJM, Díez-Villaseñor C, García-Martínez J et al. Intervening sequences of regularly spaced prokaryotic repeats derive from foreign genetic elements. *J Mol Evol* 2005;**60**:174–82.
- Moreno-Mateos MA, Vejnar CE, Beaudoin J-D et al. CRISPRscan: designing highly efficient sgRNAs for CRISPR-Cas9 targeting in vivo. *Nat Methods* 2015;**12**:982–8.
- Morse NJ, Wagner JM, Reed KB et al. T7 polymerase expression of guide RNAs in vivo allows exportable CRISPR-Cas9 editing in multiple yeast hosts. *ACS Synth Biol* 2018;**3**:51.
- Mougiakos I, Bosma EF, de Vos WM et al. Next generation prokaryotic engineering: the CRISPR-Cas toolkit. *Trends Biotechnol* 2016;**34**:575–87.
- Mougiakos I, Bosma EF, Ganguly J et al. Hijacking CRISPR-Cas for high-throughput bacterial metabolic engineering: advances and prospects. *Curr Opin Biotechnol* 2018;**50**:146–57.
- Naduthodi MIS, Barbosa MJ, van der Oost J. Progress of CRISPR-Cas based genome editing in photosynthetic microbes. *Biotechnol J* 2018;**13**:1700591.
- Nayak DD, Metcalf WW. Cas9-mediated genome editing in the methanogenic archaeon *Methanosarcina acetivorans*. *Proc Natl Acad Sci U S A* 2017;**114**:2976–81.

- Nishida K, Arazoe T, Yachie N et al. Targeted nucleotide editing using hybrid prokaryotic and vertebrate adaptive immune systems. *Science* 2016;**353**:aaf8729.
- Nødvig CS, Hoof JB, Kogle ME et al. Efficient oligo nucleotide mediated CRISPR-Cas9 gene editing in *Aspergilli*. *Fungal Genet Biol* 2018;**115**:78–89.
- Nødvig CS, Nielsen JB, Kogle ME et al. A CRISPR-Cas9 system for genetic engineering of filamentous fungi. *PLoS One* 2015;**10**:e0133085.
- Oh J-H, van Pijkeren J-P. CRISPR-Cas9-assisted recombineering in *Lactobacillus reuteri*. *Nucleic Acids Res* 2014;**42**:e131.
- Pawelczak KS, Gavande NS, VanderVere-Carozza PS et al. Modulating DNA repair pathways to improve precision genome engineering. *ACS Chem Biol* 2018;**13**:389–96.
- Pohl C, Kiel JA, Driessen AJ et al. CRISPR/Cas9 based genome editing of *Penicillium chrysogenum*. *ACS Synth Biol* 2016;**5**:754–64.
- Pohl C, Mozsik L, Driessen AJM et al. Genome editing in *Penicillium chrysogenum* using Cas9 ribonucleoprotein particles. *Methods Mol Biol* 2018;**1772**:213–32.
- Qi L, Haurwitz RE, Shao W et al. RNA processing enables predictable programming of gene expression. *Nat Biotechnol* 2012;**30**:1002–6.
- Qi LS, Larson MH, Gilbert LA et al. Repurposing CRISPR as an RNA-guided platform for sequence-specific control of gene expression. *Cell* 2013;**152**:1173–83.
- Raschmanová H, Weninger A, Glieder A et al. Implementing CRISPR-Cas technologies in conventional and non-conventional yeasts: current state and future prospects. *Biotechnol Adv* 2018;**36**:641–65.
- Reisch CR, Prather KLJ. The no-SCAR (scarless Cas9 assisted recombineering) system for genome editing in *Escherichia coli*. *Sci Rep* 2015;**5**:15096.
- Richardson CD, Ray GJ, DeWitt MA et al. Enhancing homology-directed genome editing by catalytically active and inactive CRISPR-Cas9 using asymmetric donor DNA. *Nat Biotechnol* 2016;**34**:339–44.
- Ronda C, Maury J, Jakociunas T et al. CrEdit: CRISPR mediated multi-loci gene integration in *Saccharomyces cerevisiae*. *Microb Cell Fact* 2015;**14**:97.
- Ronda C, Pedersen LE, Sommer MOA et al. CRMAGE: CRISPR optimized MAGE recombineering. *Sci Rep* 2016;**6**:19452.
- Roy KR, Smith JD, Vonesch SC et al. Multiplexed precision genome editing with trackable genomic barcodes in yeast. *Nat Biotechnol* 2018;**36**:512–20.
- Ryan OW, Cate JHD. Multiplex engineering of industrial yeast genomes using CRISPRm. *Methods Enzymol* 2014;**546**:473–89.
- Ryan OW, Skerker JM, Maurer MJ et al. Selection of chromosomal DNA libraries using a multiplex CRISPR system. *Elife* 2014;**3**. DOI: 10.7554/eLife.03703.
- Schwartz C, Curtis N, Löbs A-K et al. Multiplexed CRISPR activation of cryptic sugar metabolism enables *Yarrowia Lipolytica* growth on cellobiose. *Biotechnol J* 2018;**13**:1700584.
- Schwartz C, Frogue K, Ramesh A et al. CRISPRi repression of non-homologous end-joining for enhanced genome engineering via homologous recombination in *Yarrowia lipolytica*. *Biotechnol Bioeng* 2017;**114**:2896–906.
- Schwartz CM, Hussain MS, Blenner M et al. Synthetic RNA polymerase III promoters facilitate high-efficiency CRISPR-Cas9-mediated genome editing in *Yarrowia lipolytica*. *ACS Synth Biol* 2016;**5**:356–9.
- Serif M, Dubois G, Finoux A-L et al. One-step generation of multiple gene knock-outs in the diatom *Phaeodactylum tricornutum* by DNA-free genome editing. *Nat Commun* 2018;**9**:3924.
- Sfeir A, Symington LS. Microhomology-mediated end joining: a back-up survival mechanism or dedicated pathway? *Trends Biochem Sci* 2015;**40**:701–14.
- Shi T-Q, Gao J, Wang W-J et al. CRISPR/Cas9-based genome editing in the filamentous fungus *Fusarium fujikuroi* and its application in strain engineering for gibberellic acid production. *ACS Synth Biol* 2019;**8**:445–54.
- Shi T-Q, Huang H, Kerkhoven EJ et al. Advancing metabolic engineering of *Yarrowia lipolytica* using the CRISPR/Cas system. *Appl Microbiol Biotechnol* 2018;**102**:9541–8.
- Shi T-Q, Liu G-N, Ji R-Y et al. CRISPR/Cas9-based genome editing of the filamentous fungi: the state of the art. *Appl Microbiol Biotechnol* 2017;**101**:7435–43.
- Shmakov S, Abudayyeh OO, Makarova KS et al. Discovery and functional characterization of diverse class 2 CRISPR-Cas systems. *Mol Cell* 2015;**60**:385–97.
- Shmakov S, Smargon A, Scott D et al. Diversity and evolution of class 2 CRISPR-Cas systems. *Nat Rev Microbiol* 2017;**15**:169–82.
- Si T, Chao R, Min Y et al. Automated multiplex genome-scale engineering in yeast. *Nat Commun* 2017;**8**:15187.
- Song F, Stieger K. Optimizing the DNA donor template for homology-directed repair of double-strand breaks. *Mol Ther Nucleic Acids* 2017;**7**:53–60.
- Song L, Ouedraogo J-P, Kolbusz M et al. Efficient genome editing using tRNA promoter-driven CRISPR/Cas9 gRNA in *Aspergillus niger*. *PLoS One* 2018;**13**:e0202868.
- Stovicek V, Holkenbrink C, Borodina I. CRISPR/Cas system for yeast genome engineering: advances and applications. *FEMS Yeast Res* 2017;**17**. DOI: 10.1093/femsyr/fox030.
- Strecker J, Jones S, Koopal B et al. Engineering of CRISPR-Cas12b for human genome editing. *Nat Commun* 2019;**10**:212.
- Swarts DC, Jinek M. Cas9 versus Cas12a/Cpf1: structure-function comparisons and implications for genome editing. *Wiley Interdiscip Rev RNA* 2018;**9**:e1481.
- Swarts DC, van der Oost J, Jinek M. Structural basis for guide RNA processing and seed-dependent DNA targeting by CRISPR-Cas12a. *Mol Cell* 2017;**66**:221–233.e4.
- Swiat MA, Dashko S, den Ridder M et al. FnCpf1: a novel and efficient genome editing tool for *Saccharomyces cerevisiae*. *Nucleic Acids Res* 2017;**45**:12585–98.
- Tan SZ, Reisch CR, Prather KLJ. A robust CRISPR interference gene repression system in *Pseudomonas*. *J Bacteriol* 2018;**200**:e00575–17.
- Tian T, Kang JW, Kang A et al. Redirecting metabolic flux via combinatorial multiplex CRISPRi-mediated repression for isopentenol production in *Escherichia coli*. *ACS Synth Biol* 2019;**8**:391–402.
- Tsakraklides V, Brevnova E, Stephanopoulos G et al. Improved gene targeting through cell cycle synchronization. *PLoS One* 2015;**10**:e0133434.
- Verwaal R, Buiting-Wiessenhaan N, Dalhuijsen S et al. CRISPR/Cpf1 enables fast and simple genome editing of *Saccharomyces cerevisiae*. *Yeast* 2018;**35**:201–11, DOI: 10.1002/yea.3278.
- Vyas VK, Bushkin GG, Bernstein DA et al. New CRISPR mutagenesis strategies reveal variation in repair mechanisms among fungi. *mSphere* 2018;**3**:e00154–18.
- Wang L, Deng A, Zhang Y et al. Efficient CRISPR-Cas9 mediated multiplex genome editing in yeasts. *Biotechnol Biofuels* 2018a;**11**:277.
- Wang M, Mao Y, Lu Y et al. Multiplex gene editing in rice using the CRISPR-Cpf1 system. *Mol Plant* 2017;**10**:1011–3.

- Wang Y, Liu Y, Liu J et al. MACBETH: multiplex automated *Corynebacterium glutamicum* base editing method. *Metab Eng* 2018b;**47**:200–10.
- Weninger A, Hatzl A-M, Schmid C et al. Combinatorial optimization of CRISPR/Cas9 expression enables precision genome engineering in the methylotrophic yeast *Pichia pastoris*. *J Biotechnol* 2016;**235**:139–49.
- Wijsman M, Świat MA, Marques WL et al. A toolkit for rapid CRISPR-SpCas9 assisted construction of hexose-transport-deficient *Saccharomyces cerevisiae* strains. *FEMS Yeast Res* 2019;**19**. DOI:10.1093/femsyr/foy107.
- Wong L, Engel J, Jin E et al. YaliBricks, a versatile genetic toolkit for streamlined and rapid pathway engineering in *Yarrowia lipolytica*. *Metab Eng Commun* 2017;**5**:68–77.
- Wu WY, Lebbink JHG, Kanaar R et al. Genome editing by natural and engineered CRISPR-associated nucleases. *Nat Chem Biol* 2018a;**14**:642–51.
- Wu Y, Chen T, Liu Y et al. CRISPRi allows optimal temporal control of N-acetylglucosamine bioproduction by a dynamic coordination of glucose and xylose metabolism in *Bacillus subtilis*. *Metab Eng* 2018b;**49**:232–41.
- Yao L, Cengic I, Anfelt J et al. Multiple gene repression in cyanobacteria using CRISPRi. *ACS Synth Biol* 2016;**5**:207–12.
- Yao X, Wang X, Hu X et al. Homology-mediated end joining-based targeted integration using CRISPR/Cas9. *Cell Res* 2017;**27**:801–14.
- Zalatan JG, Lee ME, Almeida R et al. Engineering complex synthetic transcriptional programs with CRISPR RNA Scaffolds. *Cell* 2015;**160**:339–50.
- Zetsche B, Gootenberg JS, Abudayyeh OO et al. Cpf1 is a single RNA-guided endonuclease of a class 2 CRISPR-Cas system. *Cell* 2015;**163**:759–71.
- Zetsche B, Heidenreich M, Mohanraju P et al. Multiplex gene editing by CRISPR–Cpf1 using a single crRNA array. *Nat Biotechnol* 2017;**35**:31–4.
- Zhang J-L, Peng Y-Z, Liu D et al. Gene repression via multiplex gRNA strategy in *Y. lipolytica*. *Microb Cell Fact* 2018;**17**:62.
- Zhang X, Wang J, Cheng Q et al. Multiplex gene regulation by CRISPR-ddCpf1. *Cell Discovery* 2017;**3**:17018.
- Zhang YY, Wang J, Wang Z et al. A gRNA-tRNA array for CRISPR-Cas9 based rapid multiplexed genome editing in *Saccharomyces cerevisiae*. *Nat Commun* 2019;**10**:1053.
- Zhao Y, Li L, Zheng G et al. CRISPR/dCas9-mediated multiplex gene repression in *Streptomyces*. *Biotechnol J* 2018;**13**:1800121.