

Measurement of single nanoparticle anisotropy by laser induced optical alignment and Rayleigh scattering for determining particle morphology

Rademacher, Markus; Gosling, Jonathan; Pontin, Antonio; Toroš, Marko; Mulder, Jence T.; Houtepen, Arjan J.; Barker, P. F.

DOI

[10.1063/5.0128606](https://doi.org/10.1063/5.0128606)

Publication date

2022

Document Version

Final published version

Published in

Applied Physics Letters

Citation (APA)

Rademacher, M., Gosling, J., Pontin, A., Toroš, M., Mulder, J. T., Houtepen, A. J., & Barker, P. F. (2022). Measurement of single nanoparticle anisotropy by laser induced optical alignment and Rayleigh scattering for determining particle morphology. *Applied Physics Letters*, 121(22), Article 221102. <https://doi.org/10.1063/5.0128606>

Important note

To cite this publication, please use the final published version (if applicable).
Please check the document version above.

Copyright

Other than for strictly personal use, it is not permitted to download, forward or distribute the text or part of it, without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license such as Creative Commons.

Takedown policy

Please contact us and provide details if you believe this document breaches copyrights.
We will remove access to the work immediately and investigate your claim.

Measurement of single nanoparticle anisotropy by laser induced optical alignment and Rayleigh scattering for determining particle morphology

Cite as: Appl. Phys. Lett. **121**, 221102 (2022); <https://doi.org/10.1063/5.0128606>

Submitted: 29 September 2022 • Accepted: 11 November 2022 • Published Online: 28 November 2022

Published open access through an agreement with JISC Collections

 Markus Rademacher,  Jonathan Gosling,  Antonio Pontin, et al.



View Online



Export Citation



CrossMark

ARTICLES YOU MAY BE INTERESTED IN

[Generation and manipulation of multi-twisted beams via azimuthal shift factors](#)

Applied Physics Letters **121**, 221103 (2022); <https://doi.org/10.1063/5.0123142>

[Development of compact entangled photon-pair sources for satellites](#)

Applied Physics Letters **121**, 220503 (2022); <https://doi.org/10.1063/5.0109702>

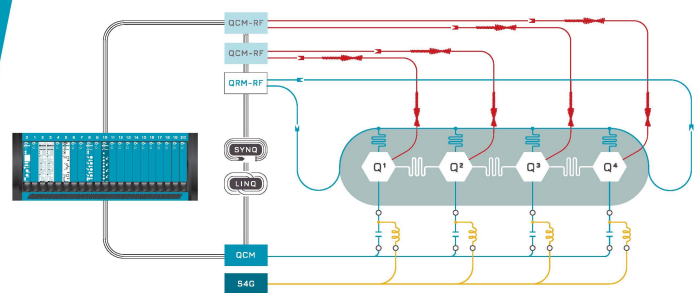
[Two-dimensional ferromagnetic materials: From materials to devices](#)

Applied Physics Letters **121**, 220501 (2022); <https://doi.org/10.1063/5.0130037>



Integrates all
Instrumentation + Software
for Control and Readout of
Superconducting Qubits

[visit our website >](#)



Measurement of single nanoparticle anisotropy by laser induced optical alignment and Rayleigh scattering for determining particle morphology

Cite as: Appl. Phys. Lett. **121**, 221102 (2022); doi: [10.1063/5.0128606](https://doi.org/10.1063/5.0128606)

Submitted: 29 September 2022 · Accepted: 11 November 2022 ·

Published Online: 28 November 2022



View Online



Export Citation



CrossMark

Markus Rademacher,¹ Jonathan Gosling,¹ Antonio Pontin,¹ Marko Toroš,² Jence T. Mulder,³ Arjan J. Houtepen,³ and P. F. Barker^{1,a)}

AFFILIATIONS

¹Department of Physics and Astronomy, University College London, London WC1E 6BT, United Kingdom

²School of Physics and Astronomy, University of Glasgow, Glasgow G12 8QQ, United Kingdom

³Optoelectronic Materials Section, Faculty of Applied Sciences, Delft University of Technology, 2629 HZ Delft, The Netherlands

^{a)}Author to whom correspondence should be addressed: p.barker@ucl.ac.uk

ABSTRACT

We demonstrate the measurement of nanoparticle anisotropy by angularly resolved Rayleigh scattering of single optical levitated particles that are oriented in space via the trapping light in vacuum. This technique is applied to a range of particle geometries from perfect spherical nanodroplets to octahedral nanocrystals. We show that this method can resolve shape differences down to a few nanometers and be applied in both low-damping environments, as demonstrated here, and in traditional overdamped fluids used in optical tweezers.

© 2022 Author(s). All article content, except where otherwise noted, is licensed under a Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>). <https://doi.org/10.1063/5.0128606>

The measurement of nanoparticle morphology is vitally important for aerosol science,^{1–3} nanoparticle production,⁴ and even identification of airborne viruses.^{5–8} For example, rapid measurement of the morphology of nanoparticles is of significance to determine their antibacterial activity and toxicity.⁹ Their geometry also determines their optical properties, which is important for cancer diagnosis and imaging.¹⁰ This also governs their specific absorption rate for *in vivo* applications of magnetic nanoparticle hyperthermia for cancer treatment.^{11–13} Although accurate measurements can be carried out using electron microscopy or x-ray diffraction, light scattering methods can rapidly characterize single particles and even large ensembles in a solution. However, as nanoparticles are often subject to translational and rotational Brownian motion, only the averaged properties of the particles can be measured. While this is sometimes sufficient, scattering from an aligned particle contains much more information when strongly suppressing Brownian motion.

More recently, the field of levitated optomechanics has greatly progressed. It offers enhanced control over the translational and rotational motion of single isolated nanoparticles, and the movement can be strongly damped, suppressing Brownian motion. Particles are held in optical, electric, or magnetic traps. In optical traps, the particles can be localized in position down to a few picometers.^{14–17} This control

has allowed these systems to be brought into the quantum regime when cooling to the quantum ground state that was recently demonstrated.^{15–17} Such experiments have paved the way for the next generation of quantum applications ranging from quantum-limited sensing of gravity¹⁸ to tests of the large-scale limit of quantum mechanics.¹⁹ Knowledge of the shape, refractive index, and other properties of the levitated nanoparticles is critical for current and future quantum experiments.^{20,21} These include quantum metrology, which aims to search for new physics beyond the standard model²² and tests of quantum mechanics in this new mesoscopic regime.²³

A long-standing problem in levitated optomechanics is obtaining detailed information on the structure and geometry of the levitated nanometer-sized objects.²⁴ A basic procedure for determining the nanoparticle's shape has been developed based on linewidth measurements of the directional damping values for different spherical nanoparticle cluster configurations.²⁵ While this technique is helpful, it is not very sensitive to small changes in the particle shape within the nanometer range and cannot be used in the over-damped regime where many optical traps operate.

Despite the importance of the laser scattering behavior of optically trapped nanoparticles to levitated optomechanics,^{26,27} the measurement of the scattering pattern of an optically trapped object has

not been used to determine the shape and geometry of a single particle. In this study, we determine single nanoparticle morphology using laser-induced optical alignment of levitated nanoparticles coupled with angularly resolved Rayleigh scattering. We present the underlying laser Rayleigh scattering theory on which the characterization technique is based and demonstrate its application to a range of trapped and oriented symmetric top nanoparticles with different morphologies. These results are compared with numerical simulations of the particle alignment. We demonstrate that this method effectively determines the geometry of the optically levitated particle, realizing a new tool for studying isolated nanoparticles.

A key ingredient in this method is the ability to align the nanoparticle to the laboratory reference frame via the optical torque induced by the linearly polarized trapping light. This feature greatly simplifies the treatment of the light scattering and increases its sensitivity to particle shape because it allows us to minimize the orientational averaging.

We consider light scattered from a linearly polarized laser along the y -axis as shown in Fig. 1, and we measure the vertically polarized light intensity I_V scattered by our nanoparticle along the x -axis. We treat the sub-wavelength nanoparticle as a dipole scatterer.²⁸ The incoming electric field is horizontally polarized with $E_{Iy} = \sqrt{2 \frac{I_y}{c\epsilon_0}}$, and the scattered intensity of the light along the x -axis is given by

$$I_V = \frac{\pi^2 c E_{Iy}^2 \chi_{yy}^2}{2 \lambda^4 r^2 \epsilon_0}, \quad (1)$$

where c is the speed of light and ϵ_0 is the vacuum permittivity.

To determine the asymmetry and the geometry of an optically trapped nanostructure from its Rayleigh scattering,²⁸ we determine the susceptibility χ_{yy} for any angle of the particle with respect to the observation direction as determined by the polarization and intensity of the levitation field. The electric susceptibility tensor χ of the nanoparticle in the body frame, when aligned with the laboratory frame, is given by

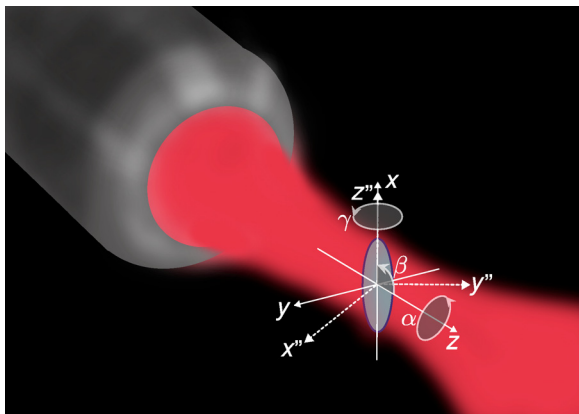


FIG. 1. Schematic representation of an ellipsoidal shaped nanoparticle trapped in an optical tweezer. The three Euler angles (α , β , and γ) and three centers of mass coordinates are shown in the convention used throughout the work (laboratory frames: x , y , and z ; body frames: x'' , y'' , and z'').

$$\chi_0 = \begin{pmatrix} \chi_0^{zz} & 0 & 0 \\ 0 & \chi_0^{yy} & 0 \\ 0 & 0 & \chi_0^{xx} \end{pmatrix}. \quad (2)$$

To predict the scattered light in our observation direction for any orientation of the nanoparticle, we calculate the susceptibility tensor by

$$\chi = R \chi_0 R^T. \quad (3)$$

The angles shown in Fig. 1 are the Euler angles, which represent the three rotations of the principal nanoparticle axes α , β , and γ , where R is the rotation matrix using the z - y' - z'' convention²⁹ and is given by $R_z(\alpha)R_y(\beta)R_z(\gamma)$ as shown in Appendix B in the [supplementary material](#). The rotation matrix R transforms the susceptibility matrix from the body frame aligned with the lab frame to the orientation determined by the trapping laser. Combining Eqs. (1)–(3), we can extract χ_0 by placing the nanoparticle in different orientations.

To illustrate how the scattered light changes for a non-spherical nanoparticle as a function of its orientation, we consider a levitated prolate spheroid for which an analytical solution exists for the susceptibility. The particle is an ellipsoid, as shown in Fig. 1, where two out of the three radii (r_x , r_y , r_z) are of the same length, and the diagonal components of the susceptibility tensor are given as³⁰

$$\chi_0^{\{zz,yy,xx\}} = \frac{4\pi r_x r_y r_z}{\epsilon_0 V} \frac{(\epsilon_l - \epsilon_m)}{3L_{\{1,2,3\}}(\epsilon_l - \epsilon_m) + 3\epsilon_m}, \quad (4)$$

where $L_1 = (1 - e^2) \left(\frac{\log(\frac{e+1}{1-e})}{2e} - 1 \right) / e^2$, $e = \sqrt{1 - \frac{r_y^2}{r_x^2}}$, and that $L_2 = L_3$ with $L_1 + L_2 + L_3 = 1$.

Using Eqs. (1)–(4), we calculate the intensity modulation as the angle α is varied over 2π with $\beta = \pi/2$ and $\gamma = 0$ fixed for different nano-ellipsoids with different degrees of asymmetry as defined by the ratio of the lengths r_x/r_y and where $r_y = r_z$. γ can be set to any value for axially symmetric particles without changing the results. These calculations are shown for three ellipsoidal nanoparticles with different asymmetry ratios in Fig. 2(a). We define the total intensity modulation depth as $\min(I_V)/\max(I_V) = I_V(\chi_0)/I_V(\chi_{\alpha=\pi/2})$, which is plotted as a function of the asymmetry parameter r_x/r_y for different ellipsoids (prolate spheroids) in Fig. 2(b). We consider the angular Brownian motion of the particle when it is optically aligned. This alignment depends on the optical field, the susceptibility, and the gas temperature. This process acts to reduce the observed asymmetry in the angular scattering. Computationally we first calculate the full scattering pattern of the particle and perform averaging on this pattern using the calculated angular dynamics of the optically trapped particle assuming a harmonic trapped motion and applying the equipartition theorem.^{14,31} We apply this averaging process to the scattering patterns shown in Fig. 2 and the solid green lines in Fig. 4. For all our calculations, we use the laboratory temperature of 295 K and the vacuum pressure of 5 mbar. The figures show that the larger the asymmetry parameter, the larger the intensity modulation depth of the vertically polarized scattered light. The modulation visible in Figs. 2(a) and 2(b) is size independent and only depends on the asymmetry ratio of the nanoparticle. This dependence on particle asymmetry is non-linear and has enhanced sensitivity when small changes in the asymmetry

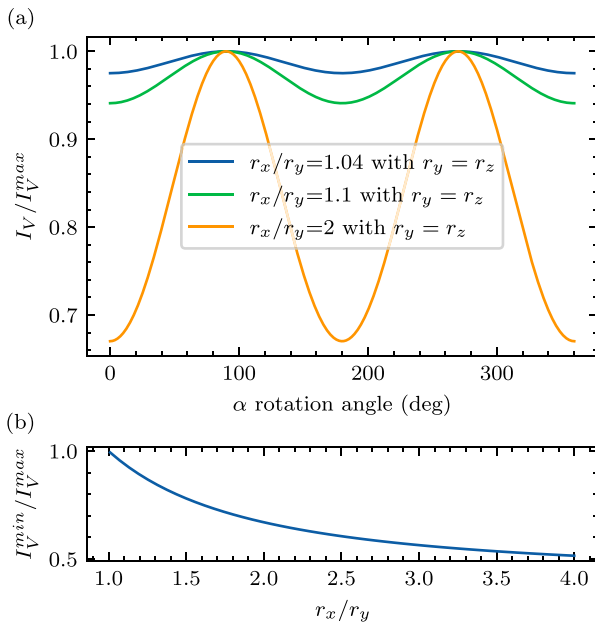


FIG. 2. (a) Relative scattering intensity of the vertical polarized light scattered by the ellipsoid $I_V/\max(I_V)$ vs the rotation angle α depicted in Fig. 1. The blue line represents an ellipsoid with 4% asymmetry. The green one shows a 10% difference for the directional radii, and the orange one has 100% asymmetry. (b) Total modulation depth of the scattering intensity of the vertical polarized light scattered by an ellipsoid $\min(I_V)/\max(I_V)$ vs the asymmetry parameter of different radii of a prolate spheroid r_x/r_y , where $r_y = r_z$.

ratio exist. Similar behavior is seen for other morphologies. When an analytical description of the susceptibility tensor χ_0 is not available, we calculate it numerically as described in Appendix A in the [supplementary material](#).

We demonstrate this angularly resolved Rayleigh scattering on different nanoparticles and verify its sensitivity to asymmetry. Figure 3 shows the experimental setup that was used. Here, the trapping light ($\lambda_{\text{trap}} = 1064$ nm, $P_{\text{trap}} = 450$ mW) was polarized by a polarizing beam splitter (PBS) and passed through a $\frac{\lambda}{2}$ -waveplate, which allowed us to rotate the polarization. Linear polarization rotation was achieved by mounting it in a remote piezo-controlled kinematic rotary mount.

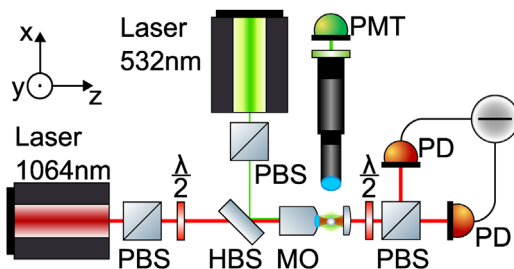


FIG. 3. Schematic of the experimental setup for characterizing the shape and geometry of levitated nanoparticles by analyzing their scattered light modulation. $\lambda_{\text{trap}} = 1064$ nm laser light in red and $\lambda_{\text{probe}} = 532$ nm probing laser light, in green.

A green probe laser ($\lambda_{\text{probe}} = 532$ nm, $P_{\text{probe}} = 5$ mW) of fixed linear polarization was Rayleigh scattered from the aligned particle. This laser illuminated the trapped nanoparticle by coupling it into the trapping beam path via a harmonic beam splitter (HBS). Both co-linear beams were passed into a microscope objective ($\text{NA}_{\text{trap}} = 0.8$) and focused. The strongly focused 1064 nm beam trapped the particle, and a lens collected the transmitted light ($\text{NA}_{\text{coll}} = 0.77$). The light was subsequently passed through a PBS where the two arms are aligned onto two separate photodiodes implementing a balanced detection scheme. We used a $\frac{\lambda}{2}$ -wave plate to balance the power on the two photodiodes. This detection scheme allows us to record a time trace of all six degrees of freedom of the levitated nanoparticle since the interaction of the motion of the nanoparticle with the trapping light changes the polarization state and direction of the laser light.^{25,32–35} The green probe beam was linearly polarized along the y-direction of Fig. 1. This beam was apertured when coupled into the microscope objective to create a much larger focal spot at the trapped nanoparticle. The larger focus allows us to reduce the light intensity on the particle so that there are no significant optical forces on it, and the light provides uniform illumination of the particle. The 532 nm scattered light from the nanoparticle was collected along the x-axis by a lens system with an effective focal length of 3.5 cm. The collected light passed through a 532 nm narrow bandpass filter and was detected on a photomultiplier tube (PMT).

The nanoparticles studied were loaded into the chamber by creating an aerosol using an asthma nebulizer.^{36,37} They were trapped at atmospheric pressure, and the pressure was then reduced to 5 mbar. The vacuum environment allowed us to compare our results with the dynamical motion of the particle in the under-damped regime. A time trace of the balanced detector for rotation was recorded as shown in Fig. 3. The power spectral density (PSD) of this time trace was calculated and used to confirm the optical trapping of non-spherical particles and the alignment via their librational motion. This PSD also shows the translational motion of the particle in the trap. We record the scattered light from a fixed direction as a function of the particle's orientation in free space. At the same time, we change the particle's orientation by rotating the trapping beam linear polarization. To do this, we rotate the half-wave plate in 120 equal increments over 360°. This waveplate rotation turns the particle 720°. The PMT voltage allows one to determine the angularly resolved Rayleigh scattering pattern.

The angularly resolved scattering and the PSDs for a range of levitated nanoparticles are shown in Fig. 4. The data include nanodumbbells of silica, octahedrons of yttrium lithium fluoride, spheroidal nanoparticles of silica, and spherical methanol nanodroplets. The normalized scattered light intensity data in Figs. 4(a), 4(c), 4(e), and 4(g) correspond to the power spectral densities shown in Figs. 4(b), 4(d), 4(f), and 4(h).

The dataset in Figs. 4(a) and 4(b) represents a silica dumbbell consisting of two $r = 71 \pm 2$ nm spheres stuck together. The dataset in Fig. 4(a) shows the distinct modulation of the scattering intensity upon turning the nanodumbbell in the 532 nm detection beam. This modulation agrees well with the calculation shown in the solid green line in Fig. 4(a). We also consider the angular Brownian motion for this calculated scattering modulation, which reduces the overall contrast in the modulation depth. We can distinctly distinguish the librational motion of the nanodumbbell from the PSD of the rotational motion in Fig. 4(b).

Figure 4(c) is the scattering modulation recorded for an octahedron-shaped nanoparticle with a short side length of $a = (77 \pm 5)$ nm and a height of $h = (113.5 \pm 9)$ nm (Appendix A in the supplementary material). This modulation also agrees with the simulations shown as a solid green line. The power spectral density for the motion of the octahedron-shaped structure is shown in Fig. 4(d). The librational peak at ~ 1 MHz is higher in frequency than the librational peaks of the nanodumbbell sample in Fig. 4(b). This difference is consistent with the simulations of the optical trapping setup.

The dataset shown in Figs. 4(e) and 4(f) represents an optically levitated pure methanol droplet. This droplet is a near-perfect sphere due to high tension forces. Within the uncertainty of our measurements, this is confirmed by the scattering dataset in Fig. 4(e), which shows no consistent modulation of the scattered intensity. In addition, the power spectral density shown in Fig. 4(f) exhibits no librational peaks in their spectrum, which is also a sign of their sphericity.

Finally, we present in Fig. 4(g) the corresponding scattering datasets, which shows a smaller modulation of the scattering intensity, compared to Figs. 4(a) and 4(c) for commercially available silica nanospheres ($r = 71 \pm 2$ nm from microparticles GmbH) while being rotated in the 532 nm illumination field. Nevertheless, we cannot identify any distinct peak of the librational motion in the PSD shown in Fig. 4(h). However, we can see an increased noise background in the frequency range between the lowest and second highest translational motion peak, which is not seen in any particle morphology pointing to a lower frequency in the optically induced alignment of the silica particles. The scattering modulation in Fig. 4(g) indicates that the nanosphere deviates from sphericity by 15% with an asymmetry ratio of $r_x/r_y = 0.85$. This divergence is equivalent to a change of 10 nm in

radius between one axis and another. This difference is calculated by fitting the experimental scattering data with Eq. (1). The procedure was repeated for several trapped nanospheres, and all show some asphericity derived from the modulation in their angular scattering pattern. For clarity, these are not shown in Fig. 4(g) but range from 0.84 to 0.86 in the asymmetry ratio. From the signal-to-noise ratio demonstrated ($\Delta I_V/I_V^{\max} = 0.061$) in Fig. 4(g) in conjunction with the nonlinear function in Fig. 2(b), we could differentiate changes between r_x and r_y of approximately 5 nm for a nanosphere of $r = 71$ nm. The ability to measure this small difference using the scattering method indicates its utility for fast measurements of the particle morphology. Such a small asphericity cannot be determined from the PSD as no librational peak can be observed at our pressures. Additionally, recent work¹⁴ in our laboratory suggests that most of the time, when loading spherical nanoparticles, we do not trap perfect spheres but rather trap nanoparticles that have the shape of an ellipsoid with a small degree of anisotropy.

In this paper, we showed that the modulation in the angularly resolved scattering of an optically suspended nanoparticle is a valuable tool to characterize small asymmetries in single nanoparticles down to the percent level. This method was demonstrated on a range of non-spherical nanoparticles and is possible due to the control in the alignment of the nanoparticle in the optical field and the strong asymmetry in the angular scattering when aligned. Although we have demonstrated this technique in an under-damped gas environment, it is also applicable for use with traditional over-damped tweezers experiments carried out in liquids for biological samples. In future experiments, cooling the angular motion degree of freedom of the sample using feedback polarization control of the trapping light or by feedback

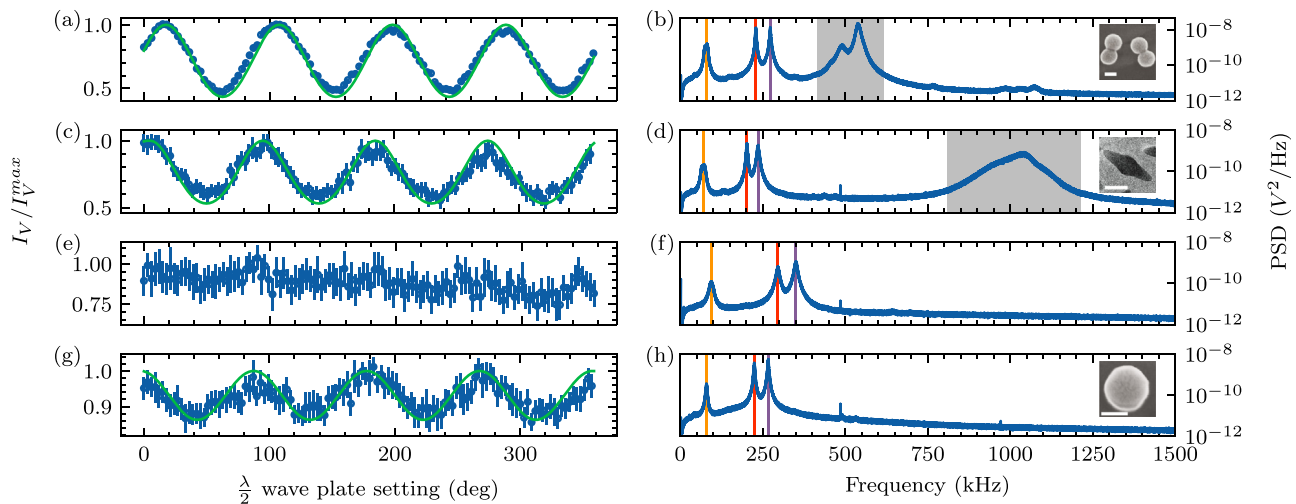


FIG. 4. Experimental scattering and time trace data: (a), (c), (e), and (g) normalized scattering intensity data $I_V/I_V^{\max}$ vs $\lambda/2$ wave plate setting, i.e., half the particle orientation angle α in free space. (b), (d), (f), and (h) Power spectral densities of the time traces of the nanoparticle's motion recorded in the PBS-detection setup. The orange, red, and purple lines represent the oscillation frequency of the z , x , and y degrees of freedom, respectively. The gray shaded area in (b) and (d) denotes the α and β degrees of freedom, which are non-linearly coupled by the γ motion.³⁸ All datasets in (b), (d), (f), and (h) correspond to the same samples used for the data in (a), (c), (e), and (g). (a) and (b) Data for a nanometer-sized dumbbell structure of two nanospheres attached. The scale bar in insets in (b), (d), and (h) represents 100 nm. The inset in (b) shows a scanning electron micrograph (SEM) of the nanodumbbells. The dataset in (c) and (d) corresponds to a nanometer-sized bi-pyramidal structure with a short length of 77 ± 5 nm and a long side of 227 ± 18 nm. The inset in (d) displays a SEM of the octahedron structure. (e) and (f) The data collected for a liquid methanol droplet suspended in the optical trap. (g) and (h) The data collected for a nanometer-sized sphere from the same sample ($r = 71 \pm 2$ nm from microparticles GmbH). The inset in (h) exhibits a SEM of the nanoparticle.

optical torque control³⁹ or by cavity cooling¹⁴ would reduce the effects of angular Brownian motion, which would enhance the modulation contrast and increase our ability to detect even more minor nanoparticle asymmetries. Trapping and cooling based on elliptically polarized light rather than the linearly polarized light here would allow this technique to be applied to other morphologies, including asymmetric tops, i.e., arbitrary shape characterization of nanoparticles.

In addition, further improvements in the detection signal-to-noise ratio would allow us to differentiate length differences as small as 0.4 nm. We expect this method could be used with other scattering and imaging modalities, such as super-resolution imaging, as part of a correlated imaging approach⁴⁰ to nanoparticle imaging and analysis. Another exciting application for this technique is characterization of compound nanoparticles with a complex internal structure.⁴¹ For example, particles used for drug delivery have anisotropic scattering due to internal polycrystal structure,⁴² shape-induced depolarization,⁴³ and filling with dopants.⁴⁴ Our technique holds promise for distinguishing these contributions, which are important for using nanoparticles in biophotonic applications. While the modulation depth shown in Fig. 4 measures optical anisotropy of the aligned nanoparticle, a measurement of the absolute amplitude provides an additional step toward determining the particle morphology. Furthermore, this technique is not limited to the Rayleigh regime as discussed here but can be further extended to larger particles using a multi-polar Mie scattering theory.³⁰ Larger particles up to 32 μ m have been shown to levitate and align using counter-propagating and gravito-optical traps.⁴⁵ This work also has important implications for more fundamental studies in levitated optomechanics, where a detailed knowledge of particle morphology is required to compare experiments with theoretical predictions.⁴⁶

See the [supplementary material](#) for detailed information for the calculation of the susceptibility tensor for arbitrary shaped particles, the definition of the three rotation matrices, and the colloidal synthesis of YLF nanocrystals.

M.R., J.G., A.P., and P.F.B. acknowledge funding from the EPSRC via Grant Nos. EP/S000267/1 and EP/W029626/1. M.R., J.G., A.P., J.T.M., A.J.H., and P.F.B. acknowledge funding from the H2020-EU.1.2.1 TEQ Project through Grant Agreement ID 766900. M.T. acknowledges funding by the Leverhulme Trust (No. RPG-2020-197).

AUTHOR DECLARATIONS

Conflict of Interest

The authors have no conflicts to disclose.

Author Contributions

Markus Rademacher: Conceptualization (equal); Data curation (equal); Formal analysis (equal); Investigation (equal); Methodology (equal); Project administration (equal); Resources (equal); Software (equal); Validation (equal); Visualization (equal); Writing – original draft (equal); Writing – review & editing (equal). **Jonathan Gosling:** Formal analysis (equal); Investigation (equal); Methodology (equal); Resources (equal); Validation (equal); Visualization (supporting); Writing – original draft (supporting); Writing review & editing

(equal). **Antonio Pontin:** Conceptualization (supporting); Investigation (equal); Methodology (equal); Resources (equal); Software (equal); Supervision (equal); Validation (equal); Writing review & editing (equal). **Marko Toroš:** Investigation (equal); Methodology (equal); Software (equal); Supervision (equal); Validation (equal); Writing review & editing (equal). **Jence T. Mulder:** Resources (lead); Writing review & editing (equal). **Arjan J. Houtepen:** Funding acquisition (equal); Resources (lead); Writing review & editing (equal). **P. F. Barker:** Conceptualization (equal); Formal analysis (equal); Funding acquisition (equal); Investigation (equal); Methodology (equal); Project administration (equal); Resources (equal); Software (equal); Supervision (equal); Validation (equal); Visualization (equal); Writing – original draft (equal); Writing – review & editing (equal).

DATA AVAILABILITY

The data that support the findings of this study are available from the corresponding author upon reasonable request.

REFERENCES

- W. Zeller, *Aerosol. Sci. Technol.* **4**, 45 (1985).
- C. Xiong and S. K. Friedlander, *Proc. Natl. Acad. Sci. U. S. A.* **98**, 11851 (2001).
- A. Karlsson, S. Török, A. Roth, and P.-E. Bengtsson, *J. Aerosol Sci.* **159**, 105828 (2022).
- T. K. Sau and A. L. Rogach, *Adv. Mater.* **22**, 1781 (2010).
- H. R. Gelderblom, *Medical Microbiology*, 4th ed. (University of Texas, Galveston, TX, 1996).
- F. V. Ignatovich, D. Topham, and L. Novotny, *IEEE J. Sel. Top. Quantum Electron.* **12**, 1292 (2006).
- M. Pan, J. Lednický, and C.-Y. Wu, *J. Appl. Microbiol.* **127**, 1596 (2019).
- J. Lukose, S. Chidangil, and S. D. George, *Biosens. Bioelectron.* **178**, 113004 (2021).
- A. Sirelkhatim, S. Mahmud, A. Seenii, N. H. M. Kaus, L. C. Ann, S. K. M. Bakhori, H. Hasan, and D. Mohamad, *Nano-Micro Lett.* **7**, 219 (2015).
- X. Huang and M. A. El-Sayed, *J. Adv. Res.* **1**, 13 (2010).
- Clinical Applications of Magnetic Nanoparticles: Design to Diagnosis/Manufacturing to Medicine*, edited by N. T. K. Thanh, 1st ed. (CRC Press, Boca Raton, 2018).
- S. K. Sharma, N. Shrivastava, F. Rossi, L. D. Tung, and N. T. K. Thanh, *Nano Today* **29**, 100795 (2019).
- J. Hilger, *Int. J. Hyperthermia* **29**, 828 (2013).
- A. Pontin, H. Fu, M. Toroš, T. S. Monteiro, and P. F. Barker, “Simultaneous cooling of all six degrees of freedom of an optically levitated nanoparticle by elliptic coherent scattering,” [arXiv:2205.10193 \[quant-ph\]](#) (2022).
- U. Delić, M. Reisenbauer, K. Dare, D. Grass, V. Vuletić, N. Kiesel, and M. Aspelmeyer, *Science* **367**, 892 (2020).
- L. Magrini, P. Rosenzweig, C. Bach, A. Deutschmann-Olek, S. G. Hofer, S. Hong, N. Kiesel, A. Kugi, and M. Aspelmeyer, *Nature* **595**, 373 (2021).
- F. Tebbenjohanns, M. L. Mattana, M. Rossi, M. Frimmer, and L. Novotny, *Nature* **595**, 378 (2021).
- M. Rademacher, J. Millen, and Y. L. Li, *Adv. Opt. Technol.* **9**, 227 (2020).
- A. Bassi, K. Lochan, S. Satin, T. P. Singh, and H. Ulbricht, *Rev. Mod. Phys.* **85**, 471 (2013).
- C. Gonzalez-Ballester, M. Aspelmeyer, L. Novotny, R. Quidant, and O. Romero-Isart, *Science* **374**, eabg3027 (2021).
- B. A. Stickler, K. Hornberger, and M. S. Kim, *Nat. Rev. Phys.* **3**, 589 (2021).
- D. C. Moore and A. A. Geraci, *Quantum Sci. Technol.* **6**, 014008 (2021).
- M. Toroš, S. Bose, and P. F. Barker, *Phys. Rev. Res.* **3**, 033218 (2021).
- F. van der Laan, R. Reimann, M. Doderer, E. Hebestreit, R. Diehl, M. Frimmer, D. Windey, F. Tebbenjohanns, and L. Novotny, *Phys. Rev. Lett.* **126**, 159901 (2021).

- ²⁵J. Ahn, Z. Xu, J. Bang, Y.-H. Deng, T. M. Hoang, Q. Han, R.-M. Ma, and T. Li, *Phys. Rev. Lett.* **121**, 033603 (2018).
- ²⁶F. Tebbenjohanns, M. Frimmer, and L. Novotny, *Phys. Rev. A* **100**, 043821 (2019).
- ²⁷C.-H. Li, J. Jing, L.-M. Zhou, Z.-H. Fu, X.-W. Gao, N. Li, X.-F. Chen, and H.-Z. Hu, *Opt. Lett.* **46**, 4614 (2021).
- ²⁸R. B. Miles, W. R. Lempert, and J. N. Forkey, *Meas. Sci. Technol.* **12**, R33 (2001).
- ²⁹G. Arfken, *Mathematical Methods for Physicists*, 3rd ed. (Academic Press, Inc., San Diego, CA, 1993).
- ³⁰C. F. Bohren and D. R. Huffman, *Absorption and Scattering of Light by Small Particles*, 1st ed. (Wiley, NY, 1998).
- ³¹Y. Arita, S. H. Simpson, P. Zemánek, and K. Dholakia, *Sci. Adv.* **6**, eaaz9858 (2020).
- ³²R. Reimann, M. Doderer, E. Hebestreit, R. Diehl, M. Frimmer, D. Windey, F. Tebbenjohanns, and L. Novotny, *Phys. Rev. Lett.* **121**, 033602 (2018).
- ³³F. Monteiro, S. Ghosh, E. C. van Assendelft, and D. C. Moore, *Phys. Rev. A* **97**, 051802 (2018).
- ³⁴F. Tebbenjohanns, A. Militaru, A. Norrman, F. van der Laan, L. Novotny, and M. Frimmer, *Phys. Rev. A* **105**, 053504 (2022).
- ³⁵M. Toroš, M. Rashid, and H. Ulbricht, *Phys. Rev. A* **98**, 053803 (2018).
- ³⁶D. R. Burnham and D. McGloin, *Opt. Express* **14**, 4175 (2006).
- ³⁷J. Millen, T. S. Monteiro, R. Pettit, and A. N. Vamivakas, *Rep. Prog. Phys.* **83**, 026401 (2020).
- ³⁸T. Seberson and F. Robicheaux, *Phys. Rev. A* **99**, 013821 (2019).
- ³⁹J. Bang, T. Seberson, P. Ju, J. Ahn, Z. Xu, X. Gao, F. Robicheaux, and T. Li, *Phys. Rev. Res.* **2**, 043054 (2020).
- ⁴⁰C. L. Fonta and B. M. Humbel, *Archiv. Biochem. Biophys.* **581**, 98 (2015).
- ⁴¹G. Chen, I. Roy, C. Yang, and P. N. Prasad, *Chem. Rev.* **116**, 2826 (2016).
- ⁴²R. E. Noskov, I. I. Shishkin, H. Barhom, and P. Ginzburg, *Nanoscale* **10**, 21031 (2018).
- ⁴³H. Bahrom, A. A. Goncharenko, L. I. Fatkhutdinova, O. O. Peltek, A. R. Muslimov, O. Y. Koval, I. E. Eliseev, A. Manchev, D. Gorin, I. I. Shishkin, R. E. Noskov, A. S. Timin, P. Ginzburg, and M. V. Zyuzin, *ACS Sustainable Chem. Eng.* **7**, 19142 (2019).
- ⁴⁴R. E. Noskov, A. Machnev, I. I. Shishkin, M. V. Novoselova, A. V. Gayer, A. A. Ezhov, E. A. Shirshin, S. V. German, I. D. Rukhlenko, S. Fleming, B. N. Khlebtsov, D. A. Gorin, and P. Ginzburg, *Adv. Mater.* **33**, 2008484 (2021).
- ⁴⁵F. Monteiro, S. Ghosh, A. G. Fine, and D. C. Moore, *Phys. Rev. A* **96**, 063841 (2017).
- ⁴⁶C. P. Blakemore, A. D. Rider, S. Roy, A. Fieguth, A. Kawasaki, N. Priel, and G. Gratta, *Phys. Rev. Appl.* **12**, 024037 (2019).