

Design of a parallel plate shearing device for visualization of concentrated suspensions

Shakeel, Ahmad; van Kan, Paul J.M.; Chassagne, Claire

DOI 10.1016/j.measurement.2019.05.101

Publication date 2019 Document Version Accepted author manuscript

Published in Measurement: Journal of the International Measurement Confederation

Citation (APA)

Shakeel, A., van Kan, P. J. M., & Chassagne, C. (2019). Design of a parallel plate shearing device for visualization of concentrated suspensions. *Measurement: Journal of the International Measurement Confederation*, *145*, 391-399. https://doi.org/10.1016/j.measurement.2019.05.101

Important note

To cite this publication, please use the final published version (if applicable). Please check the document version above.

Copyright

Other than for strictly personal use, it is not permitted to download, forward or distribute the text or part of it, without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license such as Creative Commons.

Takedown policy

Please contact us and provide details if you believe this document breaches copyrights. We will remove access to the work immediately and investigate your claim.

1	Design of a Parallel Plate Shearing Device for Visualization of		
2	Concentrated Suspensions		
3	Ahmad Shakeel, ^{1,3*} Paul J. M. van Kan, ² Claire Chassagne ¹		
4			
5	¹ Faculty of Civil Engineering and Geosciences, Department of Hydraulic Engineering,		
6	Delft University of Technology, Stevinweg 1, 2628 CN Delft, The Netherlands		
7	Dent entrensity of Teenhology, Stevniweg 1, 2020 ent Dent, The Realenands		
8 9	² Van Kan Scientific, Keucheniuspad 40, 6535 VR Nijmegen, The Netherlands		
9 10	³ Department of Chemical, Polymer & Composite Materials Engineering, University of		
11	Engineering & Technology, Lahore, KSK Campus, 54890 Pakistan		
12			
13	a.shakeel@tudelft.nl; vankanscientific@xs4all.nl; c.chassagne@tudelft.nl;		
14			
15			
16			
17			
18			
19			
20			
21			
22			
23 24	*Corresponding author		
24 25	Ahmad Shakeel		
26	Faculty of Civil Engineering and Geosciences, Department of Hydraulic Engineering,		
27	Delft University of Technology, Stevinweg 1, 2628 CN Delft, The Netherlands		
28	Email: <u>a.shakeel@tudelft.nl</u>		
29	Tel. +31(0)613091407		
	© 2019 Manuscript version made available under CC-BY-NC-ND 4.0 license		

https://creativecommons.org/licenses/by-nc-nd/4.0/

Abstract

A modified version of the commercially available RheOptiCAD® was developed to visualize the microscopic structural changes occurring in concentrated suspensions, such as the break-up of flocs in clay suspensions, under shearing action. This is made possible by replacing the inverted microscope used in the traditional RheOptiCAD set-up by an upright modular microscope equipped with a CMOS camera and epi-illumination. Our device retains the following features of the previous version of RheOptiCAD[®]: [i] uniaxial translational motion of two parallel plates, [ii] three modes of shear straining, [iii] controlled thermal environment, [iv] vacuum joining of microscopy glass slides. The validation of the new design was done using a model system of un-flocculated and flocculated kaolin suspensions and concentrated natural mud suspension. The results showed that the constructed device is a promising tool for studying, from fundamental and industrial perspectives, the microstructural behaviour of complex suspended systems under controlled thermal and mechanical conditions.

Keywords: Shearing cell, Strain-controlled, Optical microscope, Rheo-optics, Suspensions

52 **1. Introduction**

Complex systems, such as emulsions, colloidal suspensions, gels, polymeric and surfactant 53 solutions, foams and pastes are commonly part of food and non-food products, consumed on 54 daily basis. For these products, the study of their viscoelastic properties is key for industrial 55 purposes. Shearing action combined with parameters like pressure, temperature, ageing time, 56 57 ionic strength or pH can lead to structural changes in such complex systems. One of these changes is demixing or phase separation, driven by either gravitational gradient or 58 thermodynamic forces. Optical rheometry, also known as rheo-optics, is a powerful technique 59 to analyse the behaviour of these complex systems, as it allows the visualization of flow, 60 deformation and restructuration of the system under shear. Combining standard rheological 61 62 measurements with rheo-optics provides an understanding of rheological parameters such as yield strength, viscosity or thixotropy in relation with the observed structural changes. 63

With the progress in advanced microscopic techniques, many research groups have developed 64 devices that combine microscopy and rheology [1-4]. The progress in optical shearing devices 65 66 until 1998 has been summarized by Fuller [5] and Wagner [6] for 2D rheo-optics and 3D rheo-optics, respectively. Later, van der Linden et al. [7] presented the review of rheo-optical 67 devices for food and non-food systems. The rheo-optic devices developed so far are based on 68 69 either an optical device fitted to the commercial or existing shearing device, or alternatively a shearing system developed to combine with a commercial or existing optical device. The 70 71 established accuracy of commercial rheometers led researchers to favour the development of optical techniques fitted to the existing rheometers [8]. However, these devices have some 72 73 drawbacks, e.g. limited field-of-view and low magnification power. Custom-made shearing 74 cells are very interesting alternatives due to the fine tuning and huge flexibility in the materials selection for the surfaces which come into contact with the sample, the possibility to 75 analyse samples under larger deformations and the possibility to create a zero-velocity plane 76

(ZVP) at any position within the cell gap by having top and bottom plates moving in oppositedirections [9, 10].

Several optical techniques have been used in rheo-optical devices, depending on the material under investigation and the observation scale. Different shearing devices like a 4-roll mill [11], controlled strain rheometer [12], parallel plate [13, 14], and others [15] have been used so far to perform deformation under controlled environment. Table 1 summarizes some important details about the already reported rheo-optical systems, in comparison to our designed system.

Rheometer/Shearing Cell	Microscope/Camera	Advantages	Limitations	Ref.
IR-200 Rheometer without transducer, Quartz Cone & Plate/Plate-plate geometry	Upright Optical microscope, CCD camera, halogen white lamp	Rotation can be reversed, mechanism for temperature control	Largest image size 640 x 480 pixels, not suitable at high shear rates	[16]
Custom made parallel plate shear cell	Inverted confocal scanning laser microscope	Higher accuracy from mechanical point of view	Lengthy process for sample preparation, requirement of pair of cassettes for every analysis, difficulty in reproducibility, absence of temperature control mechanism	[17]
 Custom made parallel plate shear cell Anton-Paar MCR 301 Rheometer, Cone & Plate/Plate-plate geometry 	 Upright microscope objective with CCD camera, LED backlight source Inverted microscope objective with CCD camera 	 Allows to shear a very large surface of sample (~70 cm²) Measurement of stress 	 Control of gap variation between the plates was difficult, stress measurement was not possible, possible movement of only bottom plate, no mechanism for temperature control Not suitable for suspensions due to inverted microscope 	[14]
Custom made parallel plate shear cell	Confocal scanning laser microscope, CCD camera	Temperature controlling mechanism	Fixed gap, movement of only one plate	[18]
Custom made cone & plate shear cell	Inverted confocal scanning laser microscope, green He–Ne laser	Movement of both plates, imaging of planes parallel as well as perpendicular to glass plate	Fixed positon of microscope objective in cell, variation in gap width caused by a slight wobbling of the glass plate, not suitable for higher shear rates, no temperature controlling mechanism	[9]
Stress-controlled Bohlin Gemini Rheometer, Cone & Plate geometry	Inverted confocal scanning laser microscope	A wide range of shear strain, up to 50% amplitude	Fixed positon of microscope objective in cell, no recording of images (absence of camera)	[19]
Linear parallel-plate shear cell	Inverted fast confocal microscope	Wall slip prevention by coating the slides with disordered layers of colloid, solvent trap to minimize evaporation	Possible movement of only top plate, no temperature controlling mechanism	[2, 20]
Custom made parallel plate shear cell	Inverted fast-scanning confocal microscope	Teflon sheets to minimize evaporation, synchronization of image acquisition and shearing	Possible movement of only bottom plate, image size 512 × 512 pixels	[21]

85 Table 1: Key characteristics of some of the already reported rheo-optical systems

		action		
Custom made parallel plate shear cell	Inverted microscope, CCD camera	Roughened plates to minimize wall slip, movement of both plates	Image size 1024 x 1024 pixels, only oscillatory mode of shearing	[22-24]
Anton-Paar MCR 301 Rheometer, Cone & Plate geometry	Inverted confocal scanning laser microscope	Sandblasted plates to minimize wall slip, solvent trap to minimize evaporation	Image size 512 × 512 pixels, fixed gap width between cone and plate	[25]
Velocity-controlled Couette rheometer	Inverted fluorescent microscope, CCD camera	Allows to access local velocities up to 1 m/s	Not suitable for suspensions due to inverted microscope	[3]
Stress-controlled rheometer AR2000, Cone & Plate/Plate-plate geometry	Inverted confocal scanning laser microscope	Roughened plates to minimize wall slip, solvent trap to minimize evaporation	Not suitable for suspensions due to inverted microscope	[8, 26]
Custom made parallel plate shear cell	Inverted confocal scanning laser microscope	Temperature controlling mechanism, vacuum joining of glass coverslip with the plate, movement of both plates	No stress measurement, not suitable for suspensions due to inverted microscope	[10, 27]
Custom built constant stress shear cell, Cone & Plate geometry	Inverted fast-scanning confocal microscope	Solvent trap to minimize evaporation, cost-effective, availability of range and resolution of applied stresses through selection of transfer fluid	Absence of temperature controlling mechanism, not suitable for suspensions due to inverted microscope	[28]
Custom made parallel plate shear cell	Inverted confocal scanning laser microscope	Stress measurement, Solvent trap to minimize evaporation, possibility of biaxial shear experiments	Absence of temperature controlling mechanism, gluing of cover slip with the plate, not suitable for suspensions due to inverted microscope	[4]
Anton-Paar MCR 301 Rheometer, Cone & Plate/Plate-plate geometry	Inverted laser scanning confocal microscope	Wide range of applied torque, presence of normal force sensor	Not suitable for suspensions due to inverted microscope, image size 256 × 256 pixels	[29]
Rotational rheometer HAAKE MARS III, Cone & Plate/Plate-plate geometry	Inverted polarized reflected light microscope, CCD camera	Temperature controlling mechanism, in addition to light microscopy RAMAN spectroscopy measurements also available	Not suitable for suspensions due to inverted microscope setup	[30]
Custom made parallel plate shear cell	Upright optical microscope, CCD camera	Temperature controlling mechanism	Possible movement of only bottom plate, suitable for small amplitude deformation	[31-33]
Custom made parallel plate shear cell	Upright optical microscope, CMOS camera, LED light	Suitable for suspensions due to upright microscope, temperature controlling mechanism, vacuum joining of glass coverslip with the plate, movement of both plates, gap variation between two plates is possible from 0-5 mm, CMOS camera instead of CCD which provides reduced blooming and smearing, image size 2592 x 2048 pixels with square pixels for undistorted image, fluorescent marker is not required, microscope objective with large working distance	No mechanism for stress measurement	This study

Many rheo-optical devices developed so far are laboratory models and only some of them have been commercialized. The Cambridge Shearing System [31] produced by Linkam Scientific Instrument Ltd. is the first commercialized system for optical analysis under rotational shearing action. A confocal rheo-scope, combination of confocal microscope and rheometer, has also been reported by Besseling et al. [8] which possessed rotational shearing

mode. However, the size, weight (greater than 10 kg) and cost of this device seem to have 92 93 posed serious issues for its commercialization. The benefit of rotational motion over translation is the generation of an infinite shear and deformation. However, using rotational 94 mode has the technical drawback of a compromised field of vision since the axis of motor 95 rotation comes in line with the axis of observation through microscope objective. Use of 96 translational motion overcomes this disadvantage because the axis of observation is at a right 97 98 angle to the axis of motion during shearing action between e.g. parallel plates [10]. Wu et al. [17] developed a laboratory prototype of shearing cell based on translation mode. It was 99 adapted to the inverted commercial CLSM device and possessed high accuracy from 100 101 mechanical point of view.

Recently, a novel parallel plate rheological device (RheOptiCAD[®]) was designed by CAD 102 103 Instruments and reported by Boitte et al. [10]. With this device a video recording during shearing is obtained from which the structural changes in the samples can be studied (changes 104 105 in floc size, etc.). The device has primarily been designed to be mounted on an inverted 106 microscope. Even though it can be used to visualize readily opaque structures [9], the device is not suited to analyse the optical behaviour of very concentrated clay suspensions (our topic 107 108 of interest) as it makes use of transmitted light. This is why we proceeded to design an alternative set-up that would enable the study of these type of suspensions. 109

In the remainder of the article, we present the modified version of RheOptiCAD[®] with microscope system suited to analyse the behaviour of complex systems, particularly suspensions, under temperature-controlled conditions by shearing parallel-plate geometry. The design and specifications of the selected components are detailed. The set-up has been validated thanks to the pilot experiments. These experiments are done using un-flocculated and flocculated model kaolin suspensions. Additional experiments done on natural mud samples are also presented.

117 2. Design Considerations

The setup of our rheo-optical device makes use of the RheOptiCAD[®], a parallel plate shearing 118 device, commercially available from CAD Instruments (Illiers Combray, France) and 119 introduced in 2012 [10] that can easily be coupled to a microscope. In short, the 120 RheOptiCAD[®] device has been designed to enable optical analysis under a linear shearing 121 122 force. During the measurement, a strain is applied in continuous, step or oscillation mode and a video recording is obtained. The analysis can be done in a temperature-controlled 123 environment. The position of the shear plane in the cell can be adjusted by varying the 124 velocity of the top and bottom plates. 125

Depending on the microscope used, an application-specific set-up involving the RheOptiCAD[®] and this microscope must be designed. In previous applications, the subject of study often was a heavily textured colloidal system like dough [10, 27], with high viscosity and being opaque. Therefore, the RheOptiCAD[®] device was used with an inverted confocal laser scanning microscope and fluorescent markers were used to visualize the changes in dough structure. Our aim is to study concentrated colloidal suspensions under shear accounting for the following constraints:

- The visualization of concentrated (very opaque) colloidal suspensions require the use
 of an upright microscope with an epi-illumination.
- Even in the presence of a sedimentation layer, the sample thickness should be much
 larger than the size of coagulated clay particles (0.1 mm). Therefore, microscope
 objectives with a large working distance (W. D.) should be selected. A simultaneous
 design constraint is that the objectives must have an (adjustable) cover slip correction.

To study clay particles with their "native" charge, addition of fluorescent marker
 molecules is unwanted. Therefore, the microscope should operate with direct
 illumination.

In the following sections we present a complete functional design and technical
implementation, which enables this new application of the RheOptiCAD[®] system.

144 **3. Functional Design**

145 **3.1. Modification of the RheOptiCAD**[®]

The RheOptiCAD® is manufactured by CAD Instruments (Illiers-Combray, France). The 146 instrument is a modular design, contained in a cube-shaped frame with dimensions of about 147 148 20 x 20 x 20 cm and total weight of about 5 kg. The construction is designed for optimal stiffness, which is achieved by using an aluminium construction with rounded corners and a 149 minimal number of bolted joints (Fig. 1). The device can be (re)positioned in a microscopic 150 setup by means of the "handles" that form the device's skeleton. This leaves the relative 151 positioning of the internal parts unchanged. Within the cube-shaped frame, three motorized 152 153 linear-stages are mounted. These stages can move under closed-loop control and are computer-controlled. Furthermore, the temperature of the device can be actively controlled by 154 a Peltier cooler in the bottom plate, (green in Fig. 1) which is also under computer control. 155



157 FIG. 1. 3D image of the RheOptiCAD[®] rheometric device, modified for observation from above. The

microscope objective is lowered into the recessed area on the top plate for observation of particles in the samplecell between top and bottom plate.

The design of the top and bottom plate of the RheOptiCAD[®] was adapted for observation 160 from above. An opening with an oval shape was made in the top plate, to accommodate the 161 microscope objectives (18 mm x 8 mm = 144 mm²), defining the dimensions of the 162 observation window. Also, the vacuum ports in the top plate, necessary for the suction that 163 164 keeps the glass slide in position, were moved to the side. In this way, sufficient horizontal and free vertical movement of the objectives was achieved. In the design presented here, with an 165 166 upright microscope using epi-illumination and observation of reflected light, the displacement of the top plate is limited: the top plate can move with an amplitude of 12 mm, the bottom 167 plate can travel as far as 20 mm. 168

169 **3.2. Microscopy subsystem**

For the upright microscope in the setup, we have found suitable components in the Olympus
BXFM-BX3M modular microscopy/illuminator system [34]. From this modular system we
have chosen the following parts:

- BX-FM-F Focusing unit, 30 mm range, 2 µm resolution
- Märzhäuser MFD motorised Z-stage drive mounted on the fine focusing knob[35]
- BX3M-KMA-S Epi-illuminator with white LED source
- U-5RE-2 5-fold nosepiece, equipped with:
- 177 o LUCPLFLN 20x objective, coverslip correction (CC) 0-2 mm, W. D. 6.6-7.8
 178 mm, NA 0.45
- 179 o LUCPLFLN 40x objective, coverslip correction (CC) 0-2 mm, W. D. 2.7-4.0
 180 mm, NA 0.6

- U-TLU-2 Telan lens-unit (tube lens)
- U-TV1X-2-7 1x video-adapter
- U-CMAD3-1-7 C-mount adapter ring
- 184 An impression of the microscopy subsystem is given in Fig. 2.



FIG. 2. The Olympus BXFM modular microscope with focusing unit (courtesy of Olympus). This microscope
uses an LED white light source for broadband epi-illumination. Observation of the image in reflected light is
done with a CMOS camera on top of the setup (not shown)

189 **3.3. System assembly**

190 The RheOptiCAD[®] skeleton and the microscope subsystem were assembled on a solid 191 aluminium baseplate with adjustable feet for levelling. The focusing unit was attached to a 192 solid stainless-steel rod with a diameter of 32 mm, according to Olympus factory 193 specifications. The height was adjusted and fixed with an aluminium locking plate with six 194 M6 Allen bolts (Fig. 3).



196

FIG. 3. The assembled modified RheOptiCAD[®] setup.

197 **3.4. Mechanical Control**

In the RheOptiCAD[®] device, the shearing of the sample is carried out by uniaxial translation, 198 199 generated by the motion of two parallel (top and bottom) plates. Each plate is driven by its own linear stage (Nanomotion FB-075 with HR4 piezo-electric motor) (NanoMotion, 200 USA)[36]. The translational mode of shearing enables to make use of the rectangular shape of 201 202 commercial microscopy glass slides, which makes the sampling user friendly. A vertical translation with a third motorized stage enables easy separation of the top and bottom plates 203 for sample loading (Fig. 4). The sample is placed on a rectangular microscopy cover slip (24 x 204 60 mm) attached to the bottom plate. 205



207 FIG. 4. Shear movement of the top and bottom plates of the RheOptiCAD[®] device (red arrows). Vertical

208 movement of the bottom plate assembly enables loading of a suspension droplet on the bottom plate as well as

adjustment of the cell height (green arrows)

To ensure the parallelism and planarity of microscopy glass slides, important for better observation and fine-tune controlling of deformation, the glass slides were bound to the aluminium plates (top and bottom) by creating vacuum between them. A path for air was imprinted on the surface of both plates and the output point was connected to the vacuum pump (LaboPort KNF, France) having a minimum pressure limit of 160 mbar [10].

Before measurement, the bottom plate is raised until the sample is in contact with both plates. 215 Subsequently, the distance between the plates (gap width) is adjusted to the desired value, 216 217 thereby enclosing the droplet. All axes of translation are motorized with closed-loop control. 218 The absolute encoders, Renishaw (RGH24Y15D30A), with 10 nm resolution used for the horizontal translations retain their position information. The vertical encoder is recalibrated in 219 the RheOptiCAD[®] software when the top and bottom glass cover slips are replaced [10]. The 220 motor control and recording of the plate's position are done by a modular motor control 221 222 device, (Galil DMC 4040 [37]), which also generates a trigger pulse to start camera image acquisition. 223

Focusing of the objective is performed by the stepper motor in the Märzhäuser MFD motorised Z-stage drive [35]. Manual handling of the focusing knob is replaced by control from the computer user interface. At start-up, the focusing unit is driven to its top position, indicated by a micro switch. In this way, the focusing unit is calibrated.

228 **3.5. Optical image acquisition**

The optical layout of the microscope system is presented in Fig. 5. The LED source, at the back of the setup, provides white light. The source is imaged at the back focal plane of the objective (Köhler illumination). The light is focused on the sample by the objective.



232

FIG. 5. The optical layout of modified RheOptiCAD[®] device. A 1" CMOS camera (see insert) is fitted on the
epi-illumination module of the microscope.

235 We have chosen plan-fluorite objectives for this assembly, for the following reasons:

• planarity of the field of view is essential

colour correction as provided by plan-apochromatic lenses is not necessary for our
 experiments. We want to observe, locate and track particles under shear; therefore, we
 use a monochrome camera for optimal resolution.

availability and affordability of long-working distance objectives with cover slip
 correction

The objectives mentioned above provide a working distance which is suitable for the 0-5 mm gap width of the RheOptiCAD[®]. Furthermore, the coverslip correction ring enables improvement of the image of particles near the bottom of the sample. The objective collects the reflected light from a horizontal slice in the sample. With the infinity-corrected optics chosen for this setup, a sharp image is created by a tube lens between the semi-transparent mirror and the camera. The focal length of this tube lens determines the image size on the camera. The Olympus tube lens and C-mount adapter for a 1" digital camera was chosen.

The camera selected was a USB 3.0 connected monochrome CMOS camera with a 1" target (12.5 x 10 mm) and a resolution of 2592 x 2048 pixels (UI-3180CP-M-GL Rev.2, IDS GmbH, D). This camera is based on a PYTHON5000 CMOS-chip (ON Semiconductor) [38] with some interesting features for microscopy:

- square 4.8 µm x 4.8 µm pixels for undistorted images
- a global shutter with various external trigger options via the control connector, this
 enables synchronization with the motion of the RheOptiCAD[®] plates.
- choice of a Reduced Area-of Interest (ROI) and increased framerate. This opens the option of using the camera at a resolution of 2048 x 2048 pixels, matching the circular (flat) field of view of the microscope. Depending on the USB connection, framerates of more than 80 Hz can be achieved.
- a CMOS sensor, apart from high resolution and high frame rate, provides reduced
 blooming and smearing compared to CCD devices. This is an advantage in particle
 tracking and analysis. Table 2 compares the features of the previous and current
 versions of the RheOptiCAD.
- 265 Table 2: Differences between the two versions of the RheOptiCAD systems

Component name	RheOptiCAD [®] [8]	Our device
Modular upright microscope		Х
Inverted microscope	Х	

CMOS camera with square pixels and reduced blooming and smearing		Х
CCD camera	Х	
Epi-illumination		Х
Vacuum joining of glass slides	Х	Х
Peltier system for temperature control	Х	Х

267 **3.6. Zero velocity plane**

To analyse the structural changes in the sample under shear within the observation window, it is essential for the object to be in the zero-velocity plane (ZVP). The position of this ZVP between the plates in z-axis can be changed just by playing with the velocities of top and bottom plates (see Fig. 6), according to the following equation:

272
$$z_0 = \frac{e}{\frac{v_2}{v_1} + 1}$$
(3)

with z_0 being the position of the ZVP in the z-axis direction, with the origin just underneath the top plate. v_1 and v_2 , respectively, the velocities of the top and bottom plates (mm s⁻¹), and *e* the gap width (mm).



276

FIG. 6. Movement of top and bottom plates along x-axis to change the location of ZVP (red arrows). Green arrows represent

278 the vertical movement (z-axis) of bottom plate for loading the sample and height adjustment

279 **3.7. Temperature control**

Temperature could be controlled within the range of $10-80^{\circ}$ C using a Peltier system TEC-1090 Controller/Peltier Driver (Meerstetter Engineering, CH) (30 mm × 30 mm) mounted on the bottom plate. Water circulation, in the copper part of Peltier system, was used to regulate the sample temperature and a thermistor was fitted to the aluminium body of the bottom plate to monitor the temperature. Heating and cooling rates were optimized within the range of 1-20°C min⁻¹ using the PID controller.

286 **3.8. Control software**

To make a user-friendly shearing device, a software was developed by CAD instruments, comprising of graphic interface, to define and control all the parameters related to the plates like, position of plates, mode of shear strain, velocity, amplitude, experiment time, frequency and gap width. Three modes of shear strain, after positioning the sample and setting the gap width, are available:

- Step-strain for sudden deformation by having amplitude of each plate as variable (both
 plates are mobile)
- 294 2. continuous strain for linear deformation by having amplitude of each plate and295 experimental time as variables (both plates are mobile)
- 3. oscillatory strain for sinusoidal deformation by having amplitude and frequency of oneof the plates as variable (one plate is mobile)

After each experiment, a data file is designed to be created automatically, which consists of the data supplied by the encoders of motorized stages. Different variables like position, time and velocity of top plate, bottom plate and gap width are recorded. Galil DMC-4040 acquisition system allows the recording of data points. Fig. 7 shows the scheme of complete

software configuration and Fig. 8 shows the developed shear cell with an upright opticalmicroscope.



304

FIG. 7. Schematic representation of complete software configuration; PC = personal computer; OM = optical
 microscope. The motor controller triggers continuous camera acquisition when the shear motion starts.



307

308

FIG. 8. Shear cell combined with an upright optical microscope and a camera

309 4. Validation Experiments

310 **4.1. Kaolin suspension**

The validation of the new set-up was performed with kaolin suspensions in water. Unflocculated kaolin suspensions were prepared by dispersing small amount of kaolin (Imerys, England) in distilled water. Two commercial polyelectrolytes, Zetag 4120 (anionic copolymer of acrylamide and acrylic acid) and Zetag 8125 (cationic copolymer of acrylamide and quaternized ammonium cationic monomer) were used to prepare flocculated kaolin suspensions, by simply dispersing small amounts of polyelectrolytes and kaolin in distilled water. For the optical analysis, we used our new optical microscope equipped with the 20x objective, having 0.45 NA and 6.6-7.8 mm of working distance. The gap between the two plates was varied from 100 to 10 μ m for different samples. In all cases, the sample was in contact with both upper and lower plates. This ensures that the samples were deformed instead of displaced. The temperature was maintained at 20°C for all the investigations. The LED light source was used and the images (2592 x 2048 pixels) were recorded in the x-y plane.

Several investigations were performed in oscillation mode using frequencies f between 0.5 324 and 2 Hz and amplitudes A ranging from 0.1 to 0.5 mm for the bottom plate. The value of the 325 amplitude was carefully chosen to make sure that the particles remain in the frame of view 326 327 during the whole experiment. Fig. 9 shows the snapshots from the video recording for the unflocculated kaolin suspension. It can be easily seen from the images that the unmodified 328 kaolin particles are very small and homogeneously dispersed within the water. Under 329 330 oscillatory shear, the particles showed a little bit movement due to the absence of any interactions between the particles. 331



FIG. 9. Images of un-flocculated kaolin suspensions subjected to oscillation at (a) t = 0 s (b) t = 5 s; Gap width = 10 µm; f = 0.5 Hz; A = 0.1 mm. Slight movement of clay particles can be seen due to absence of interactions. Scale bar represents 70 µm Figs. 10 and 11 present the images of the kaolin suspensions containing polyelectrolytes. All these images corroborate the formation of flocculated structures by addition of polyelectrolytes. Fig. 10 displays the break-up of a flocculated structure by the application of

a oscillatory shear for kaolin particles coated with cationic polyelectrolyte. Fig. 11b shows the
stretching of a flocculated structure made of kaolin and anionic polyelectrolyte after 5 s of
oscillatory shear at an amplitude of 0.5 mm and a frequency of 2 Hz.

341



FIG. 10. Images of cationic polyelectrolyte-based kaolin suspensions subjected to oscillation at (a) t = 0 s (b) t = 3 s (c) t = 5s; Gap width = 100 μ m; f = 1 Hz; A = 0.4 mm. Sequence of images shows the break-up of flocs. Scale bar represents 70 μ m



345

FIG. 11. Images of anionic polyelectrolyte-based kaolin suspensions subjected to oscillation at (a) t = 0 s (b) t = 5 s; Gap width = 100 μ m; f = 2 Hz; A = 0.5 mm. Second image presents the stretching of flocs after 5 s. Scale bar represents 70 μ m

348 **4.2. Natural mud sample**

A natural mud sample, collected from Port of Hamburg (Germany), was chosen for the 349 350 investigation. The natural mud samples were placed on the bottom plate of the device and a gap of 100 µm was set to perform the experiments at 20°C. Firstly, the samples were 351 352 subjected to continuous strain by setting the movement of both plates in opposite directions. 353 The images selected from the video recording (which can be found in the online 354 supplementary material) are shown in Fig. 12. Fig. 12b shows the breakage/separation of a bigger flocculated structure into two flocs which further divided into more smaller flocs as 355 356 shown in Fig. 12c.







359 FIG. 12. Images of a natural mud suspension subjected to continuous strain at (a) t = 0 s, (b) t = 5 s, (c) t = 10 s. Sequence of 360 images shows the structural break-up during shear. The black circle shows a small floc that has detached from a bigger one 361 on the right (not in view). Scale bar represents 70 µm

The optical shearing of the samples was also performed in oscillation mode by oscillating the 362 bottom plate at 1 Hz with the amplitude of 0.4 mm. The images selected from the video 363 recording are shown in Figs. 13 and 14. Fig. 13b shows the breakage of a flocculated 364 structure after 3 s of oscillation motion. Fig. 14a shows the presence of a large particle (clay) 365 in the suspension, which displayed the rotational motion during oscillatory shearing as 366 indicated by the arrows in Fig. 14b. This large particle also creates a void during oscillation 367 after 10 s, as shown in Fig. 14c. 368



369

370 371 FIG.13. Images of a natural mud suspension subjected to oscillation at (a) t = 0 s, (b) t = 3 s. The black circle shows the result of the breakage of bigger flocs by the presence of a void (white colour) in the second image. Scale bar represents 70 µm



FIG. 14. Images of a natural mud suspension subjected to oscillation at (a) t = 0 s, (b) t = 5 s, (c) t = 10 s. Arrows shows the direction of rotational motion of particle during shearing. The white area inside the black circle represents a void created by the tumbling motion of the large particle. Scale bar represents 70 μ m

377 **5.** Conclusion

This study presents the modification of an already reported rheo-optical device[10]. The 378 modified system enables the observation of sedimentating and/or concentrated suspensions, 379 by using an upright optical microscope configuration instead of an inverted one as in the 380 original device. An optical microscope with epi-illumination using reflected light was used to 381 get the optical signature of suspensions. Proof-of-concept experiments performed by using 382 un-flocculated and flocculated kaolin suspensions and a natural mud suspension, confirmed 383 the applicability of our device for investigating complex systems. Successive snapshots taken 384 from the video recording of these suspensions under shear revealed the structural changes of 385 386 these systems as a function of the shearing action. The new rheo-optical device will be used, in the future, to perform state of the art research in the field of sediment rheology by linking 387 the qualitative structural break-up and build-up (thixotropy) of mud suspensions observed by 388 the present device to the quantitative rheological measurements obtained from a conventional 389 rheometer. 390

In future applications/research, some technical modifications can be done to optimize the presented device such as: [i] incorporating a device for stress measurement (material's response to the applied deformation), [ii] having a translation motion of plates in y-axis, in addition to x-axis, [iii] adding the possibility to have oscillation of both plates simultaneously, instead of one plate, in oscillation mode, [iv] modifying the surface properties of the glass slides in case of sticky materials which would also enable to study particle-wall interactions, and [5] improving the data analysis software for quantitative investigation.

398 Acknowledgements

The authors would like to thank CAD Instruments, France, in particular Claude Vizcaino, and Olympus, The Netherlands particularly Ronald van Dijk, for their technical contribution. This study was partially supported by funding from the Netherlands Organization for Scientific Research (NWO), project no. 850.13.031.

403 **References**

- 404 [1] P. Schall, D.A. Weitz, F. Spaepen, Structural Rearrangements That Govern Flow in Colloidal 405 Glasses, Science, 318 (2007) 1895-1899.
- 406 [2] N. Koumakis, M. Laurati, S.U. Egelhaaf, J.F. Brady, G. Petekidis, Yielding of Hard-Sphere Glasses
 407 during Start-Up Shear, Phys. Rev. Lett., 108 (2012) 098303.
- 408 [3] J. Goyon, A. Colin, G. Ovarlez, A. Ajdari, L. Bocquet, Spatial cooperativity in soft glassy flows, 409 Nature, 454 (2008) 84-87.
- 410 [4] N.Y.C. Lin, J.H. McCoy, X. Cheng, B. Leahy, J.N. Israelachvili, I. Cohen, A multi-axis confocal 411 rheoscope for studying shear flow of structured fluids, Rev. Sci. Instrum., 85 (2014) 033905.
- 412 [5] G.G. Fuller, Optical rheometry of complex fluid interfaces, Current Opinion in Colloid & Interface413 Science, 2 (1997) 153-157.
- 414 [6] N.J. Wagner, Rheo-optics, Current Opinion in Colloid & Interface Science, 3 (1998) 391-400.
- [7] E. van der Linden, L. Sagis, P. Venema, Rheo-optics and food systems, Current Opinion in Colloid &
 Interface Science, 8 (2003) 349-358.
- [8] R. Besseling, L. Isa, E.R. Weeks, W.C.K. Poon, Quantitative imaging of colloidal flows, Adv. Colloid
 Interface Sci., 146 (2009) 1-17.
- 419 [9] D. Didi, W. Hans, B. Alfons van, I. Arnout, Confocal microscopy of colloidal dispersions in shear
- 420 flow using a counter-rotating cone-plate shear cell, J. Phys.: Condens. Matter, 16 (2004) S3917-421 S3927.
- 422 [10] J.-B. Boitte, C. Vizcaïno, L. Benyahia, J.-M. Herry, C. Michon, M. Hayert, A novel rheo-optical
 423 device for studying complex fluids in a double shear plate geometry, Rev. Sci. Instrum., 84 (2013)
 424 013709.
- 425 [11] L. Hamberg, P. Walkenström, M. Stading, A.-M. Hermansson, Aggregation, viscosity
 426 measurements and direct observation of protein coated latex particles under shear, Food
 427 Hydrocolloids, 15 (2001) 139-151.
- 428 [12] F. Mighri, M.A. Huneault, Dispersion visualization of model fluids in a transparent Couette flow 429 cell, J. Rheol., 45 (2001) 783-797.
- 430 [13] Y. Deyrail, P. Cassagnau, Phase deformation under shear in an immiscible polymer blend:
 431 Influence of strong permanent elastic properties, J. Rheol., 48 (2004) 505-524.
- 432 [14] V. Grenard, N. Taberlet, S. Manneville, Shear-induced structuration of confined carbon black
 433 gels: steady-state features of vorticity-aligned flocs, Soft Matter, 7 (2011) 3920-3928.
- 434 [15] B. Noetinger, L. Petit, E. Guazzelli, M. Clément, Réalisation d'une cellule à cisaillement elliptique,
 435 Rev. Phys. Appl. (Paris), 22 (1987) 1025-1032.
- 436 [16] T. Kume, K. Asakawa, E. Moses, K. Matsuzaka, T. Hashimoto, A new apparatus for simultaneous
- 437 observation of optical microscopy and small-angle light scattering measurements of polymers under438 shear flow, Acta Polym., 46 (1995) 79-85.
- 439 [17] Y.L. Wu, J.H.J. Brand, J.L.A.v. Gemert, J. Verkerk, H. Wisman, A.v. Blaaderen, A. Imhof, A new
- parallel plate shear cell for in situ real-space measurements of complex fluids under shear flow, Rev.
 Sci. Instrum., 78 (2007) 103902.
- [18] E. Tamborini, L. Cipelletti, L. Ramos, Plasticity of a Colloidal Polycrystal under Cyclic Shear, Phys.
 Rev. Lett., 113 (2014) 078301.
- [19] A. Basu, Q. Wen, X. Mao, T.C. Lubensky, P.A. Janmey, A.G. Yodh, Nonaffine Displacements in
 Flexible Polymer Networks, Macromolecules, 44 (2011) 1671-1679.
- [20] R. Besseling, E.R. Weeks, A.B. Schofield, W.C.K. Poon, Three-Dimensional Imaging of Colloidal
 Glasses under Steady Shear, Phys. Rev. Lett., 99 (2007) 028301.
- 448 [21] J. Zausch, J. Horbach, M. Laurati, S.U. Egelhaaf, J.M. Brader, V. Th, M. Fuchs, From equilibrium to
- steady state: the transient dynamics of colloidal liquids under shear, J. Phys.: Condens. Matter, 20(2008) 404210.
- 451 [22] E.D. Knowlton, D.J. Pine, L. Cipelletti, A microscopic view of the yielding transition in 452 concentrated emulsions, Soft Matter, 10 (2014) 6931-6940.

- 453 [23] G. Petekidis, A. Moussaïd, P.N. Pusey, Rearrangements in hard-sphere glasses under oscillatory
 454 shear strain, Physical Review E, 66 (2002) 051402.
- [24] D. Chen, D. Semwogerere, J. Sato, V. Breedveld, E.R. Weeks, Microscopic structural relaxation in
 a sheared supercooled colloidal liquid, Physical Review E, 81 (2010) 011403.
- 457 [25] T. Sentjabrskaja, P. Chaudhuri, M. Hermes, W.C.K. Poon, J. Horbach, S.U. Egelhaaf, M. Laurati,
- 458 Creep and flow of glasses: strain response linked to the spatial distribution of dynamical 459 heterogeneities, Scientific Reports, 5 (2015) 11884.
- 460 [26] R. Besseling, L. Isa, P. Ballesta, G. Petekidis, M.E. Cates, W.C.K. Poon, Shear Banding and Flow-461 Concentration Coupling in Colloidal Glasses, Phys. Rev. Lett., 105 (2010) 268301.
- 462 [27] J.B. Boitte, M. Hayert, C. Michon, Observation of wheat flour doughs under mechanical 463 treatment using confocal microscopy and classification of their microstructures, Journal of Cereal 464 Science, 58 (2013) 365-371.
- [28] H.K. Chan, A. Mohraz, A simple shear cell for the direct visualization of step-stress deformationin soft materials, Rheol. Acta, 52 (2013) 383-394.
- 467 [29] S.K. Dutta, A. Mbi, R.C. Arevalo, D.L. Blair, Development of a confocal rheometer for soft and 468 biological materials, Rev. Sci. Instrum., 84 (2013) 063702.
- 469 [30] A.P. Kotula, M.W. Meyer, F. De Vito, J. Plog, A.R.H. Walker, K.B. Migler, The rheo-Raman
- 470 microscope: Simultaneous chemical, conformational, mechanical, and microstructural measures of471 soft materials, Rev. Sci. Instrum., 87 (2016) 105105.
- 472 [31] S. Wannaborworn, M.R. Mackley, Y. Renardy, Experimental observation and matching numerical
- simulation for the deformation and breakup of immiscible drops in oscillatory shear, J. Rheol., 46(2002) 1279-1293.
- [32] K. Tanaka, K. Yonetake, T. Masuko, R. Akiyama, Shearing microscopy using polarized optical
 microscope with shear stage and spectral analyser to study liquid crystalline polymers, Journal of
 Microscopy, 205 (2002) 15-20.
- 478 [33] M.R. Mackley, S. Wannaborworn, P. Gao, F. Zhao, The optical microscopy of sheared liquids 479 using a newly developed optical stage, Microscopy and Analysis, 69 (1999) 25–27.
- 480 [34] https://www.olympus-ims.com/en/microscope/bxfm.
- 481 [35]
- 482 <u>https://www.marzhauser.com/nc/en/pim/productdetail/pimview/single/pimid/p60.html?filter=p50</u>
- 483 <u>&cHash=083883ba4af55655852019fc9ee88051</u>.
- 484 [36] <u>http://www.nanomotion.com/motion-product/fb075-linear-stage</u>.
- 485 [37] <u>http://www.galilmc.com/motion-controllers/multi-axis/dmc-40x0</u>.
- 486 [38] <u>https://www.onsemi.com/pub/Collateral/NOIP1SN5000A-D.PDF</u>.