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## A CHLORHEXIDINE-RELEASING EPOXY-BASED COATING ON TITANIUM IMPLANTS PREVENTS *STAPHYLOCOCCUS AUREUS* EXPERIMENTAL BIOMATERIAL-ASSOCIATED INFECTION

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### Abstract

Prevention of biomaterial-associated infections (BAI) remains a challenging problem, in particular due to the increased risk of resistance development with the current antibiotic-based strategies. Metallic orthopaedic devices, such as non-cemented implants, are often inserted under high mechanical stress. These non-cemented implants cannot be protected by *e.g.* antibiotic-releasing bone cement or other antimicrobial approaches, such as the use of bioactive glass. Therefore, in order to avoid abrasion during implantation procedures, we developed an antimicrobial coating with great mechanical stability for orthopaedic implants, to prevent *Staphylococcus aureus* BAI. We incorporated 5 and 10 wt % chlorhexidine in a novel mechanically stable epoxy-based coating, designated CHX<sub>5</sub> and CHX<sub>10</sub>, respectively. The coatings displayed potent bactericidal activity *in vitro* against *S. aureus*, with over 80 % of the release (19 µg/cm<sup>2</sup> for CHX<sub>5</sub> and 41 µg/cm<sup>2</sup> for CHX<sub>10</sub>) occurring within the first 24 h. In mice, the CHX<sub>10</sub> coating significantly reduced the number of CFU (colony forming units), both on the implants and in the peri-implant tissues, 1 d after *S. aureus* challenge. The CHX<sub>10</sub>-coated implants were well-tolerated by the animals, with no signs of toxicity observed by histological analysis. Moreover, the coating significantly reduced the frequency of culture-positive tissues 1 d, and of culture-positive implants 1 and 4 d after challenge. In summary, the chlorhexidine-releasing mechanically stable epoxy-based CHX<sub>10</sub> coating prevented implant colonisation and *S. aureus* BAI in mice and has good prospects for clinical development.

**Keywords:** antimicrobial coating, chlorhexidine, *Staphylococcus aureus*, titanium, *in vivo* test, mouse, infection, histology.

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### Introduction

Medical devices manufactured from titanium, such as prosthetic joints and dental implants, are widely used in modern medicine. Titanium's lightness, good mechanochemical and osseo-integrative properties make it an ideal material for bone implant application (Web ref. 1). Unfortunately, as with any inserted or implanted foreign body material (Elek and Conen, 1957; Zimmerli and Sendi, 2011), a significant problem with titanium implants is the risk of infection. These so-called biomaterial-associated infections (BAIs) are feared complications in orthopaedic and trauma surgery with tremendous consequences for the patients. The majority of BAIs are caused by staphylococci, especially *Staphylococcus epidermidis* and *Staphylococcus aureus* (*S. aureus*) (Anderson and Marchant, 2000; O'Gara and Humphreys, 2001). Formation of a biofilm consisting of bacteria, bacterial products and host proteins on the biomaterial surface is considered the major component in the pathogenesis of BAI (Costerton *et al.*, 1999). In addition, the presence of the foreign body material leads to colonisation of tissue surrounding implants (Broekhuizen *et al.*, 2008a; Engelsman *et al.*, 2010; Nejadnik *et al.*, 2008; Vuong *et al.*, 2004; Zaat *et al.*, 2010).

In order to prevent infection of medical devices, a variety of surface modifications have been developed. These include non-adhesive, contact-killing and tissue-integrating surfaces, as well as coatings for the controlled release of antimicrobial agents (Brooks *et al.*, 2013; Busscher *et al.*, 2012). To prevent both, colonisation of the implant itself and of the tissue surrounding the implant, an antimicrobial-releasing coating is preferred. However, the

use of antibiotics on a medical device is discouraged by government regulatory agencies, especially the American Food and Drug Administration (FDA), in view of the increasing development of resistance (Brooks *et al.*, 2013; FDA, 2007). As an alternative to antibiotics, commonly used antiseptics, such as chlorhexidine, might be used. Chlorhexidine (CHX) has broad antimicrobial activity and has been shown to prevent infection in patients and in animal models. It is used topically, for site preparation in surgical procedures, and also intracorporally, *e.g.* in abdominal surgery for rapid sterilisation of exposed tissues prior to wound closure (Wilkins and Unverdorben, 2013). Moreover, chlorhexidine is widely applied as an irrigant fluid in dentistry with low toxicity and good tolerability in soft tissue (Iqbal, 2012). In biomaterials, chlorhexidine has been applied in combination with silver-sulfadiazine in a coating on intravenous catheters, which decreases catheter-related infection in humans (Maki, 1997). Experimental titanium tibia intramedullary nails coated with chlorhexidine and chloroxylenol have reduced rates of orthopaedic device-related infection in rabbits (Darouiche *et al.*, 1998a; Darouiche *et al.*, 1998b).

In the present study we aimed to design an antimicrobial coating for titanium implants for general application in orthopaedics and trauma, with high mechanical stability, in order to avoid abrasion during implantation procedures. Coatings and materials manufactured from epoxy resins are among the few available that meet this requirement (Berruet *et al.*, 1987; Garcia *et al.*, 2009). Epoxy resins can be catalytically cross-linked or they can be cured using reagents such as polyamines. The typically good adhesion to metallic substrates and good mechanical properties of epoxy-based coatings (Foyet *et al.*, 2009; Meis *et al.*, 2014; Wu *et al.*, 2014) prompted us to investigate an epoxy-amine coating network containing chlorhexidine as the antimicrobial agent for orthopaedic implants. Dopamine is added to the coating because the catechol moiety of dopamine is known for its strong adhesion to a range of metallic substrates, including medical-grade titanium (Fan *et al.*, 2006; Shi *et al.*, 2008; Watson *et al.*, 2010).

Therefore, the purpose of the present study was to develop an epoxy-based chlorhexidine-releasing coating for titanium implants with potent *in vitro* as well as *in vivo* antibacterial activity in a murine subcutaneous implant infection model. In this model, the release of chlorhexidine proved to be effective in preventing *S. aureus* colonisation of the implants and surrounding tissue without signs of toxicity.

## Materials and Methods

### Coating preparation

Coating formulations as described in Table 1 were prepared as follows. For solution A, isophorone diamine (IPD; Sigma-Aldrich  $\geq 99\%$ ) was dissolved in ethanol (EtOH) at room temperature, while stirring. Dopamine hydrochloride (dopamine; Sigma-Aldrich) was added to this solution and left stirring until completely dissolved. Next, glycerol diglycidyl ether (epoxy resin; Sigma-Aldrich, technical grade) was added to the mixture. The resulting solution A

was left to stand for 1 h at room temperature. For solution B, chlorhexidine diacetate salt hydrate [Sigma-Aldrich, bis(biguanide) antimicrobial] was dissolved in EtOH at room temperature, while stirring. For the unloaded control coatings, no chlorhexidine was added to solution B. Just prior to the coating procedures, solutions A and B were mixed. The weights of the solids used for the solutions A and B are listed in Table 1. The coating formulations were designated CHX<sub>0</sub> (no chlorhexidine), CHX<sub>5</sub> (5 wt % chlorhexidine) and CHX<sub>10</sub> (10 wt % chlorhexidine). Two batches of the CHX<sub>0</sub> and CHX<sub>10</sub> coating formulations were prepared. The chemical composition was kept maximally constant in both batches with particular attention to the ratio between the amino groups and the epoxy groups determining the crosslink density. The coating formulations of CHX<sub>0</sub> (1st batch), CHX<sub>5</sub> and CHX<sub>10</sub> (1st batch) were used to coat aluminium sheets and titanium disks for the *in vitro* assays. The second batch CHX<sub>0</sub> and CHX<sub>10</sub> formulations were used to coat the titanium implants for the *in vivo* assays. Formulations were prepared fresh prior to application to the different materials.

### Application of coating to aluminium sheets and titanium discs by spin-coating

For the *in vitro* experiments, coatings were applied on aluminium sheets (5 × 5 cm), Q-panels (15.3 × 10.1 cm) and medical grade titanium [Ti 6Al-4V ELI, surface finish 2B (ASTM A480) (Web ref. 2)] discs (diameter 20 mm, height 1 mm), which had been cleaned by submerging in ethanol and acetone and subsequent treatment with ultrasound for 5 min. Aliquots of approximately 1 mL and 0.1-0.2 mL of the coating formulation were applied to the surface of aluminium sheets and titanium disks, respectively, and the samples were spin-coated at a rotation speed of 1,000 rpm for 30 s. The samples were air-dried for 0.5-2 h and subsequently placed in an oven at ~90 °C for 3 h.

### Application of coatings to titanium implants by dipping

For the *in vivo* experiments, medical-grade titanium implants (10 × 4 × 1 mm, with a slit to allow cutting of the implant after explantation, see below) were manufactured by Flowcut (Nederweert, The Netherlands) and coated using the following procedure. First, the implants were cleaned by boiling them for 1 h in 10 wt % aqueous NaOH, followed by ultrasonic treatment in ethanol and acetone for 5 min. The implants were dip-coated to allow complete coverage of the surface in spite of their complex shape. The implants were connected to a paperclip and submerged completely in the dipping solution using a dip coater (Specialty Coating Systems, type PL-3201, SCS, Indianapolis, IN, USA) with a dip speed of 2 mm/s. The dipped implants were dried for 0.5-1 h and placed in a convection oven at 90 °C for 3 h for curing. The paperclip was removed and the coated implants sterilised by autoclaving at 121 °C for 20 min.

### *In vitro* chlorhexidine release assay

Samples of 1 × 5 cm were cut from the coated aluminium sheets and incubated with 4 mL of phosphate buffered

**Table 1.** Coating formulation compositions. Batch 1 was used for spin-coating of the aluminium sheets, Q-panels and titanium disks; batch 2 for dip-coating of the *in vivo* titanium implants. To obtain a similar coating thickness in both coating procedures, the viscosity of the formulation had to be smaller in case of the dip-coating procedure and therefore a reduced solid in EtOH content was used in batch 2.

		EtOH (g)		Solids (g)								
Coating	Batch	Sol. A	Sol. B	Epoxy		IPD		Dopamine		CHX		NH/epoxy
CHX <sub>0</sub>	1	5.00	10.00	4.993	(64.9 %)	1.731	(22.5 %)	0.969	(12.6 %)	-		1.04
CHX <sub>5</sub>	1	5.00	10.00	4.992	(61.8 %)	1.720	(21.3 %)	0.969	(12.0 %)	0.369	(4.9 %)	1.03
CHX <sub>10</sub>	1	5.00	10.00	5.021	(59.0 %)	1.719	(20.2 %)	0.962	(11.3 %)	0.809	(9.5 %)	1.03
CHX <sub>0</sub>	2	10.00	15.00	5.012	(65.0 %)	1.740	(22.6 %)	0.962	(12.5 %)	-		1.07
CHX <sub>10</sub>	2	10.00	15.00	5.012	(58.5 %)	1.740	(20.3 %)	0.962	(11.2 %)	0.858	(10.0 %)	1.07

saline (PBS; pH = 7.0) at 37 °C, while shaking at 100 rpm. At different time points (0, 1, 2, 3 and 4 d) the medium was exchanged with fresh PBS and the chlorhexidine concentration in the removed medium was determined by ultraviolet-visible spectroscopy (UV-Vis). The absorption values at  $\lambda = 255$  nm were converted to mass concentrations of chlorhexidine through a calibration curve ( $m = 22.427 \times A + 0.2474$ ; with m: mass of chlorhexidine, A: measured absorbance at  $\lambda = 255$  nm) and these values were plotted against time in a cumulative way. The lower limit of detection for the absorption was 0.005 AU (arbitrary units). In case of an absorption above 1.5 AU, the sample was diluted and measured again. The amount of chlorhexidine released at 96 h was defined as the total releasable amount. Percentage release at other time points was calculated based on this amount. The results of one representative experiment are shown.

### Mechanical stability assays

#### Solvent resistant assay

A double rub test was used to assess the mechanical stability of the coatings under dry conditions. A number of the coated aluminium Q-panels were washed in PBS at 37 °C for 48 h, to assess the mechanical stability of the coatings under wet conditions. The double rub test was performed essentially as described in the solvent resistance standard test method [ASTM D5402 (Web ref. 3)], with some modifications. In short, a piece of felt (diameter 2 cm), soaked in EtOH (more appropriate solvent than methyl ethyl ketone (MEK) used in ASTM D5402), was moved back and forth (1 double rub) over a distance of 12 cm (in 1 s) using an automated rubbing apparatus (DSM, in house equipment) on a defined area of the coated substrate with a constant load of 400 g. When macroscopic damage to the coating was observed, the test was stopped and the number of rubs was noted. A maximum of 200 double-rubs were applied and the resistance was rated by eye on a 0 to 5 scale, with 0 meaning penetration of the substrate in 200 double rubs or less and 5 meaning no effect on surface after 200 double rubs.

#### Tape adhesion test

The adhesion of the coating to the substrate was tested essentially as described in the tape standard test method

[ASTM D3359 (Web ref. 4)]. Due to the coating hardness, it was not possible to make cuts as described in ASTM D3359 prior to the procedure. In short, 25 mm wide semi-transparent pressure-sensitive tape was pressed firmly onto a defined area of the coating and pulled off rapidly under an angle close to 180°. The test was repeated on two other sites of the coated aluminium Q-panels and the level of adhesion was rated by eye on 0- 5 scale, with 0 meaning full removal of the coating and 5 meaning no peeling or removal of the coating.

### Scanning electron microscopy

A JEOL JSM-IT100 scanning electron microscope (SEM; JEOL, Tokyo, Japan) was used to examine non-coated and CHX<sub>10</sub>-coated titanium implants. Before imaging, specimens were sputter-coated with a carbon layer of ~6 nm thickness. The images were collected at an acceleration voltage of 5-15 kV.

### Atomic force microscopy

Atomic force microscopy (AFM) was performed with an AFM Dimension Edge (Bruker, Camarillo, CA, USA). Non-coated and CHX<sub>10</sub>-coated titanium implants were scanned in tapping mode with a standard silicon cantilever at a resonance frequency of 370 kHz and a scan rate of 0.5 Hz using a tip with a radius of 10 nm and a height of 10-15  $\mu\text{m}$ . The average surface roughness ( $R_a$ ) was quantified on three different locations of each sample using images of  $100 \times 100 \mu\text{m}^2$  and presented as average with standard deviation.

### Bacteria

*S. aureus* ATCC 49230 is a biofilm-producing strain (Li *et al.*, 2008) originally isolated from a patient with osteomyelitis (Smeltzer *et al.*, 1997). In the *in vivo* studies, in order to verify the identity of the challenge strain after re-isolation, we constructed a green fluorescent *S. aureus* ATCC 49230 strain by introducing plasmid WVV189, essentially as described earlier (Riool *et al.*, 2014). This plasmid carries the  $gfp_{uvr}$  gene under control of a constitutive *S. aureus* promoter and a chloramphenicol resistance gene for selection (Rooijackers *et al.*, 2005). The strain, *S. aureus* ATCC 49230 GFP, was designated *S. aureus* GFP in this paper, for brevity. During growth, the plasmid was



stably maintained by the *S. aureus* GFP bacteria, even in absence of chloramphenicol. Chloramphenicol (10 µg/mL) was, however, always added to cultures of the strain used to prepare inocula, in order to avoid any potential loss of the plasmid. The susceptibility to chlorhexidine and the growth rate in tryptic soy broth (TSB; BD Difco) of *S. aureus* GFP and of the wild type *S. aureus* ATCC 49230 were equal (data not shown). In previous studies we showed that the presence of plasmid WW189 did not influence the virulence of *Staphylococcus epidermidis* in mice (Riool *et al.*, 2014).

#### Surface bactericidal activity assay

A modified version (Necula *et al.*, 2009) of the Japanese Industrial Standard test for surface microbicidal activity [JIS Z 2801:2000 (Japanese Standards Association, 2000)] was used to evaluate the bactericidal activity of quadruplicate coated titanium disks against *S. aureus*. In short, *S. aureus* was cultured shaking at 37 °C in TSB with 10 µg/mL chloramphenicol, up to the logarithmic growth phase. Subsequently, the bacteria were diluted with TSB to 10<sup>7</sup> colony forming units (CFU)/mL, based on the optical density at 620 nm (OD<sub>620</sub>). UV-sterilised nitrocellulose filter disks (diameter 13 mm) were placed on a Colombia sheep blood agar plate and 20 µL of the diluted bacterial culture, containing 2 × 10<sup>5</sup> CFU, were pipetted onto the filters. The medium was absorbed by the agar, while the bacteria were retained on the filter. 20 µL of 1 % TSB in 10 mM phosphate buffer were pipetted centrally on the surface of each titanium disk and an inoculated filter disk was carefully placed on top, with the bacteria contacting the coated surface. All disks with bacterial filters were placed individually in Petri dishes and incubated at 37 °C for 24 h in a humid atmosphere. After incubation, each disk and the corresponding filter were placed together in 5 mL of TSB, sonicated for 30 s in a sonicator water bath (Elma Transsonic T460, 35 kHz; Elma Schmidbauer GmbH, Singen, Germany) and vortexed for 1 min to dislodge adherent bacteria. This procedure does not affect bacterial viability (Boelens *et al.*, 2000a). Per sample, two independent 10-fold serial dilutions were made in a microtiter plate and duplicate 10 µL aliquots of the undiluted suspension and of the dilutions were pipetted onto blood agar plates. The blood agar plates were incubated overnight at 37 °C and number of colonies was counted the following day.

#### Mouse experimental biomaterial-associated infection model

An established subcutaneous biomaterial-associated infection model was used to assess prevention of infection (Boelens *et al.*, 2000c; Broekhuizen *et al.*, 2009; Christensen *et al.*, 1983; Riool *et al.*, 2014). In this model, planktonic bacteria are injected along the implant, as opposed to biofilm treatment models, where bacterial biofilm are pre-grown on the implant surface prior to implantation (Riool *et al.*, 2014). The mouse study was approved by the Animal Ethical Committee of the Academic Medical Center at the University of Amsterdam, The Netherlands. Specific pathogen-free C57BL/6J OlaHsd immune competent female mice (Envigo, Horst, The

Netherlands), 7 to 9 weeks old and weighing 17 to 20 g, were used.

Mice were anaesthetised with 2 % isoflurane (Pharmachemie) in oxygen in a laminar flow cabinet, followed by a subcutaneous injection of buprenorphine (0.05 mg/kg; Temgesic, RB Pharmaceuticals Limited) for pain control, 15 min prior to the surgical procedure. The backs of the mice were shaved and disinfected with 70 % ethanol. On each side, an incision of 0.5 cm was made, 1 cm lateral to the spine. Subsequently, the (coated) titanium biomaterials were implanted subcutaneously with minimal tissue damage using a transponder, specifically designed for these implants (Riool *et al.*, 2014). The incisions were closed with a single 0/6-vicryl stitch (Vicryl, Ethicon).

*S. aureus* GFP was cultured in TSB with 10 µg/mL chloramphenicol to the mid-logarithmic growth phase. The bacteria were washed in 0.9 % NaCl (saline) and resuspended in saline, based on the OD<sub>620</sub>, to 4 × 10<sup>6</sup> [inoculum size was based on previous experience with *S. aureus* in the mouse BAI model (de Breij *et al.*, 2016)] or 4 × 10<sup>7</sup> CFU/mL. Immediately following implantation, 25 µL of either of the *S. aureus* GFP inoculum suspensions, containing 10<sup>5</sup> or 10<sup>6</sup> CFU, respectively, were injected along the (coated) implants using a repetitive injector (Stepper model 4001-025; Dymax Corporation, Torrington, CT, USA). Groups of 9 mice with 2 implants each were used in the experiments. Mice were housed singly in individually ventilated cages (IVCs) and were provided with sterile food and water *ad libitum*.

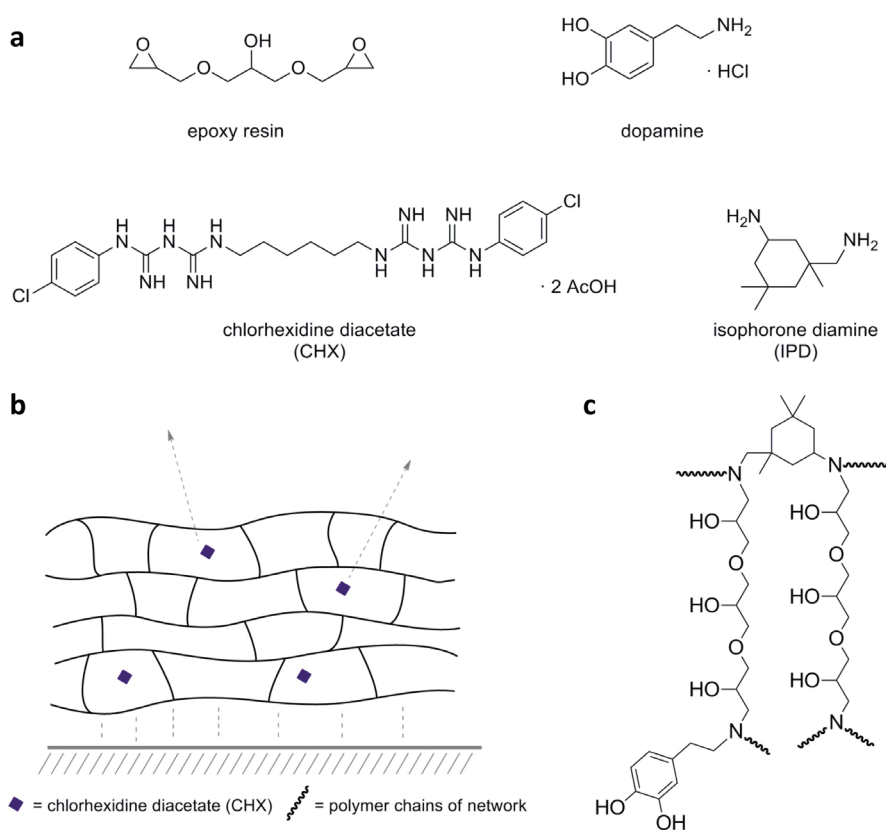
1 or 4 d after implantation, mice were anaesthetised with 2 % isoflurane in oxygen and buprenorphine was administered for pain control 15 min before biopsies were taken. Standardised biopsies (diameter 12 mm) were taken from the implantation sites as described previously (Boelens *et al.*, 2000a; Riool *et al.*, 2014) and subsequently mice were euthanised by cervical dislocation. Each biopsy included skin, subcutaneous tissue and the implant. The implant was separated from the tissue and both implant and tissue were used for quantitative culture of bacteria. The implants were vortexed briefly in 0.5 mL of PBS to remove non-adherent bacteria and then sonicated in saline for 5 min in a water bath sonicator to dislodge adherent bacteria. The tissue samples were homogenised in 0.5 mL of PBS using 5 zirconia beads (2 mm diameter, BioSpec Products, Bartlesville, OK, USA) and the MagnaLysor System (Roche), with 3 cycles of 30 s at 7000 rpm, with 30 s cooling on ice between cycles. Like the sonication procedure of the implants, this homogenisation procedure does not affect the viability of staphylococci (Boelens *et al.*, 2000a). The sonicates and homogenates were 10-fold serially diluted and plated on blood agar plates. In addition, the sonicated implants and 50 µL of each homogenate were cultured in 5 mL TSB containing 0.5 % (v/v) Tween-80, shaking at 37 °C for 48 h. The cultured *S. aureus* GFP bacteria were distinguished from mouse flora by inspection under exposure to UV light in a G:box (Syngene). The number of cultured bacteria was expressed as log CFU per implant or per homogenised biopsy. The lower limit of detection was 10 CFU. This value was assigned to samples positive in broth and negative in blood agar plate culture. For statistical analysis and to visualise the data

on a logarithmic scale, a value of 1 CFU per sample was assigned when no growth had occurred in either culture system.

### Histology

Tissue biopsies were embedded in plastic [methyl methacrylate/butyl methacrylate (MMA/BMA); Merck Schuchart] and 3  $\mu\text{m}$  sections were cut from these samples. In each experimental group, 4 biopsies were randomly selected for histological examination and one or two microscopic sections per biopsy were immunohistochemically stained. The sections were deplastified by incubating twice in 100 % acetone for 6 min, washed in demineralised water for 5 min and incubated in methanol with 0.3 %  $\text{H}_2\text{O}_2$  for 20 min at RT (room temperature) in the dark, to block endogenous peroxidase activity. The sections were subjected to heat-induced epitope retrieval (HIER) (van der Loos, 2010), performed in a PreTreatment Module (Lab Vision) by incubation in citrate buffer pH 6.0 (TA-250-PM1X, Thermo Fisher Scientific) for 20 min at 98 °C, rinsed with tap water and incubated with Superblock (AAA999, Klinipath) for 10 min at RT. Sections were incubated overnight at 4 °C with rat anti-mouse monoclonal F4/80 (macrophages; MCA497GA, clone: CI:A3-1, Serotec) diluted 1: 1000 in Tris-HCl-buffered saline (TBS; 50 mM

Tris, 0.9 % NaCl), subsequently with rabbit anti-rat IgG (6130-01, fab2; SBA/ITK) diluted 1: 3000 in TBS with 20 % normal mouse serum for 30 min at RT, followed by undiluted BrightVision anti-rabbit AP (DPVB-55AP, Immunologic) for 30 min at RT and finally with Vector Blue (SK-5300, Vector Labs) for 10 min at RT, with three TBS washes between all incubations. After rinsing with tap water, a HIER-citrate antigen retrieval step of 10 min at 98 °C and a Superblock step were performed as described above. Next, sections were incubated with 1: 10000 diluted rabbit anti-*S. aureus* polyclonal IgG (20920, Abcam) for 1 h at RT, with undiluted BrightVision anti-rabbit AP for 30 min at RT and with Vector Red (SK-5100, Vector Labs) for 10 min at RT, again with three TBS washes between all incubations. After another HIER-citrate (10 min at 98 °C) and Superblock incubation, sections were incubated with 1: 1000 diluted rat anti-mouse Ly-6G-FITC (neutrophils; 553126, BD) in TBS for 2 h at RT, with 1: 1000 diluted rabbit anti-FITC (4510-7804, Nuclilab) in the presence of 5 % normal mouse serum in TBS for 15 min at RT, with undiluted BrightVision anti-rabbit IgG-HRP (DPVR-110HRP, Immunologic) for 30 min at RT, and finally stained for ~8 min with DAB (BS04-110, Immunologic) until they turned yellow/brownish. Sections were then washed with tap water, stained with 10-fold diluted haematoxylin (4085-9002, Klinipath) for 2 min, washed



**Fig. 1.** Overview of the coating components (a) and architecture (b,c). (a) The coating components: epoxy resin (*i.e.* the diglycidyl ether of glycerol), dopamine, chlorhexidine diacetate (CHX) and isophorone diamine (IPD). (b) Schematic representation of the coating architecture, including chlorhexidine diacetate (CHX; diamonds) release from the epoxy-based coating on a substrate (////). (c) Representation of a (possible) crosslink within the network.

with tap water and dried on a heat block at 50 °C. A drop of VectaMount (H-5000, Vector Labs) was applied and the sections were covered with a coverslip.

Between 2 and 5 multispectral imaging data sets of the immunohistochemically stained sections were acquired at 420 to 720 nm with intervals of 20 nm, using the Nuance Imaging System (PerkinElmer/Caliper Life Science). The data sets were spectrally unmixed to identify the different cell types (macrophages, neutrophils and bacteria). In the negative controls without primary antibodies no staining was observed.

### Statistical analysis

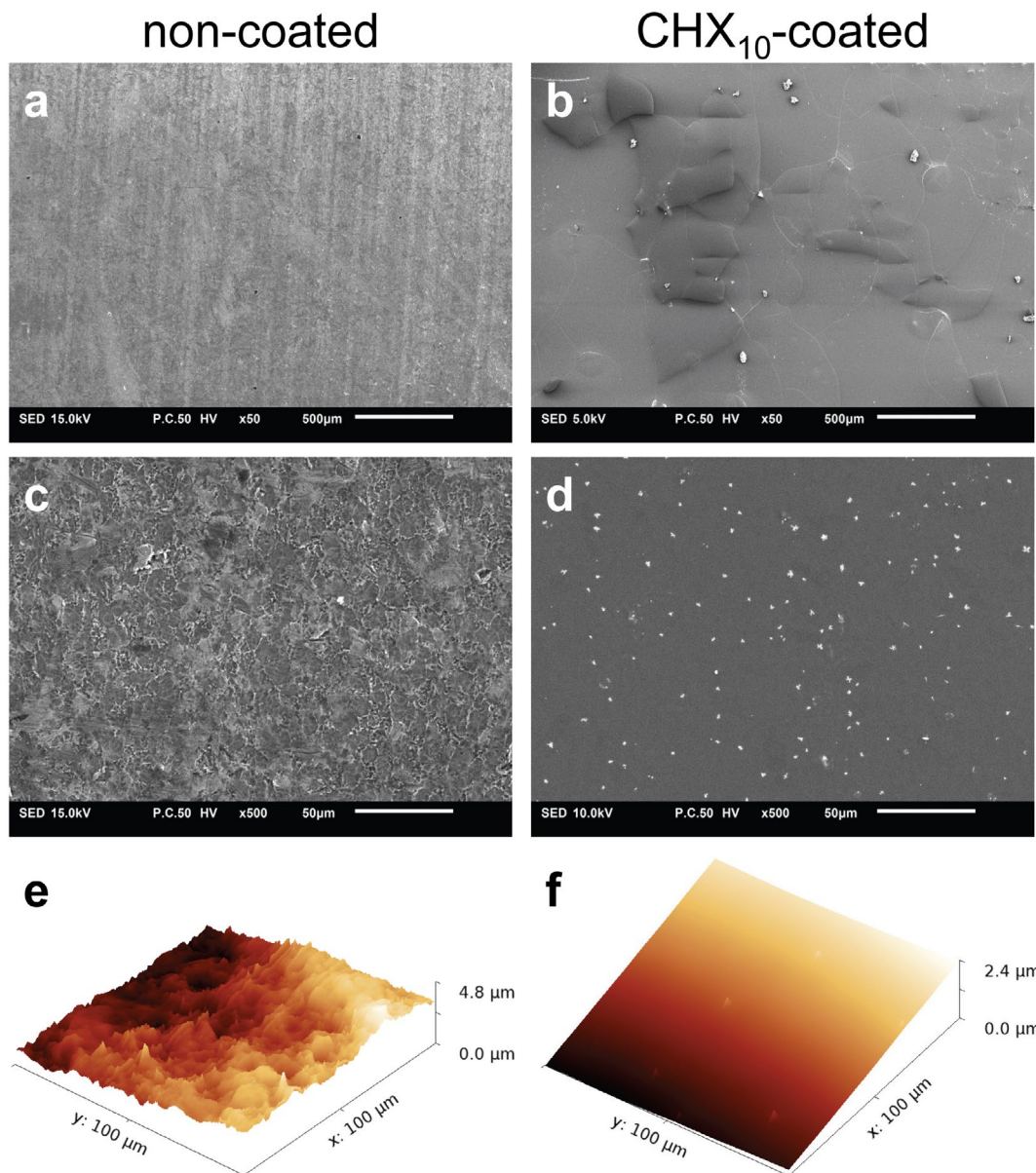
All statistical analyses were performed in Graphpad Prism. For CFU counts, two-sample comparisons were made using a two-tailed Mann-Whitney rank sum test. The significance of differences between the frequencies of categorical

variables was determined using Fisher's exact test. For all tests,  $p \leq 0.05$  was considered significant.

## Results

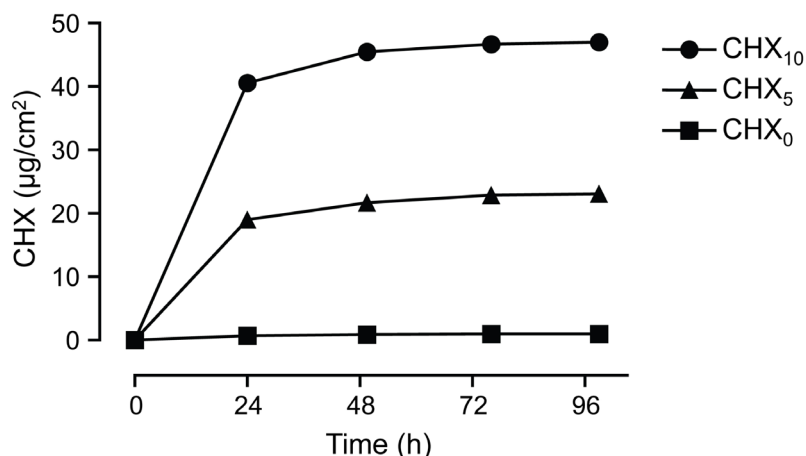
### Coating synthesis and characterisation

A procedure for the preparation of the coating formulation was established utilising the diglycidyl ether of glycerol as the epoxy resin, IPD for curing and dopamine to optimise the adhesion to titanium (Fig. 1a). Ensuring a close to equimolar ratio of NH/epoxy in the coating formulations (Table 1) resulted in a highly cross-linked network (Fig. 1c) when the procedure was applied to aluminium sheets and Q-panels as well as to titanium disks. The coatings showed excellent resistance to rubbing, with no visible damage after 200 double-rubs (classification: 5). Moreover,

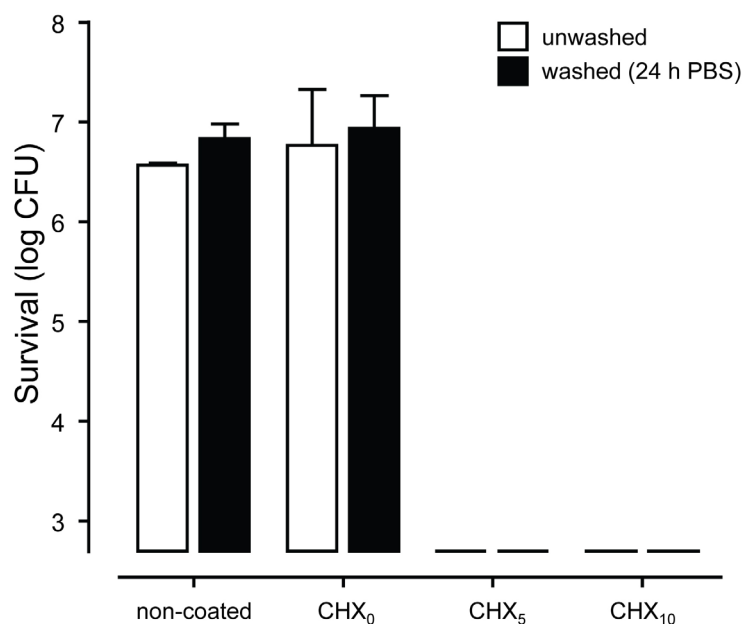


**Fig. 2.** Surface morphology of non-coated (left panels) and CHX<sub>10</sub>-coated titanium implants (right panels) shown by SEM recordings at 50× (a,b) and 500× magnification (c,d) and 3D morphology by AFM analysis (e,f).





**Fig. 3.** Cumulative release of chlorhexidine ( $\mu\text{g}/\text{cm}^2$ ) from aluminium sheets coated with CHX<sub>0</sub> (no CHX; squares), CHX<sub>5</sub> (5 wt % CHX; triangle) or CHX<sub>10</sub> (10 wt % CHX; circles) over time. Results of one representative experiment.



**Fig. 4.** Surface bactericidal activity of chlorhexidine-releasing epoxy-based coating on titanium disks against  $2 \times 10^5$  CFU of *S. aureus*. The survival of the bacteria 24 h after application on unwashed (white bars) and washed (24 h in PBS; black bars) disks is shown. Non-coated, titanium disks without coating; CHX<sub>0</sub>, titanium disks with coating not containing chlorhexidine; CHX<sub>5</sub> and CHX<sub>10</sub>, titanium disks with coating containing 5 or 10 wt % chlorhexidine, respectively. The detection limit was log 2.7 CFU of *S. aureus*.

no visual damage was observed upon tape stripping (classification: 5), proving a good adhesion of the coating to the substrate. The mechanical properties of the coatings did not change when the coated substrates had been incubated in PBS at 37 °C for 48 h prior to the mechanical testing. Next, we prepared the same coating formulations with chlorhexidine diacetate added at approximately 5 and 10 wt % of the total solid content or without this agent (Fig. 1b). Separate batches of coating formulations were prepared for the *in vitro* studies (CHX<sub>0</sub>, CHX<sub>5</sub> and CHX<sub>10</sub>) and for the *in vivo* studies (CHX<sub>0</sub> and CHX<sub>10</sub>) (Table 1). No differences were observed between the mechanical properties of the separate batches of identical formulations.

The SEM analyses of the non-coated surfaces showed a rough topography with visible grooves (Fig. 2a) and enhanced local roughness (Fig. 2c), which was also affirmed by the 3D morphology as assessed by AFM, with an estimated average surface roughness ( $R_a$ ) of  $861.9 \pm 27$  nm (Fig. 2e). In contrast, the CHX<sub>10</sub>-coated samples showed a smooth (Fig. 2f;  $R_a$  of  $530.5 \pm 15$  nm), compact and non-porous layer completely covering the original titanium topography (Fig. 2b,d). In addition, white

structures protruding from the surface were visible on the coated samples, both in SEM (Fig. 2b,d) and AFM (Fig. 2f). The nature of these protrusions remains elusive.

#### *In vitro* chlorhexidine release

The release curves showed that the total amount of released chlorhexidine could be adjusted by changing the concentration of chlorhexidine in the formulation, *i.e.* 5 versus 10 wt % chlorhexidine (CHX<sub>5</sub> and CHX<sub>10</sub> in Fig. 3). Chlorhexidine was released from the CHX<sub>5</sub>- and CHX<sub>10</sub>-coated aluminium sheets in a burst-release fashion with > 80 % of the release, 19 and 41  $\mu\text{g}/\text{cm}^2$  chlorhexidine, respectively, occurring in the first 24 h. After 4 d, 23 and 47  $\mu\text{g}/\text{cm}^2$  chlorhexidine were released from the CHX<sub>5</sub>- and CHX<sub>10</sub>-coated sheets, respectively.

#### *In vitro* surface bactericidal activity

The CHX<sub>5</sub> and CHX<sub>10</sub> coatings applied on titanium disks completely killed the *S. aureus* inoculum after 24 h of incubation (Fig. 4). After washing with PBS for 24 h, both the CHX<sub>5</sub> and CHX<sub>10</sub> coatings remained fully bactericidal. The coating without chlorhexidine (CHX<sub>0</sub>) did not reduce



survival of *S. aureus*, showing that the bactericidal activity of the CHX<sub>5</sub> and CHX<sub>10</sub> coatings was due to the release of chlorhexidine.

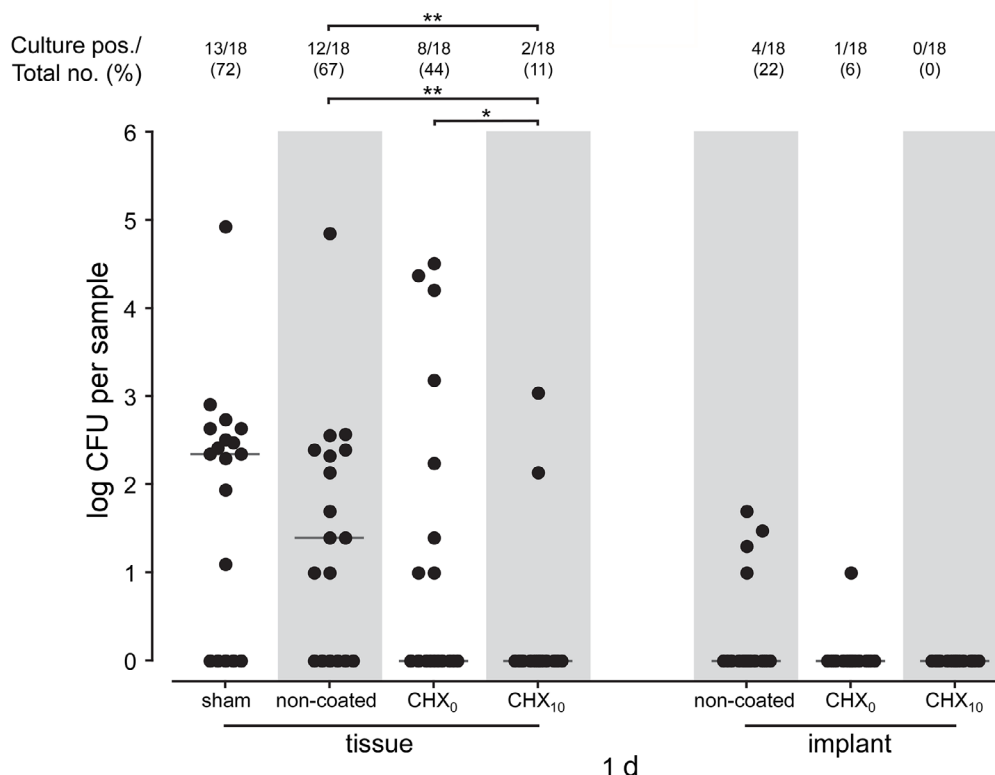
### ***In vivo* antibacterial activity of CHX<sub>10</sub>-coated titanium implants in mouse experimental BAI**

In a first experiment, mice were either sham-operated or they received non-coated, CHX<sub>0</sub>- or CHX<sub>10</sub>-coated titanium subcutaneous implants. An *S. aureus* GFP inoculum of 10<sup>5</sup> CFU was subsequently injected at the site of sham surgery in the sham-operated mice or along the implants in the other mice. A potentially protective effect of the CHX<sub>10</sub> coating against colonisation of the implants could not be assessed, since the level of implant colonisation in the animals receiving non-coated or CHX<sub>0</sub>-coated implants appeared to be unexpectedly low (Fig. 5). The level of tissue colonisation did however allow evaluation. 1 d after challenge, tissues from sham-operated mice and from mice receiving non-coated implants did not differ in frequency of culture positive tissues nor in number of CFU cultured from these tissues. In the group of mice with CHX<sub>10</sub>-coated implants, the frequency of culture-positive tissues was lower than in the mice with non-coated implants ( $p = 0.0016$ ) and the number of CFU retrieved was lower than from tissues of mice with CHX<sub>0</sub>-coated ( $p = 0.029$ ) or non-coated implants ( $p = 0.0022$ ) (Fig. 5).

No apparent macroscopic signs of toxicity or inflammation were observed in any of the mice.

In order to allow evaluation of an effect of the CHX<sub>10</sub> coating on implant colonisation, we repeated the experiment with an *S. aureus* GFP inoculum of 10<sup>6</sup> CFU with expected higher levels of *in vivo* colonisation of non-coated implants. We used the coating with the greatest chlorhexidine content (CHX<sub>10</sub>) and non-coated implants for comparison. With this increased inoculum size, both the implant surfaces and the peri-implant tissues were strongly colonised in the group of mice with non-coated implants (Fig. 6). In this group, 1 d after challenge, 100 % of the tissues and 89 % of the implants were culture positive. Similarly, at day 4, 83 % of the tissues and 28 % of the implants were culture positive and number of CFU retrieved from the implants and the surrounding tissue was larger with this increased inoculum size than with an inoculum of 10<sup>5</sup> CFU used in the previous experiment (compare Fig. 6 to Fig. 5).

1 d after challenge, the CHX<sub>10</sub> coating significantly reduced the number of culture-positive implants ( $p < 0.001$ ) as well as the number of CFU in the tissue ( $p = 0.0015$ ) and on the implants ( $p < 0.001$ ). 4 d after challenge, the group with CHX<sub>10</sub>-coated implants had significantly smaller number of culture-positive tissues ( $p = 0.0153$ ) and culture-positive implants ( $p = 0.0455$ ) than mice with non-coated implants. 1 d after challenge, only 4 of the



**Fig. 5.** Bactericidal activity of chlorhexidine-releasing epoxy-based coatings applied on titanium implants against  $1 \times 10^5$  CFU of *S. aureus* GFP in mouse experimental biomaterial-associated infection 1 d after implantation. Sham, surgery but no implant; non-coated, titanium implant without coating; CHX<sub>0</sub>, titanium implant with coating not containing chlorhexidine; CHX<sub>10</sub>, titanium implant with coating containing 10 wt % chlorhexidine. Results are expressed as the frequencies and percentages of culture-positive samples and as the log of the numbers of CFU retrieved. The horizontal line represents the median log CFU. Frequencies of culture-positive samples and log numbers of CFU were analysed by Fisher's exact and Mann-Whitney tests, respectively. \*  $p \leq 0.05$  and \*\*  $p \leq 0.01$ .

CHX<sub>10</sub>-coated implants were culture positive, yielding a small number of CFUs and the surfaces of all CHX<sub>10</sub>-coated implants were culture negative 4 d after challenge. No apparent macroscopic signs of toxicity or inflammation were observed in the peri-implant tissue of any of the mice. Thus, the release of chlorhexidine proved to be effective to prevent *S. aureus* colonisation of the implant and the surrounding tissue *in vivo* with no visible adverse effects.

### Histology

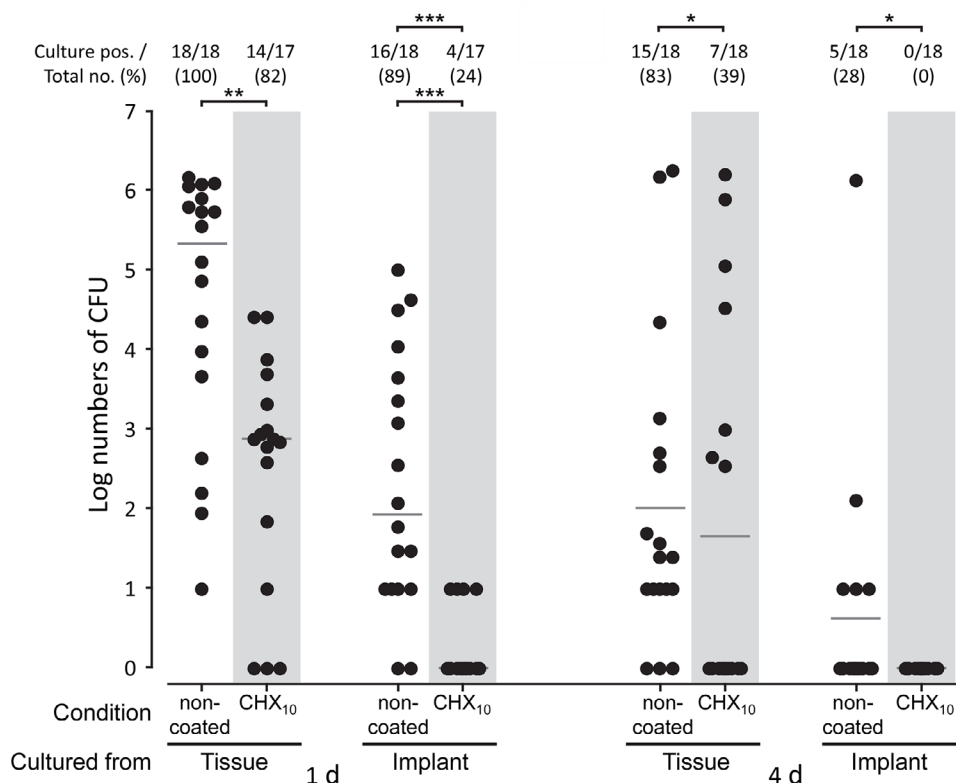
To assess the influence of the coatings on the local immune response, we simultaneously stained polymorphonuclear neutrophils (PMNs), macrophages and bacteria in single histological slides of tissue sections and visualised the cells using multispectral imaging. 1 d after challenge, a strong infiltration of PMNs as well as of macrophages around the non-coated titanium samples was observed (Fig. 7a). At this early time point, clearly less PMNs were present around the CHX<sub>10</sub>-coated implants. 4 d after challenge, the foreign body response had progressed from neutrophil to macrophage (F4/80 positive cells) predominance, around both the non-coated and the CHX<sub>10</sub>-coated titanium implants (Fig. 7c,d). So, in line with our macroscopic observations, there were no microscopic signs of *in vivo* adverse effects, such as excessively increased or protracted neutrophil-dominated inflammatory responses

nor necrosis, which might have been caused by the coating components or the chlorhexidine released.

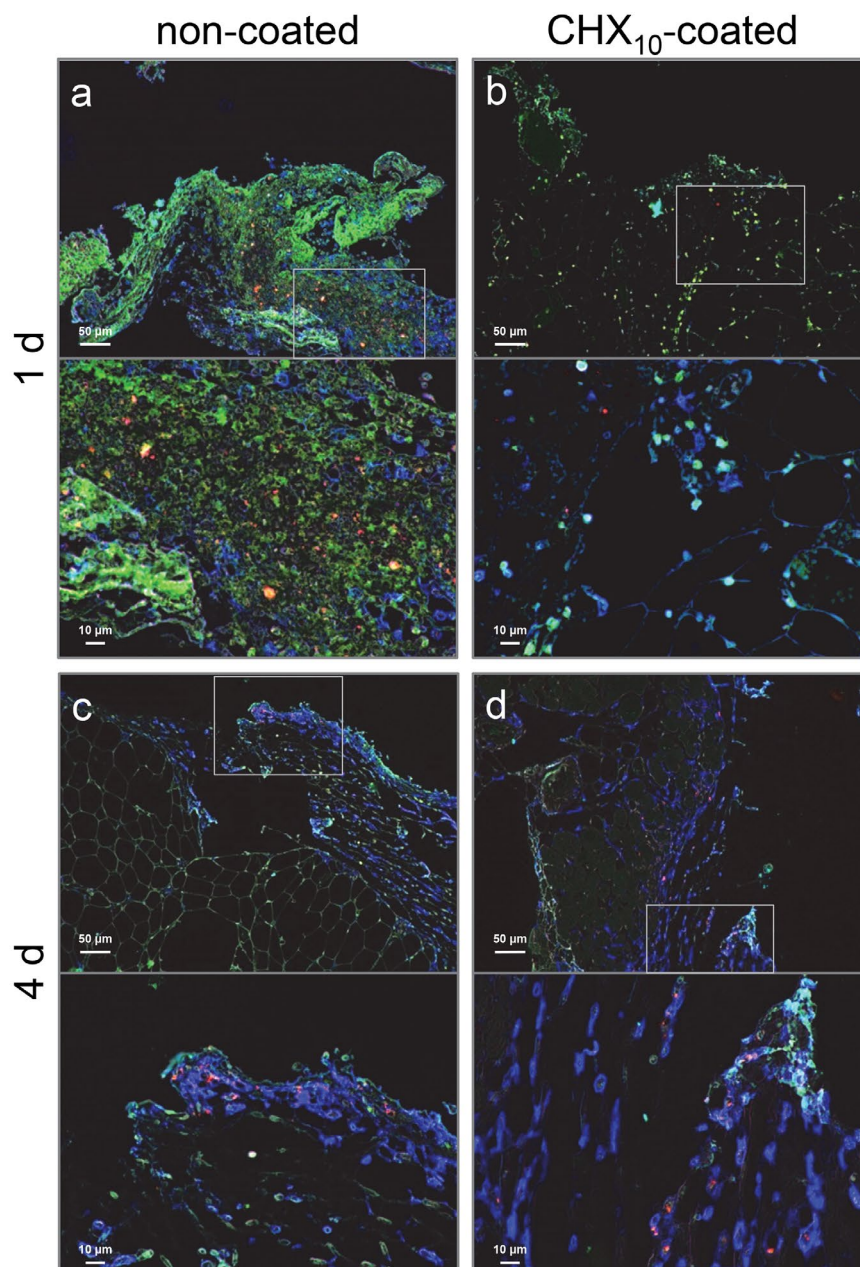
1 d after challenge, many bacteria were observed in the tissue around the non-coated implant (Fig. 7a) and only few bacteria around the CHX<sub>10</sub>-coated implant (Fig. 7b), which was in accordance with the quantitative culture results (Fig. 6). The bacteria were mainly co-localised with the PMNs. Although the number of culture-positive tissues around CHX<sub>10</sub>-coated implants 4 d after challenge was smaller compared to the tissues around non-coated implants, the number of CFUs retrieved was similar. This was also true for the number of bacteria observed by histology in the tissue surrounding the non-coated (Fig. 7c) and CHX<sub>10</sub>-coated implants (Fig. 7d). 4 d after challenge the bacteria were mainly co-localised with macrophages.

### Discussion

We have developed an epoxy-based mechanically stable coating for titanium implants providing a predetermined release of chlorhexidine to prevent biomaterial-associated infection (BAI). The epoxy-based coatings containing 5 or 10 wt % chlorhexidine, CHX<sub>5</sub> and CHX<sub>10</sub>, displayed an initial burst release in the first 24 h and the residual release occurring within 4 d. Additional *in vitro* studies showed that



**Fig. 6.** Bactericidal activity of chlorhexidine-releasing epoxy-based coating on titanium against  $1 \times 10^6$  CFU *S. aureus* GFP in mouse experimental biomaterial-associated infection 1 d (left) and 4 d (right) after implantation. Non-coated, titanium implant without coating; CHX<sub>10</sub>, titanium implant with coating containing 10 wt % chlorhexidine. Results are expressed as the frequencies and percentages of culture-positive samples and as the log of the numbers of CFU retrieved. The horizontal line represents the median log CFU. Frequencies of culture-positive samples and log numbers of CFU were analysed by Fisher's exact and Mann-Whitney tests, respectively. \*  $p \leq 0.05$ , \*\*  $p \leq 0.01$  and \*\*\*  $p \leq 0.001$ .



**Fig. 7.** Histological sections of biopsies of mice with non-coated titanium implants (non-coated (a,c) or with CHX<sub>10</sub>-coated implants (b,d), challenged with  $1 \times 10^6$  CFU of *S. aureus* GFP and sacrificed after 1 d (a,b) or 4 d (c,d). High magnification images, recorded from boxed area, are shown below the respective images. Macrophages immunostained with F4/80, neutrophils with Ly-6 and bacteria with anti-*S. aureus* antibodies are shown in multispectral images as blue, green and red, respectively. Co-localisation of macrophages and bacteria are shown in pink, co-localisation of macrophages and neutrophils in light-blue and co-localisation of bacteria and neutrophils in yellow. The images are representative for sections from different levels of the biopsies examined.

the coating completely eradicated the *S. aureus* inoculum even after washing for 24 h in PBS. The CHX<sub>10</sub> coating prevented mouse subcutaneous biomaterial-associated *S. aureus* infection and did not show macroscopic nor microscopic signs of adverse effects.

Coatings and materials derived from epoxy resins have found widespread use in numerous applications. Because of their good adhesion properties and excellent mechanical stability, epoxy-based coatings are highly suited for metallic orthopaedic implants, which are often inserted under mechanical stress. These coatings are particularly

suited for non-cemented implants, which in contrast to cemented implants cannot be protected by *e.g.* antibiotic releasing bone cement (Lynch *et al.*, 1987; Nijhof *et al.*, 2000) or other antimicrobial approaches, such as the use of bioactive glass (Drago *et al.*, 2013; Romano *et al.*, 2014), and are more prone to recurrent infection (Dale *et al.*, 2009; Neut *et al.*, 2015).

Perhaps the most well-known epoxy resin, imparting good mechanical properties to its derived coatings, is the diglycidyl ether of bisphenol-A (DGEBA) (Garcia *et al.*, 2009). Indeed, cured networks of DGEBA and



isophorone diamine (IPD) for biomedical applications showed respectable mechanical properties and good biocompatibility *in vitro* (Garcia *et al.*, 2009). However, bisphenol A derivatives are under continuous debate for their safety (Testai *et al.*, 2016). Therefore, we selected the diglycidyl ether glycerol as the epoxy resin, to be cured with IPD, a system shown to be haemocompatible (Garcia *et al.*, 2016) and not toxic in a rabbit bone implant model (Odekerken, 2015). Ensuring a nearly stoichiometric ratio of NH to epoxy groups resulted in a strongly crosslinked network. To further enhance adhesion of the coating to the titanium, dopamine was added to the formulation. The coating did not show any damage after extensive testing in the double rub as well as tape adhesion assays, illustrating its excellent mechanical properties. The coating therefore is a promising candidate for application to titanium implants in orthopaedics and dentistry.

Although antibiotic releasing coatings are widely used for different medical applications, *e.g.* in sutures and central venous and urinary tract catheters, antibiotic release coatings for orthopaedic devices remain mainly experimental. A study in rabbits with titanium fixation plates coated with poly-L-lactide (PLLA), containing a combination of rifampicin and fusidic acid, placed on the tibia, showed a significant reduction in *S. aureus* infection after 28 d (Källicke *et al.*, 2006). Direct application of rifampicin and minocycline on titanium significantly reduced *S. aureus* colonisation of femoral medullary nails and device-related osteomyelitis in rabbits after 1 week (Darouiche, 2007). In a rabbit tibia *S. aureus* infection model, the application of tobramycin to hydroxyapatite-coated titanium implants reduced infection rates and improved osseointegration after 28 d (Moojen *et al.*, 2009). In another rabbit study, the combination of hydroxyapatite with gentamicin showed significant reduction in *S. aureus* infection rates, good biocompatibility and bone integration as observed with pure hydroxyapatite (Alt *et al.*, 2011). A gentamicin-loaded biodegradable poly(D,L-lactide) (PDLLA) coating on titanium Kirschner wires showed a significant reduction in implant-related infection after 6 weeks of implantation in a rat tibia infection model (Lucke *et al.*, 2003). A recent prospective study of the first commercially available antibiotic-releasing intramedullary tibia nail, with a coating of PDLLA containing gentamicin, has shown promising results, with no deep surgical wound infections recorded within the first six months after implantation of the coated nails (Fuchs *et al.*, 2011). However, antibiotic-releasing coatings have two major disadvantages. Due to the worldwide rapid increase in antibiotic resistance, there is a risk that a patient will have an infection with a bacterium, which is resistant to the released antibiotic. Conversely, due to the local release, graded concentrations of antibiotic will be present near the implant, increasing the risk of selection of resistant bacteria and thus contributing to resistance development and ensuing implant infection.

In order to avoid antibiotics, we sought to use an antiseptic as the antimicrobial agent for our coating. One of the most frequently used antiseptics is chlorhexidine, with a broad spectrum of activity against Gram-positive and Gram-negative bacteria and yeasts, and with acceptable

tolerability and a good safety record (McDonnell and Russell, 1999). Chlorhexidine has been approved by the American Food and Drug Administration (FDA) for application to intracorporally used medical devices, such as surgical meshes (Choe *et al.*, 2000; Pérez-Köhler *et al.*, 2015) and intravenous catheters (Maki, 1997) and has a history of successful clinical (Karpanen *et al.*, 2016) and experimental use in preventing biomaterial-associated and other infections (Darouiche *et al.*, 2008; Sjollemma *et al.*, 2014). Chlorhexidine combined with chloroxyleneol in a dip-coating on rabbit tibia intramedullary nails prevented experimental *S. aureus* osteomyelitis (Darouiche *et al.*, 1998a; Darouiche *et al.*, 1998b).

Chlorhexidine is an important agent in the prevention of the spread of antibiotic resistant organisms and hospital-acquired infections (Johnson *et al.*, 2015). The effectiveness and widespread use of chlorhexidine has led to some concerns regarding the emergence of bacterial resistance (Edmiston *et al.*, 2013; Horner *et al.*, 2012), including MRSA resistance (Johnson *et al.*, 2015). However, reported levels of susceptibility of the bacterial isolates are still several orders of magnitude below the clinically applied dosages (Horner *et al.*, 2012).

We tailored our coatings to release most of the chlorhexidine within the first 24 h. As proof of concept for prevention of infection, we studied efficacy in our mouse subcutaneous implant infection model. The CHX<sub>10</sub> coating proved to be highly efficacious in preventing *S. aureus* infection and did not cause any signs of adverse effects. Interestingly, 1 d post challenge only few bacteria were observed in histology and there was only a relatively low level of neutrophil influx in the tissue surrounding the CHX<sub>10</sub>-coated implants. Likely, this small influx is due to the efficacy of the coating to kill *S. aureus* rapidly, thereby reducing *e.g.* local complement activation and consequent neutrophil attraction.

Studies in a rabbit tibia intramedullary nail infection model, with titanium implants with the same epoxy-based chlorhexidine-releasing coatings as the one used in our study, showed no signs of local nor of systemic toxicity during the 6 week experiment (Odekerken, 2015). Moreover, significantly greater bone apposition on the implant surface of both the CHX<sub>0</sub> (98.8 % coverage) and CHX<sub>10</sub> (81.3 % coverage) than on non-coated titanium nails (71.5 % coverage) was observed after 6 weeks. However, in contrast to our mouse results, no protection against *S. aureus* infection was observed. This may have been due to the prolonged time period of the rabbit experiments. In the first post-operative week, the rabbits showed reduced <sup>18</sup>F-FDG tracer uptake suggesting reduced levels of inflammation. This might indicate that a partial suppression of the *S. aureus* infection might have occurred in the first days of the experiment. Apparently, the chlorhexidine reservoir and its release profiles would need to be optimised for the rabbit model, which certainly is possible with this coating technology.

Bacteria colonising the surface of a biomaterial not only are a focus of a localised biofilm infection, but can also be the source of tissue colonisation (Riool *et al.*, 2014). Conversely, bacteria residing in the tissue can be a cause of infection after re-implantation, in experimental infection



(Engelsman *et al.*, 2010) as well as in patients. Tissue-residing bacteria can be hard to eradicate by antibiotic treatment (Broekhuizen *et al.*, 2008b; Nejadnik *et al.*, 2008) and may necessitate a prolonged regimen of systemic and local antibiotic treatment prior to re-implantation in patients (Zimmerli *et al.*, 2004). Thus, prevention of bacterial colonisation of tissue surrounding implants is of vital importance. In our study, large numbers of bacteria were cultured from the tissues around the non-coated implants. Particularly, 4 d after challenge, these bacteria were co-localised with macrophages, suggesting that the macrophages were not effectively killing the bacteria. This phenomenon has been observed in many earlier studies (Boelens *et al.*, 2000a; Boelens *et al.*, 2000b; Broekhuizen *et al.*, 2010). However, from the tissue around the CHX<sub>10</sub>-coated implants, only few bacteria were cultured and few bacteria were seen by histology. Apparently, the rapid initial release of chlorhexidine killed the vast majority of the infecting bacteria, preventing formation of a biofilm on the implant surface as well as colonisation of the tissue, thus protecting both these sites against infection.

### Conclusions

Taken together, our mechanically stable chlorhexidine-releasing epoxy-based coating on titanium holds promise for further development towards clinical application. Because of its mechanical stability, it is suited for all types of bone implants, including but not limited to non-cemented orthopaedic implants and possibly dental implants as well. Our coating provides a good alternative for coatings releasing conventional antibiotics and therefore has the potential not only of preventing implant infection but also contributing to reduction of antibiotic resistance development.

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## Discussion with Reviewers

**David Grainger:** Do the authors envision this non-degrading coating to be used in limited, designated places on the implant, well-away from the cement-less bone on-growth areas? How is this rationalised if infection often

occurs where bone on-growth is also supposed to occur?

**Authors:** Initial studies have indicated that the coating does not necessarily inhibit – or even promotes – bone apposition and might therefore also be used to prevent infection of bone on-growth areas of the implant.

**David Grainger:** Would be a possibility to use this approach with cemented implants, where antibiotic loaded cements might synergies chlorhexidine release? Often antibiotics are combined with antiseptics as the authors cite for catheters (rifampin/chlorhexidine), silver/minocycline, *etc.* So is this not appropriate to be used with antibiotic loaded cements?

**Authors:** This certainly is a possibility but it requires further investigation regarding the optimal combination of chlorhexidine and antibiotics. The combination of antiseptics and antibiotics is not that common though. The references cited have predominantly studied the use of either combinations of two antiseptics (*e.g.* chlorhexidine and silver-sulfadiazine; Maki, 1997) or of two antibiotics (*e.g.* rifampicin and minocycline; Darouiche, 2007).

**Yannis Missirlis:** What is the shear rate experience by the coating/substrate interface, when the artificial hip is forcefully inserted in place?

**Authors:** We are not aware of the exact shear rate in this particular case, as the present study was the first step in the development of a novel mechanically stable antimicrobial-releasing coating. Indeed, insertion of a prosthetic hip may be associated with greater shear rates, with possible consequences for any coating applied on the implant. Further research should focus on the possible applications of this coating.

**Yannis Missirlis:** Is fatigue shear to be experienced at the interface during everyday use of the implant?

**Authors:** Depending on the application, fatigue shear might be experienced at the interface (also see question above). Whether this would compromise the functionality of the implant in the long term requires further long term research in relevant animal models.

**Yannis Missirlis:** How much more does dopamine facilitate adhesion to Ti surfaces? Does the Ti surface have to be prepared for this purpose?

**Authors:** Dopamine facilitates adhesion because of its catechol moiety, being able to attach to the surface oxide layer of the titanium substrate (Ye *et al.*, 2011). Apart from cleaning, the surfaces did not need any preparation to attain adhesion. However the increase in adhesion due to dopamine was not quantified during the coating design phase. Epoxy/amine coatings without dopamine (or an appropriate primer layer) detached from the metal substrates upon submersion in water.

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**Editor's note:** The Scientific Editor responsible for this paper was Thomas Fintan Moriarty.