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Aqueous chemoenzymatic one-pot enantioselective synthesis of tertiary α-aryl cycloketones via Pd-catalyzed C–C formation and enzymatic C–C asymmetric hydrogenation†

Pengqian Luan,a Yunting Liu,*a,b Yongxing Li,a Ran Chen,a Chen Huang,a Jing Gao,a Frank Hollmann c and Yanjun Jiang*a

An aqueous chemoenzymatic cascade reaction combining Pd-catalyzed C–C formation and enzymatic C–C asymmetric hydrogenation (AH) was developed for enantioselective synthesis of tertiary α-aryl cycloketones in good yields and excellent enantioselectivities. The stereopreference of the enzyme in AH of α-aryl cyclohexenones was studied. An enantiocomplementary enzyme was obtained by site-directed mutation.

Biocatalysts have become one of the fundamental pillars in the toolbox of synthetic chemistry due to their high stereoselectivity. More importantly, biocatalysis can produce enantiomer-enriched molecules in a green way based on the mild reaction conditions and the use of water as the preferred solvent, which meets the requirements of environmentally friendly catalytic processes and sustainable development strategy.1 Although the catalytic repertoire of enzymes is striking, many valuable transformations catalyzed by artificial catalysts until now have no known enzyme-catalysed counterparts,2 and the catalytic capabilities of enzyme towards non-natural substrates are limited.3 The chemoenzymatic catalysis has proven to be effective to tackle these limitations, where the wide catalytic capabilities of chemocatalysts and the exquisite selective properties of biocatalysts are usually merged in a one-pot aqueous system.4 The pioneering work on combining Pd-catalyzed C–C bond formation with the enzymatic carbonyl reduction developed by Gröger,5a provides a paradigm for aqueous chemoenzymatic cascade catalysis, which is becoming a popular trend in asymmetric synthesis.5 However, this strategy often suffers from the poor stability of the free enzymes and the incompatibilities between the two catalytic disciplines,6 motivating the development of stable and compatible systems to realize aqueous chemoenzymatic processes.

Chiral tertiary α-aryl cycloketones are prevalent structural motifs in pharmaceuticals, bioactive molecules and natural products.7 The enantioselective synthesis of such acidic α-H-containing compounds has always been a challenging task due to their prone-to-racemization nature under basic conditions.8 Metal-catalyzed asymmetric α-arylation of preformed soft enolates with low basicity carbonyls,9 and enantioselective protonation of achiral enolate complexes using Lewis-acid assisted chiral Brønsted acid catalysts,10 have proven to be a powerful tool to deliver tertiary α-aryl cyclohexanones (Scheme 1a and b). Shi et al. achieved the asymmetric synthesis of more challenging tertiary α-aryl cyclopentanones by enantioselective

Scheme 1 Strategies for asymmetric synthesis of chiral tertiary α-substituted cycloketones.
epoxidation of benzylidene cyclobutane and subsequent epoxide rearrangement (Scheme 1c).\textsuperscript{12} Albeit effective, none of these approaches enable the direct use of ketone substrates. The asymmetric hydrogenation (AH) of readily available prochiral enones catalyzed by transition metals or organocatalysts, is one of the straightforward and simple strategies for access to chiral ketones.\textsuperscript{12} Although great progress has been made in AH of acyclic and exocyclic enones,\textsuperscript{13,14} only few methodologies have been developed for endocyclic enones,\textsuperscript{15} and no reports on AH of α-aryl endocyclic enones. In addition, for most of the above strategies, toxic organic solvents and expensive chiral ligands are inevitably used, deviating from the principles of Green Chemistry. Therefore, it is still highly urgent to develop a facile, general and green method for enantioselective synthesis of chiral tertiary α-aryl cycloketones.

Old yellow enzymes (OYEs) are an important class of ene-reductases (ERs), which possess the ability to generate up to two chiral centers via the stereospecific trans-hydrogenation (hydride addition/protonation) of conjugated C=C bonds.\textsuperscript{16} OYEs-catalyzed AH has emerged as a valuable synthetic approach, which enabled various chiral compounds to be reduced in mild reaction conditions with high enantioselectivities, such as α-alkyl endocyclic enones.\textsuperscript{17} However, the ability of OYEs in AH of α-aryl endocyclic enones and the integration of OYEs with other catalytic disciplines have been rarely investigated.\textsuperscript{18} Herein, we developed an aqueous chemoenzymatic two-step one-pot cascade reaction for the direct synthesis of enantioenriched tertiary α-aryl cycloketones from readily available 2-iodocycloenones. In this process, the aryl groups were installed through Suzuki-Miyaura coupling (SMC) catalyzed by an immobilized Pd catalyst (DON@Pd), and subsequently, the α-H-containing stereocenters were constructed by OYE-catalyzed AH of α-aryl endocyclic enones (Scheme 1d).

To test the feasibility of this strategy, 2-phenylcyclohexenone (1a) synthesized by SMC reaction of 2-iodocyclohexenone and phenylboronic acid,\textsuperscript{19} was selected as a model substrate. For the AH reaction, we evaluated three different OYEs (TsOYE from\textit{ Thermus scotoductus}, OYE1 from\textit{ Saccharomyces pastorianus} and YqJ from\textit{ Bacillus subtilis}) in form of freeze-dried enzyme solution (pH 7.5) using DMF (v/v 10%) as cosolvent to overcome the poor substrate solubility in water. Unfortunately, 1a was not transformed by TsOYE and OYE1, only YqJ could produce (R)-2-phenylcyclohexanone (2a) in extremely low yield (9%), albeit with high stereoselectivity (Table 1, entries 1–3). The R configuration of 2a was determined by optical rotation compared with the literature data.\textsuperscript{20} Although the yield and the ee value were further increased when the reaction was conducted under diluted conditions, it was still far from satisfactory (Table 1, entries 4 and 5). Subsequent experiments revealed a significant cosolvent effect on the catalytic performance of YqJ. Water-immiscible DME and acetone led to no conversion (Table 1, entries 6 and 7), DMSO and THF caused decreases in the enantioselectivity of the AH (Table 1, entries 8 and 9). Water-immiscible isooctane and ionic liquids (ILs) significantly improved the yield and enantioselectivity of the reaction (Table 1, entries 11–13). The best result was obtained with [BMIm][NTf2] (20% v/v) as cosolvent (Table 1, entry 14), which was attributed to its better biocompatibility and extraction ability.\textsuperscript{21} In this biphasic system, the substrate was partitioned into the IL phase, which served as substrate reservoir and product sink,\textsuperscript{22} thereby preventing the interaction between the substrate/cosolvent and the YqJ. However, a further increase to 1 : 1 (v/v) [BMIm][NTf2] proved to be harmful to YqJ (Table 1, entry 15). The effect of cosolvents on relative enzyme activity was also evaluated, and the results were summarized as shown in Fig. S1.† The water-immiscible solvents (DMF, DME, DMSO, THF and acetone) had a severely deleterious effect on enzyme activity, while the water-immiscible solvents (n-hexane, isooctane and ILs) were more biocompatible, especially [BMIm][NTf2], which was consistent with the reaction results.

According to literature,\textsuperscript{17,23} the type of cofactor and recycling system can cause remarkable variation of yields and stereoselectivities. This effect was also examined in our reactions (Table 1, entries 16–18). Using NADPH gave better results than using NADH. With catalytic amounts of NADPH, the recycling system of glucose dehydrogenase (GDH)/glucose (Glu) pair achieved 79% yield and 97% ee (Table 1, entry 18). Furthermore, recombinant cells co-expressing the enzymes of YqJ and GDH (YqJ-RBS-GDH) were constructed to avoid

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**Table 1** Optimization studies of OYEs-catalyzed AH of 2-phenylcyclohexenone\textsuperscript{a}

<table>
<thead>
<tr>
<th>Entry</th>
<th>Cosolvents</th>
<th>Enzymes</th>
<th>Yield\textsuperscript{b} (%)</th>
<th>ee\textsuperscript{b} (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1\textsuperscript{c}</td>
<td>DMF (10%)</td>
<td>OYE1</td>
<td>n.d.</td>
<td>—</td>
</tr>
<tr>
<td>2</td>
<td>DMF (10%)</td>
<td>TsOYE</td>
<td>n.d.</td>
<td>—</td>
</tr>
<tr>
<td>3</td>
<td>DMF (10%)</td>
<td>YqJ</td>
<td>9</td>
<td>85</td>
</tr>
<tr>
<td>4\textsuperscript{f}</td>
<td>DMF (10%)</td>
<td>YqJ</td>
<td>26</td>
<td>85</td>
</tr>
<tr>
<td>5\textsuperscript{f}</td>
<td>DMF (10%)</td>
<td>YqJ</td>
<td>42</td>
<td>87</td>
</tr>
<tr>
<td>6</td>
<td>DME (10%)</td>
<td>YqJ</td>
<td>n.d.</td>
<td>—</td>
</tr>
<tr>
<td>7</td>
<td>Acetone (10%)</td>
<td>YqJ</td>
<td>42</td>
<td>60</td>
</tr>
<tr>
<td>8</td>
<td>DMSO (10%)</td>
<td>YqJ</td>
<td>39</td>
<td>59</td>
</tr>
<tr>
<td>9</td>
<td>THF (10%)</td>
<td>YqJ</td>
<td>43</td>
<td>92</td>
</tr>
<tr>
<td>10</td>
<td>n-Hexane (10%)</td>
<td>YqJ</td>
<td>58</td>
<td>94</td>
</tr>
<tr>
<td>11</td>
<td>Isooctane (10%)</td>
<td>YqJ</td>
<td>64</td>
<td>95</td>
</tr>
<tr>
<td>12</td>
<td>[BMIm][PF6] (10%)</td>
<td>YqJ</td>
<td>70</td>
<td>95</td>
</tr>
<tr>
<td>13</td>
<td>[BMIm][NTf2] (10%)</td>
<td>YqJ</td>
<td>76</td>
<td>95</td>
</tr>
<tr>
<td>14</td>
<td>[BMIm][NTf2] (20%)</td>
<td>YqJ</td>
<td>32</td>
<td>67</td>
</tr>
<tr>
<td>15</td>
<td>[BMIm][NTf2] (50%)</td>
<td>YqJ</td>
<td>54</td>
<td>90</td>
</tr>
<tr>
<td>16\textsuperscript{f}</td>
<td>[BMIm][NTf2] (20%)</td>
<td>YqJ + FDH</td>
<td>43</td>
<td>82</td>
</tr>
<tr>
<td>17</td>
<td>[BMIm][NTf2] (20%)</td>
<td>YqJ + GDH</td>
<td>79</td>
<td>97</td>
</tr>
<tr>
<td>18</td>
<td>[BMIm][NTf2] (20%)</td>
<td>YqJ-RBS-GDH</td>
<td>89</td>
<td>99</td>
</tr>
<tr>
<td>19</td>
<td>[BMIm][NTf2] (20%)</td>
<td>YqJ-RBS-GDH</td>
<td>86</td>
<td>99</td>
</tr>
</tbody>
</table>

\textsuperscript{a} Reactions were carried out at 30 °C in Tris-HCl buffer (pH 7.5) using 1a (5 mM), OYE whole-cell lyophilizates (0.2 g), NADPH as hydrogen source. \textsuperscript{b} The yields and the values of ee were determined by HPLC. \textsuperscript{c} Using 1a (2 mM). \textsuperscript{d} Using 1a (1 mM). \textsuperscript{e} NADH as hydrogen source. \textsuperscript{f} Using 1a (25 mM). n.d. = not determined.
adding extra GDH (Fig. S2†), and they produced 2a in a higher yield and stereoselectivity (Table 1, entry 19). This result was mainly ascribed to that the two enzymatic transformations occurred within the cell, which enhanced the cofactor regeneration efficiency.21 Under the optimal solvent system, the substrate concentration could increase to 25 mM with only a slight decrease in yield (Table 1, entry 20).

The integration of SMC and enzymatic hydrogenation in the same reaction medium was explored to realize the two-step one-pot chemoenzymatic cascade synthesis of enantioenriched tertiary α-aryl cycloketones. Preliminary experiments revealed that the direct combination of the two steps caused incomplete bioreduction (only 41% overall yield), which was ascribed to the inhibition of enzyme by the residual boronic acid.26 Because the heterogeneous Pd/C could not convert boronic acid completely, especially in the case of using recycled catalyst. While the homogeneous catalysts, such as Pd[PPh3]4 and Pd(OAc)2/PPh3, could make the coupling reaction proceed more thoroughly, the metal cation and phosphine ligand were also inhibitors of the enzyme.24 To solve this problem, an immobilized-Pd catalyst was fabricated using our previously reported dendritic organosilica nanoparticles as carrier (DON@Pd, the particle size and the loading amount of Pd were calculated to be 1.8 nm and 5 wt%, respectively) (Fig. S3–S5†). The wrinkle structure with high specific surface area is conductive to the formation of ultra-small and high dispersed Pd nanoparticles. After a simple optimization of the reaction conditions (Table S1†), the obtained DON@Pd produced 1a in almost quantitative yield. Fortunately, its combination with YqjM in a biphasic system where Pd-catalyzed C–C formation was performed in the IL phase, and enzymatic C=C AH was performed in the buffer phase, could produce the final product 2a in 81% yield and 99% ee. It should be mentioned that the yield of cascade synthesis was higher than that of stepwise synthesis (63%).

After the optimized reaction protocol for the chemoenzymatic asymmetric synthesis of tertiary α-aryl cyclohexanones was established, the substrate scope of the reaction was surveyed. As indicated in Scheme 2a, substrates containing electron-donating and electron-withdrawing substituents were converted into the corresponding target products (2α–k) in moderate to high yields (40–81%) and excellent enantioselectivities (91–99% ee). The ortho-substituents of the aryl groups caused much slower reaction rates and lower yields due to the increased steric bulk (2b, 2g and 2l). In addition, the five- and seven-membered ring substrates were also investigated. The tertiary α-cyclopentanones (2f–o) were also obtained in good yield (71–78%) and excellent enantioselectivities (96–99% ee). However, α-phenyl cycloheptenone (2p) could not be converted, which may be due to that the substrate specificity of YqjM is generally limited to small molecules.26 The enzymatic AH of α-alkyl enones in this biphasic system was also evaluated (3a–d), achieving improved yields and increased substrate concentrations compared to the previous reported results (Scheme 2b and Table S2†).

As shown in previous reports,27 the stereoselectivity of the YqjM strongly depends on the substrate and is hardly predictable. We also observed a striking switch of stereopreference in the reduction of α-phenyl and α-methyl cyclohexenones (2a and 3a). In order to rationalize this phenomenon, induced docking experiments were performed based on the crystal structure of YqjM. In the process of YqjM-catalyzed hydrogenation, H-bonding formed between His167 and C=O bond of the substrate is essential for the activation of C=C bond (Fig. S6–S8†). Ala-60 and Ile-69 build up the entrance of the active site cavity forming a deep and small pocket on the bottom, which can accommodate the methyl group of 3a, but cannot accept the sterically hindered phenyl ring of 2a (Fig. S6 and S7†), thus forcing 2a to turn around and to bind in the opposite orientation. In addition, the reorientation by 180° of 2a in the active site resulted in a strong π–π interaction between the phenyl ring of 2a and the imidazole ring of His167 (Fig. S7†), in which the distance between the two planes is about 3.3–3.7 Å (Fig. S9†). The effective H-bond and π–π interactions bring the substrate closer to the active center of the enzyme, which is responsible for the high level of enantio-induction.

The access to enantiocomplementary products is highly valuable for manufacturing drugs since the two mirror-image forms of a chiral molecule usually exhibit different pharmacological activities. Site-directed mutation was performed to develop an enantiocomplementary variant of YqjM for obtaining the opposite enantiomer products. We envisioned that widening the active pocket to accommodate the sterically hindered phenyl group might be a viable solution. After careful screening of the amino acids around the active center, a

![Scheme 2](image-url)
double amino acid mutation (Gly-60 and Gly-69 were installed in place of Ala-60 and Ile-69, respectively) was found to induce complete reversal of the enantioslectivity of YqJm (Fig. S8†). After a slight optimization of the reaction conditions, (S)-2a was produced by the mutant YqJm through the above developed chemoenzymatic process with 42% yield and 99% ee (Scheme 3a).

To expand the applicability of the chemoenzymatic protocol, a three-step one-pot cascade reaction was developed for asymmetric synthesis of chiral α-aryl cycloalkanols via the combination of this process with AH of C≡O bond catalyzed by alcohol dehydrogenase from *Rhodococcus ruber* (ADH-A).29 Without any purification of intermediates, enantioenriched (1S,2R)-2-phenylcyclohexanol-1-ol (4) was directly produced from 2-iodocyclohexenone in 62% yield and 98% ee (Scheme 3b).

Finally, the recyclability of the both types of catalysts was investigated by the model reaction of chemoenzymatic production of (R)-2a under the optimal conditions. The DON@Pd could be recovered by centrifugal separation before enzymatic AH, and its high catalytic performance was completely sustained for 5 cycles with no significant leaching and aggregation of Pd NPs (Fig. S10 and S11†). However, the recovery of the catalytically active cells was difficult due to significant cell lysis observed under these reaction conditions. To solve this problem, the two enzymes [YqJm and GDH] were purified and co-immobilized on DONs (for details see ESI†). Under optimized immobilization conditions, the maximum enzyme loading was 230 mg g<sup>-1</sup> (Fig. S12†). Compared with the whole cells, the co-immobilized catalyst exhibited obviously higher stabilities to organic solvents, temperatures and pH, as well as higher storage stability (Fig. S13–S16†). While the whole cells were almost inactivated after recycled 5 times (Fig. S17†), the co-immobilized enzymes used 5 times still maintained a moderate activity, producing (R)-2a in 52% yield and 93% ee (Fig. S18†).

**Conclusions**

In conclusion, an aqueous chemoenzymatic approach for asymmetric synthesis of enantioenriched tertiary α-aryl cycloketones has been developed for the first time. The transformation relies on the sequential cascade reaction combining Pd-catalyzed SMC and YqJm-catalyzed AH of α-aryl endocyclic enones. This process can be expanded to synthesize chiral α-aryl cycloalkanols bearing two contiguous stereocenters by combining ADH-catalyzed AH of C≡O bond. An enantiocomplementary variant of the YqJm was also obtained by site-directed mutation. This concept, which integrates the versatile reactivity of chemocatalysis and the exquisite selectivity of biocatalysis, is a green, reliable alternative and supplement to the existing approaches for the asymmetric synthesis of chiral molecules.

**Conflicts of interest**

There are no conflicts to declare.

**Acknowledgements**

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**Notes and references**


The R configuration of 2a was determined by optical rotation compared with the literature data. The optical rotation of the product 2a is [α]D22 20 = +108.9° (c 0.45, CHCl3, 99% ee) [Lit. [α]D22 20 = +102° (c 0.45, CHCl3, 93% ee)], see: C. Cheon, O. Kanno and F. Toste, J. Am. Chem. Soc., 2011, 133, 13248–13251.


